

# Chemistry and Pharmacology of Flavonoids- A Review

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## ABSTRACT

Several modern and most of the traditional drugs have been developed from natural sources. Flavonoids or bioflavonoids, most ubiquitous polyphenolic compounds, are secondary metabolites of plants and fungal origin. Apart from their biological functions in plants (protection against herbivores, ultraviolet radiation and pathogens), they perform myriads of pharmacological activities in humans as well. Though flavonoids are not acknowledged as nutrients still their intake on regular basis is considered fruitful for human health. Flavonoids are biosynthesized through phenylpropanoid pathway and contain a C6-C3-C6 carbon framework. The present review has reviewed the chemistry, structure and classification of flavonoids. Additionally, their occurrence and chemical properties have also been explored. Moreover, we discuss about the different mechanisms through which flavonoids act like direct radical scavenging, leukocyte immobilization and interaction with different enzymes. Flavonoids own a number of pharmacological activities such as anti-parkinson, anti-ulcer, spasmolytic, anti-depressant, anti-bacterial, anti-hypertensive, anti-diabetic, anti-inflammatory and anti-cancer. This review intent to give healthy information for formation of new flavonoid based pharmaceutical formulation to act against various diseases.

**Key words:** Flavonoids, Polyphenolic compound, Pharmacological activity, Mechanism, Biosynthesis.

## INTRODUCTION

'Flavonoid' word was initially derived from 'flavous', a latin word which means yellow, resembling flavonoid's colour in nature.<sup>1-3</sup> Despite of its meaning, plentiful of other flavonoids are white and the chief flavonoid-related anthocyanins are purple, red or blue in colour as well.<sup>3</sup> Flavonoids or bioflavonoids, are a category of secondary metabolites of plants and fungal origin.<sup>4-9</sup> They are a class of natural compounds having variable phenolic structures.<sup>6</sup> A new substance was screened out from oranges in 1930 which was thought to be a member of a new class of vitamins and was designated as vitamin P.<sup>7,10,11</sup> Later on, that substance was confirmed to be a flavonoid, called rutin.<sup>11</sup> The term "flavonoid" is basically used to describe a broad assemblage of nat-

ural compounds that contain a C6-C3-C6 carbon framework or more accurately a phenylbenzopyran functionality.<sup>12</sup> The position at which the aromatic ring links with the benzopyrano functionality helps to determine the three classes into which this group of natural compounds can be divided: the flavonoids (2-phenylbenzopyrans) Figure 1, isoflavonoids (3-phenylbenzopyrans) Figure 2 and the neoflavonoids (4-phenylbenzopyrans) Figure 3 and (Chalcone) Figure 4. These groups generally share a common precursor (chalcone) and are therefore structurally and biogenetically related.<sup>13-17</sup> Flavonoids possess different pharmacological activities and act through several mechanisms. All the healthy information regarding flavonoids will be discussed in this paper.

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## Chemistry and Structure

Flavonoids are naturally occurring compounds present in plants. They have variable phenolic structures. Stereochemically flavonoids are composed of a 15-Carbon skeleton comprising of two benzene rings (A and B as shown in Figure 1) which are linked through a heterocyclic pyrane ring (C). Flavonoids have been classified into a number of classes. Biosynthesis of flavonoids occurs via phenylpropanoid pathway. In this pathway phenylalanine which is an amino acid, gets transformed into 4-coumaroyl-CoA and this 4-coumaroyl-CoA conjugates with malonyl-CoA to give chalcones consisting of two phenyl rings. Conjugate ring-closure of chalcones produces a three-ringed similar form of flavonoids called as flavone. This pathway continues via a sequence of various enzymatic modifications to form flavanones, dihydroflavonols and anthocyanins. Along with these compounds flavan-3-ols, proanthocyanidins (tannins), flavonols and several other poly-phenolics can also be formed.<sup>19</sup> Flavonoid's basic structure is aglycone (Figure 1).<sup>20</sup> In the structure of flavonoids, a six-member ring that is condensed with the benzene ring can either be a  $\alpha$ -pyrone (flavanones and flavonols) or its dihydro-derivative (flavanones and flavonols).<sup>20,22-26</sup> Flavanones differ from flavonols by lacking a hydroxyl group (OH) at the 3- position and a C2-C3 double bond.<sup>27,28</sup> Different class of Flavonoids are frequently hydroxylated at different positions (2,3,3',4',5,5' and 7). The carbohydrates (D-glucose, L-rhamnose, glucorhamnose, galactose or arabinose) are formed via glycosidic linkage generally positioned at positions 3 or 7.<sup>6,29</sup>

## Occurrence and Properties of Flavonoids

Flavonoids are composed of a wide-range of polyphenolic entities having a benzo- $\gamma$ -pyrone system and are pervasive in plants.<sup>30-31</sup> Because flavonoids are secondary metabolites (biosynthesized through shikimic acid pathway) of plants, they are consumed by man via food too.<sup>30,32</sup> Flavonoids are polyhydroxyphenols which are synthesized by plants to act against microbial infec-

tion.<sup>22,33</sup> The major dietary sources of flavonoids within the reach of human beings are vegetables and fruits including tea and wine.<sup>20,34</sup> Greater than 8000 compounds possessing basic flavonoid structure have been found, out of which several compounds are responsible for magnificent colors of various parts of plants (for e.g.-leaves, flowers and fruits). These secondary metabolites provide protection to plants against herbivores, ultraviolet radiation and pathogens (bacteria, virus and other microorganisms).<sup>35,36</sup> Various reports related to different substances of plant origin have stated the impact of flavonoids on metabolism of thyroid hormone, chemically distinct from ascorbic acid which has been assigned as vitamin P and has been considered useful in hemorrhage.<sup>10</sup> Flavonoids, in common, are hydrophilic substances having low values of log P and low molecular weights. It is considered that transport of flavonoids across cell membrane should take place through particular transmembrane transporters instead of simple diffusion through lipid part of the cell membrane.<sup>37</sup> Flavonoids act as functional foods by promoting health and preventing diseases.<sup>35,38</sup> Regardless of such useful properties of flavonoids, one major con is their low bioavailability which can differ between individual flavonoids in a specific class and between different classes as well.<sup>38</sup> Variability in bioavailability of different flavonoids can be detected from the data of comparative urinary excretion of daidzin and anthocyanins intake which was 43% and 0.3% respectively.<sup>38,39</sup> The bioavailability of flavonoids with complex higher structures and high molecular weights can even be lower.<sup>38,40</sup> Classification of flavonoids is presented in Table 1.

## Working Mechanisms

### Antioxidative Effects

The most illustrated property of almost every class of flavonoids is their ability to function as antioxidants. All the body cells and tissues are always vulnerable to

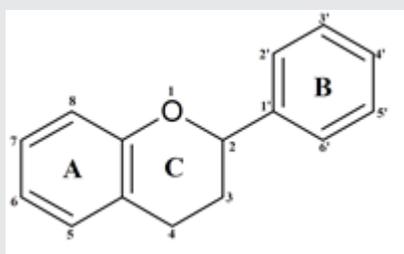


Figure 1: Basic structure of flavonoids.

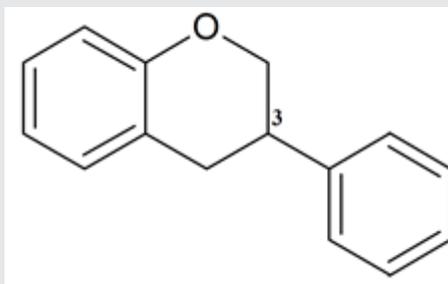
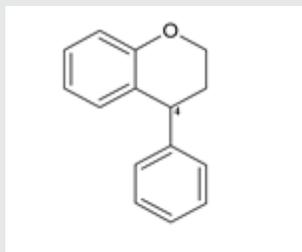
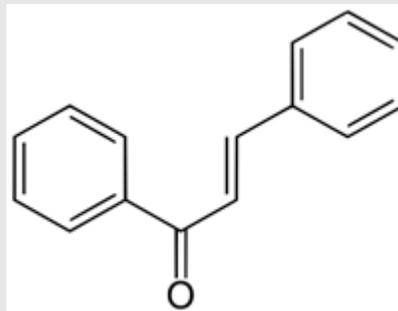


Figure 2: Basic structure of isoflavonoids.



**Figure 3: Basic structure of neoflavonoid.**

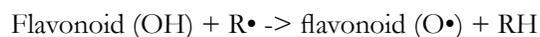


**Figure 4: Basic structure of chalcone.**

the devastating effect of reactive oxygen species and free radicals, which are formed during normal oxygen metabolism or are induced by damage due to exogenous factors.<sup>41,42</sup> The mechanisms and the procedure via which free radicals interrupt normal cellular functions are not entirely known. However lipid peroxidation has been assumed to be one of the most important mechanisms through which free radicals act. Lipid peroxidation generally causes damage to cell membranes. This damage results in shifting of the cell's net charge which causes a change in cell's osmotic pressure and this leads to swelling and ultimately cell death. Another way by which free radicals cause tissue damage is by causing general inflammatory response by attracting inflammatory mediators.<sup>43,44</sup> Human body's antioxidant defence mechanism include both enzymatic (glutathione peroxidase, superoxide dismutase and catalase) and non-enzymatic (ascorbic acid,  $\alpha$ -tocopherol and glutathione) parts. A hike in the formation of ROS during damage or injury results in utilization and reduction of the cell's scavenging compounds. At single point of time, flavonoids can interrupt more than three different ROS generating systems that are explained below. Flavonoids may upregulate the effect of endogenous antioxidants as well.<sup>41,44</sup>

#### **Direct Radical Scavenging Action**

Damage due to free radicals can be prevented by flavonoids in various ways. Out of them one is the direct scavenging of radicals. As per the mechanism, radicals oxidize the flavonoids and themselves get reduced. The outcome of this is that the reduced radicals are now in a less-reactive and more stable form. Flavonoids react with reactive part of the free radical to stabilize it.<sup>41</sup> As hydroxyl group of flavonoids is highly reactive, the radicals are stabilized according to the mentioned equation:<sup>45,46</sup>



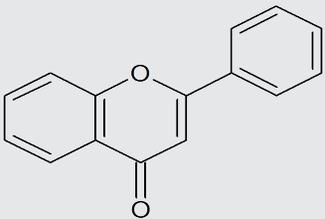
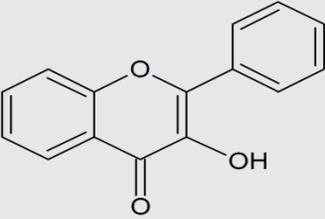
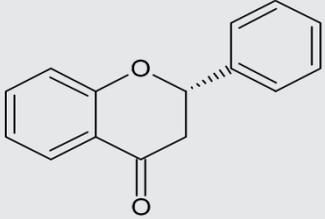
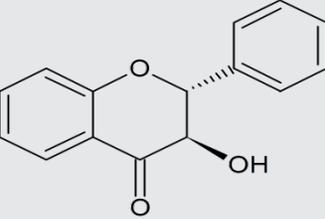
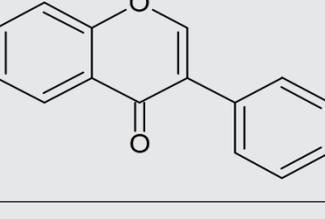
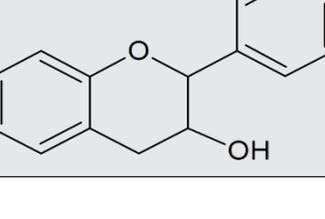
Where  $\text{R}\cdot$  symbolizes a free radical and  $\text{O}\cdot$  symbolizes an oxygen free radical. Some flavonoids inhibit superoxides while other flavonoids inhibit highly reactive peroxy nitrite, which is an oxygen-derived radical. Rutin and epicatechin both are extremely strong radical scavengers.<sup>47</sup> Rutin inhibits xanthine oxidase enzyme and this ability of rutin helps it to act as a powerful scavenger. *In vitro* LDL oxidation can be inhibited by flavonoids via their radical scavenging property.<sup>48</sup> This effect of flavonoids on LDL oxidation helps to protect LDL particles and may be used as a preventive measure against atherosclerosis.<sup>41,45</sup>

#### **Nitric Oxides**

Occurrence of ischemia-reperfusion injury has been lowered by quercetin and various other flavonoids which act via interfering with the action of inducible nitric oxide synthase.<sup>49,50</sup> Nitric oxide is produced by various cell of the body including macrophages and endothelial cells. Dilation of blood vessels in the body is maintained by nitric oxide which is released by the action of nitric oxide synthase<sup>51</sup> but at the same time release of nitric oxide in higher amount in macrophages results in oxidative damage. Nitric oxide reacts with free radicals to form extremely devastating peroxy nitrite which causes irreversible damage to the cell membrane by directly oxidizing the low-density lipoproteins.<sup>52</sup> Flavonoids act as antioxidants by scavenging the free radicals. Less damage to the body will be caused as free radicals will no more be available to react with nitric oxide.<sup>53-55</sup> Several studies have reported that flavonoids can scavenge nitric oxide molecules directly and this makes nitric oxide to be considered as a radical.<sup>56-58</sup> Thus it has been evaluated that scavenging of nitric oxide perform a crucial role in determining the therapeutic potential of different flavonoids.<sup>58</sup> According to a report nitric oxide has been dose dependently inhibited by a flavonoid named Silibin.<sup>59</sup>

#### **Xanthine Oxidase Pathway**

Table 1: Classification of Flavonoids<sup>6,18</sup>.

Class of flavonoid	Structural backbone	Examples
Flavones		Luteolin, Apigenin, Chrysin
Flavonols		Quercetin, Kaempferol, Galangin, Rutin, Myricetin
Flavanones		Hesperidin, Naringenin
Flavanonol		Aromadedin, Taxifolin
Isoflavones		Genistein, Daidzein, Glycitein, Formononetin
Flavan-3-ols		Catechin, Galliccatechin, Epicatechin

After ischemic reperfusion, xanthine oxidase pathway has been considered as a vital route through which oxidative injury is caused to different cells and tissues.<sup>60,61</sup> Xanthine gets metabolised to uric acid via involvement of both enzymes xanthine oxidase and xanthine dehydrogenase. Under normal physiological conditions xanthine dehydrogenase is present in the form of an enzyme which gets interconverted to xanthine oxidase (source of free radicals) during oxidative stress. In the reoxygenation phase superoxide free radicals are released as a result of reaction of xanthine oxidase with available molecular oxygen. Flavonoids, quercetin and silibin have been proved to decrease oxidative injury by inhibiting xanthine oxidase activity.<sup>62,63</sup> Luteolin (3',4',5',7-tetrahydroxyflavone) has been reported to be a potent inhibitor of xanthine oxidase.<sup>64</sup>

### **Leukocyte Immobilization Mechanism**

Another crucial mechanism for the production of reactive oxygen species, release of mediators of inflammation (bradykinin, PGE<sub>2</sub>) and cytotoxic oxidants is the strong adhesion of leukocytes to the endothelial wall. This route is responsible for further activation of the complement system as well.<sup>41,45</sup> Unlike during normal conditions where leukocytes roam freely, at the time of inflammation and ischemia several complement factors and endothelium derived mediators make leukocytes adhere to the endothelial wall. Because of this leukocytes get immobilized and degranulation of neutrophils starts. As an outcome, inflammatory mediators and oxidants are released which cause damage to various cells and tissues. The concentration of immobilized leukocytes in ischemia-reperfusion injury was reported to downturn after oral administration of fraction of purified micronized flavonoid.<sup>41,45,65</sup> Administration of flavonoids resulted in lowering of the concentration of immobilized leukocytes which is assumed to be related to the decline in total serum complement. This can be considered as a defensive mechanism against inflammation associated disorders (e.g.: reperfusion injury).<sup>65,66</sup> Stimulation of degranulation of neutrophils can be inhibited by many flavonoids without having any effect on production of superoxide.<sup>67</sup> Modulation of Ca<sup>2+</sup> channels of the plasma membrane by flavonoids seem to have inhibitory effect on degranulation of mast cells.<sup>68</sup>

### **Interaction with Other Enzyme Systems**

The main effects of flavonoids are an outcome of their radical scavenging property.<sup>69</sup> Interaction of flavonoids with several enzyme functions is another route via which flavonoids perform their action. Moreover, some effects can be an outcome of combination of two mech-

anisms i.e. interaction with enzyme systems and radical scavenging. Lipid peroxidation occurs when reactive chemical species containing oxygen are present in the vicinity of iron.<sup>70</sup> Some flavonoids have been recognized to chelate iron,<sup>71</sup> by that they inhibit the formation of free radicals. Quercetin is one such flavonoid which is acknowledged for its iron-stabilizing and iron-chelating properties. Directly inhibiting lipid peroxidation is one more protective action which flavonoids perform.<sup>72</sup> Explicit flavonoids decrease inflammation by scaling down the adhesion and confinement of inflammatory cells to the endothelial wall by decreasing complement activation.<sup>66,73</sup> Diminishing the peroxidase release is another characteristic of flavonoids. Diminished production of peroxidase impedes the production of reactive oxygen species by neutrophils by meddling with the activation of  $\alpha$ 1-antitrypsin.<sup>74,75</sup> Thereafter a gradual deactivation of proteolytic enzymes was reported to occur in neutrophils.<sup>75</sup> Flavonoids, having ability to obstruct various enzyme systems, hamper arachidonic acid's metabolism as well.<sup>76</sup> This characteristic of flavonoids allows them to perform their antithrombotic and anti-inflammatory actions. Process of inflammation starts with the production and release of arachidonic acid. Chemotactic agents (movement towards a chemical gradient) are formed from neutrophils consisting of lipoxygenase. Release of cytokines is provoked by them as well.<sup>41,77</sup>

## **PHARMACOLOGICAL PROPERTIES**

### **Anti-Parkinson**

It is a progressive degenerative disorder. Progressive degeneration of neurons occurs in substantia nigra pars compacta and nigrostriatal tract.<sup>78</sup> The etiology of Parkinson's disease is extremely complicated with various factors playing roles such as environment, genetics and aging.<sup>79</sup> Neurodegeneration occurs as a result of several biological processes involving oxidative stress,<sup>80-83</sup> augmented iron deposition,<sup>84-86</sup> DNA damage,<sup>87,88</sup> lipid peroxidation,<sup>89</sup> reduced glutathione (GSH) levels,<sup>90,91</sup> oxidation of protein,<sup>88</sup> and elevated superoxide dismutase level.<sup>92,93</sup> Lipopolysaccharide, an external stimuli could generate ROS which can alleviate the endogenous antioxidant enzymes specifically glutathione peroxidase, catalase and superoxide dismutase and leads to upswing in lipid peroxidation and cell death.<sup>94,95</sup> Mitochondrial metabolism can get affected by ROS directly. ROS causes lipid peroxidation that progressively causes cytochrome-c's leakage from mitochondria and ultimately cell death. External stimuli also trigger proapoptotic caspases by activating MAPK-induced inflammatory medi-

ators which cause cellular apoptosis.<sup>96</sup> Commencement of proinflammatory cytokine genes (iNOS, TNF- $\alpha$  and IL-1 $\beta$ ) expressions induced by NF- $\kappa$ B, is also caused by MAPK family.<sup>96,97</sup> Flavonoids for example emodin,<sup>98</sup> kaempferol,<sup>99</sup> genistein<sup>100</sup> and morin<sup>101</sup> have been proved to suppress secretion of TNF- $\alpha$ . Naringenin has been reported to alleviate expression of NF- $\kappa$ B, iNOS and COX-2.<sup>102</sup>

### Anti-Ulcer

Peptic ulcer occurs in that part of the gastrointestinal tract which is exposed to gastric acid and pepsin. It results probably due to an imbalance between the aggressive (acid, pepsin, bile and *H. pylori*) and defensive (gastric mucus, nitric oxide and bicarbonate secretion) factors.<sup>103</sup> The probable anti-ulcer effect of hesperidin has been due to its antioxidant and mucoprotective effect. Hesperidin impedes oxidative cell injury by augmenting the levels of certain enzymes (superoxide dismutase, catalase and glutathione) in gastric mucosa. Free radicals play major role in formation of stomach ulcers. Hesperidin allows the regeneration of ulcerated tissue and prevented hemorrhagic injury of gastric mucosa.<sup>104</sup> Quercetin has been found to have antiulcer activity in animals.<sup>105-107</sup> It acts by inhibiting the enzyme histidine decarboxylase<sup>108, 109</sup> and thus reduce the formation of histamine in the gastric mucosa, which stimulates the parietal cells and pepsinogen responsible for the secretion of hydrochloric acid and pepsin respectively.<sup>110</sup> Manuka honey, which is rich in flavonoids, preserves the gastric mucosal GSH.<sup>111</sup> GSH and gastric mucus both act as a barrier against gastric mucosal injury.<sup>112</sup> *Matricaria chamomilla*, which contains apigenin-7-O- $\beta$ -glucoside-6''acetate, apigenin-7-O-galactoside-6'' acetate and apigenin-7-O- $\beta$ -glucoside, has been found to exhibit antiulcer effect.<sup>113</sup>

### Spasmolytic

Spasmolytic effect occurs by blocking M<sub>3</sub> receptor (visceral smooth muscle contraction is elicited through M<sub>3</sub> receptor).<sup>114</sup> Catechin promotes vasodilation by activating muscarinic receptors on the endothelium and hence stimulates endothelium-dependent nitric oxide production.<sup>115</sup> Studies have shown that catechin has been reported to have a vasodilator effect mediated through numerous pathways including reduction in Ca<sup>2+</sup> uptake, upturn of cyclic adenosine monophosphate (cAMP) levels and inhibition of protein kinase C.<sup>116</sup> Probable Ca<sup>2+</sup> channel blocking action adds to the already compelling profile of catechin.<sup>115</sup> Thyme extract and flavones inhibit responses of particular receptors like histamine, L-noradrenaline and acetylcholine by inhibiting responses to such agonists which stimulate

these receptors.<sup>117</sup> Flavonoids suppress contraction of smooth muscles caused by influx of extra-cellular Ca<sup>2+</sup> into the guinea-pig longitudinal muscle.<sup>118</sup>

### Anti-Depressant

In depression, monoaminergic transmission in the brain gets affected (5-HT and/or NE gets depleted).<sup>119</sup> Several flavonoids including quercetin have shown inhibitory action against MAO-A.<sup>120,121</sup> Monoamine oxidase-A is responsible for oxidative deamination of 5-HT and NA. Hence, manifestations of depression can be ameliorated by inhibiting MAO-A.<sup>122</sup> Intake of reserpine repeatedly causes cognitive deficit and elevate cellular oxidative stress. Quercetin shows a protective effect against reserpine induced dysfunctions.<sup>123</sup> A downturn in the levels of SOD and CAT was noted in groups that were provided with reserpine. Mice treated with *Hypericum bookerianum* (EEHh) and its glycosidic flavonoid enriched extract (GFHh) were able to maintain normal levels of SOD and CAT enzymes.<sup>124</sup> Flavonoids, quercetin and rutin, act against dysfunctions induced by reserpine by scavenging upon reserpine generated oxygen-derived free radicals.<sup>125</sup> Antidepressant property of hesperidin was displayed by inhibiting L-arginine/ nitric oxide/ cyclic-GMP pathway and by elevating levels of BDNF in the brain, specifically in hippocampus.<sup>126,127</sup> A study showed antidepressant action of vitexin which was mediated through heightened levels of catecholamines (dopamine, adrenaline and noradrenaline) in the synaptic cleft and by interacting with dopaminergic, serotonergic and noradrenergic receptors.<sup>128</sup> Other flavonoids including fisetin,<sup>129</sup> quercetin,<sup>130</sup> naringenin,<sup>131,132</sup> nobiletin (a dietary flavonoid),<sup>133</sup> luteolin<sup>134</sup> and kaempferitrin<sup>135</sup> also have reported antidepressant activity.

### Anti-Bacterial

An Antibacterial agent is the one which interrupts the propagation and growth of bacteria.<sup>136</sup> Apigenin-7-O-triglycoside, apigenin, luteolin-7-O-neohesperidoside, lucenin-2, saponarine and vitexin are some of the flavonoids which are isolated from mosses and have been proved to possess inhibitory effect against various bacteria. They have been shown to have antibacterial effect against several bacteria including *Enterobacter cloacae* and *Pseudomonas aeruginosa*.<sup>137</sup> Golnar extract has played a successful role in preventing food poisoning as it exhibited antibacterial action against both gram positive and gram negative food poisoning causing bacteria.<sup>138</sup>

### Anti-Hypertensive

When given chronically, Quercetin showed a gradual dose dependant and sustain fall in BP of rats.<sup>139</sup> Quercetin inhibits oxygen-derived free radicals and exer-

cises its inhibitory action against several transcription factors, enzymes and ion channels as well.<sup>140</sup> Hence several changes in cell functioning and gene expression are caused by quercetin by interfering with different signal relay pathways. Thus quercetin's vasodilatory action is a possible mechanism via which it shows its antihypertensive effect. ROS have been assumed to have pathophysiological role in essential hypertension and therefore decrease in cellular oxidative stress by quercetin could be considered as a possible mechanism via which it shows its antihypertensive effect. In spontaneously hypertensive rats (SHRs) quercetin alleviated superoxide ions which is related to down regulation of NADPH oxidase subunits.<sup>141</sup> Assessment of BP in rats at the end of 5 weeks treatment demonstrated that quercetin exhibited remarkable reduction in diastolic, systolic and mean arterial BP in SHRs. Quercetin markedly decreased both heart rate and BP in spontaneously hypertensive rats.<sup>142</sup> In male wistar rats antihypertensive action of flavonoids chrysin and luteolin was efficiently investigated. Chrysin and luteolin both have the capability to reduce BP and heart rate of diabetic rats which is associated with their vasorelaxation action.<sup>143,144</sup> Kaempferol does not need functional endothelium to initiate vasodilation of blood vessels and hence considered as endothelium-independent vasodilator which acts similar to sodium nitroprusside. Epicatechin and myricetin both flavonoids showed their inhibitory action against vasoconstrictors (endothelin-1 and angiotensin II).<sup>146,147</sup> The fall in blood pressure has been accomplished by various flavonoids via their effects on functions of epithelium. Different classes of flavonoids such as flavanones (naringin and hesperidin), flavanols (epicatechin) and flavones (luteolin, buddleioside and chrysin), all have exhibited vasodilatory effect.<sup>145,148-150</sup> Many studies have showed that naringin, hesperidin, quercetin and epicatechin have augmented nitric oxide synthase activity and bioavailability in the endothelium which enhanced endothelial function.<sup>151-155</sup> Acetylcholine-induced vasodilation was improved *invitro* by naringin, hesperidin, luteolin and epicatechin.<sup>144,146,155</sup>

### Anti-Inflammatory

The antiinflammatory activity of flavonoids is mediated through a number of mechanisms including inhibition of proinflammatory enzymes like lipooxygenase, Cyclooxygenase-2 and iNOS. At molecular level, flavonoids stimulate protein kinase C, phase II antioxidant and detoxifying enzymes and mitogen activated protein kinase (MAPK). Flavonoids also show inhibitory action against NF- $\kappa$ B.<sup>156-158</sup> In peritoneal macrophages of rats, kaempferol and quercetin inhibit COX-2.<sup>159</sup> Catechin

quite infirmly inhibits Cyclooxygenase-2 and that too at an extremely large concentration.<sup>160</sup> Whereas some flavonoids like myricetin, kaempferol, quercetin and morin act by inhibiting lipooxygenase. Apigenin, quercetin and luteolin inhibit COX-2 at very high concentrations and inhibit NO production.<sup>161</sup> Catechin and quercetin showed synergistic inhibitory action against tumor necrosis factor alpha (TNF- $\alpha$ ) and Interleukin 1 beta (IL-1 $\beta$ ) and augment the release of IL-10, also known as human cytokine synthesis inhibitory factor.<sup>156</sup> Genistein has been proved to inhibit TNF- $\alpha$ , IL-1 $\beta$  and IL-6 in LPS induced RAW cells.<sup>156</sup> Quercetin showed its affect on iNOS and TNF- $\alpha$  in RAW cells treated with LPS by blocking MAPK and AP-1 DNA binding.<sup>163,164</sup>

### Anti-Diabetic

Diabetes mellitus is a metabolic disorder indicated by conditions such as hyperglycaemia, negative nitrogen balance, hyperlipidaemia, glycosuria and sometimes ketonaemia. Two major types of diabetes mellitus are Type-I Insulin-dependent diabetes mellitus (IDDM) and Type-II Noninsulin-dependent diabetes mellitus (NIDDM).<sup>165</sup>

Various researchers have proved that flavonoids scale down diabetes mellitus either by avoiding glucose absorption or by improving glucose tolerance.<sup>166</sup> *Invitro* experiments have stated that isoflavones of soyabean extract (daidzein and genistein) impede absorption of glucose in the small intestinal brush-border-membrane (BBM) vesicles of rabbits.<sup>167</sup> Naringenin also decreased the level of uptake of glucose into the BBM vesicles of diabetic rats equivalent to normal rats.<sup>168</sup> Several flavonoids including (-)-epigallocatechin, (-)-epigallocatechin gallate, myricetin, quercetin, apigenin and (-)-epicatechin gallate ameliorate diabetes mellitus by inhibition of Na<sup>+</sup> dependent glucose transporter-1 (SGLT-1).<sup>169</sup> In both *invivo* and *invitro* conditions of animal tissues, non-glycosylated flavonoids showed reduction in glucose absorption under Na<sup>+</sup> dependent conditions.<sup>170,171</sup> Flavonoids alleviate diabetes mellitus via several mechanisms. Most common being reduction in glucose absorption. Another mechanism via which flavonoids act is by inhibiting the activity of  $\alpha$ -glucosidase in the small intestine. Kaempferol, luteolin, galangin and chrysin showed  $\alpha$ -glucosidase inhibitory activity in both *invivo* and *invitro* conditions when used to study their roles in absorption and metabolism of glycosides.<sup>172</sup>

Amentoflavone, daidzein, luteolin and luteolin 7-O-glucoside have been proved to be the strongest inhibitors of  $\alpha$ -glucosidase among the twenty-one tested compounds.<sup>173</sup> Orientoside too was shown to impede

$\alpha$ -glycosidase function. The lowest possible level of blood glucose is achieved after 4 h of dosing of quercetin.<sup>174</sup> A research has proved that rutin has higher activity as compared to ellagic acid, boswellic acid and quercetin. They act by upsurging the peripheral utilization of glucose and by obstructing the glucose transporter function in intestine. Hypoglycemic activity of these four flavonoids were noticed in this mentioned order- Rutin > Quercetin > Ellagic acid > Boswellic acid.<sup>175</sup> Diosmin stimulates the production of insulin from  $\beta$ -cells of pancreas.<sup>176-178</sup> Studies revealed that in contrast with casein, soya protein isolate show better hypotriglyceridemic effect. This proposed that partly isoflavones are responsible for this activity.<sup>179</sup> Another study has reported that genistein improves hyperglycemia, promotes cAMP/PKA signaling pathway and causes human vascular endothelial inflammation *ex vivo*.<sup>180</sup> Several reports have shown that genistein ameliorates glucose tolerance, hyperglycemia and blood insulin level in obese diabetic mice without having any affect on fat deposit, peripheral insulin sensitivity, body weight gain, plasma lipid profile and food intake.<sup>181</sup> Intake of genistein resulted in improved cardiac remodeling advancement in experimentally induced diabetes which was mediated partly by inhibiting the actions of CRP (C-reactive protein), TGF- $\beta$ 1 (Transforming growth factor  $\beta$ 1) and TNF- $\alpha$  (Tumor necrosis factor- $\alpha$ ).<sup>182</sup> By controlling intracellular signaling mechanism of AMPK (AMP-activated kinase), genistein, EGCG and capsaicin improve obesity.<sup>183</sup>

### Anti-Cancer

In cancer uncontrolled cellular function occurs. Cancerous cells escape normal cellular functions and normal homeostasis via suppression of tumor suppressor gene and alteration of normal cellular physiological functions and structure. Flavonoids of tea obstruct epidermal growth factor and platelet-derived growth factor mediated signal relay pathways. Malignant cells affect various events such as angiogenesis.<sup>184</sup> Tyrosine kinase is mediator of signal transduction process which causes cell proliferation, migration, differentiation and apoptosis. This tyrosine kinase is inhibited by flavonoids quercetin and genistein.<sup>185,186</sup> Flavonoids namely apigenin, luteolin and quercetin arrest cell growth and cause apoptosis mediated via p53 as stated by different reports.<sup>187</sup> These flavonoids have inhibitory and protective action against breast tumor as reported by several researchers. Genistein administration has improved the early maturation and differentiation of the mammary glands, which is assumed to be the mechanism of tumor obstructing activity of soya. Tumor inhibitory activity of isofla-

vones has been demonstrated through various studies conducted on different models.<sup>188</sup> Women who eat up high amount of tofu have been observed to have lower incidence of breast cancer.<sup>189</sup> Seventh Day Adventists and Japanese studies have stated that consumption of high amount of soya milk and tofu is correlated with less chances of prostate cancer.<sup>190,191</sup> Flavonoids in tea have been proved to have anti-cancer effect as stated by different studies.<sup>192,193</sup> Antitumor effect has been shown by oncamex, a new flavonoid, in animal models of breast carcinoma.<sup>194</sup>

Apigenin was observed to possess skin papillomas inhibitory activity and was seen to prevent their conversion to cancer as well.<sup>195</sup> Luteolin acts by penetrating into the skin for treatment and prevention of skin cancer.<sup>196</sup> Quercetin has also been reported to have activity against hepatic-cancer.<sup>197</sup> Kaempferol showed productive results in ovarian cancer by lowering vascular endothelial growth factor (VEGF) expression which causes increment in vascular proliferation and permeability.<sup>198</sup> Myricetin and baicalein showed cytotoxic activity against leukemia, an another type of cancer.<sup>199</sup> Quercetin was reported to impede thyroid cell growth by inhibiting insulin modulated AKT kinase activity. It downturns TSH- inflected RNA levels in sodium iodide symporter (NIS) gene and therefore considered to be a new disrupter of thyroid function which can be used in thyroid cancer.<sup>200</sup> Proliferation of KAT 18 and HTH 7 have been inhibited by Chrysin both time and dose dependently. An upturn in cleaved polyADP ribose polymerase (responsible for DNA repair, genomic integrity and apoptosis), cleaved caspase-3, along with a downturn in Mcl-1, cyclin D1 and XIAP (play role in the control of mitotic cell death) was detected.<sup>201</sup> Via the mechanism of induction of differentiation, human U937 leukemia cell line is inhibited by a novel flavonoid III-10.<sup>202,203</sup> Several researchers have reported the anticancer activity of alcoholic extracts of *Gracilaria tenuistipitata* in squamous cell carcinoma of mouth. Programmed cell death is induced by the alcoholic extract by enhancing ROS initiation, mitochondrial depolarization and DNA damage.<sup>204,205</sup> Another flavonoid Epigallocatechin-3-gallate (EGCG) has demonstrated to inhibit angiogenesis in the chorioallantoic membrane.<sup>206-208</sup>

### CONCLUSION

Flavonoids are naturally occurring compounds present in plants. The major dietary sources of flavonoids within the reach of human beings are vegetables and fruits including tea and wine. In the present study we discussed the chemistry and pharmacological activities such as anti-parkinson, anti-ulcer, spasmolytic,

anti-depressant, anti-bacterial, anti-hypertensive, anti-diabetic, anti-inflammatory and anti-cancer. This review intent to give healthy information for formation of new flavonoid based pharmaceutical formulation to act against various diseases.

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None

## CONFLICT OF INTEREST

The authors declare that there is no conflict of interest.

## ABBREVIATIONS

**LDL:** Low density lipoprotein; **ROS:** Reactive oxygen species; **PGEA2:** Prostaglandin E2; **MAPK:** Mitogen-activated protein kinase; **TNF:** Tissue necrotic factor; **SOD:** Superoxide dismutase; **BDNF:** Brain-derived neurotrophic factor.

## REFERENCES

- Rao V, Kiran SD, Rohini P, Bhagyasree P. Flavonoid: A review on Naringenin. *Journal of Pharmacognosy and Phytochemistry*. 2017;6(5):2778-83.
- Bhat SV, Nagasampagi BA, Sivakumar M. *Chemistry of natural products*. 1<sup>st</sup> ed. New Delhi: Narosa publishing house pvt. ltd. 2005.
- Mills S, Bone K. *Principles of herbal pharmacology in principles and practice of phytotherapy, modern herbal medicine*. 2<sup>nd</sup> ed. USA: Churchill Livingstone. 2013.
- Panche AN, Diwan AD, Chandra SR. Flavonoids: an overview. *Journal of Nutritional Science*. 2016;5:1-15.
- Falcone FML, Rius S, Casati P. Flavonoids: biosynthesis, biological functions and biotechnological applications. *Frontiers in Plant Science*. 2012;3:1-15.
- Kumar S, Pandey AK. Chemistry and biological activities of flavonoids: an overview. *The Scientific World Journal*. 2013;1-16.
- Tapas AR, Sakarkar DM, Kakde RB. Flavonoids as nutraceuticals: a review. *Tropical Journal of Pharmaceutical Research*. 2008;7(3):1089-99.
- Jones R, Ougham H, Thomas H, Waaland S. *Molecular life of plants*. 2<sup>nd</sup> ed. USA: Wiley-Blackwell. 2012.
- Heldt HW, Piechulla B. Phenylpropanoids comprise a multitude of plant secondary metabolites and cell wall components. In: *Plant Biochemistry*. 4<sup>th</sup> ed. New York. Elsevier Academic. 2011;431-49.
- Caballero B, Trugo L, Finglas P. *Encyclopedia of food sciences and nutrition*. 2<sup>nd</sup> ed. London: Academic Press. 2003.
- Watson RR, Zibadi S. Bioactive dietary factors and plant extracts in dermatology. New York: Humana Press. 2013.
- Verma AK, Pratap R. The biological potential of flavones. *Natural Product Reports*. 2010;27(11):1571-93.
- Grotewold E. *The science of flavonoids*. New York: Springer science + Business Media. 2006.
- Alzand KI, Mohamed MA. Flavonoids: Chemistry, biochemistry and antioxidant activity. *Journal of Pharmacy Research*. 2012;5:4013-2.
- Samanta A, Das G, Das SK. Roles of flavonoids in plants. *International Journal of Pharmacy and Pharmaceutical Sciences*. 2011;6(1):12-35.
- Pandey RP, Sohng JK. Genetics of Flavonoids. In: Ramawat, Gopal K, Mérillon, Michel J. *Natural Products-Phytochemistry, Botany, Metabolism of Alkaloids, Phenolics and Terpenes*. Germany: Springer-Verlag Berlin Heidelberg. 2013;1617-45.
- Andersen OM, Markham KR. *Flavonoids: chemistry, biochemistry and applications*. US: CRC Press. 2005.
- Kawser HM, Abdal DA, Han J, Yin Y, Kim K, Kumar SS, et al. Molecular mechanisms of the anti-obesity and anti-diabetic properties of flavonoids. *International Journal of Molecular Sciences*. 2016;17(4):1-32.
- Kuete V. *Medicinal plant research in Africa: Pharmacology and chemistry*. 1<sup>st</sup> ed. UK: Newnes. 2013.
- Mendes AP, Borges RS, Neto AM, de Macedo LG, da Silva AB. The basic antioxidant structure for flavonoid derivatives. *Journal of Molecular Modeling*. 2012;18(9):4073-80.
- Ververidis F, Trantas E, Douglas C, Vollmer G, Kretzschmar G, Panopoulos N. *Biotechnology of flavonoids and other phenylpropanoid-derived natural products*. Part I: Chemical diversity, impacts on plant biology and human health. *Biotechnology Journal: Healthcare Nutrition Technology*. 2007;2(10):1214-34.
- Novza YA, Popova EM. Flavonoids: chemistry and biological activities. *Problems of Environmental Biotechnology*. 2016;1:1-10.
- Farooq MA. Flavonoids their chemistry, spectra studies and uses -a review. *Indo American Journal of Pharmaceutical Sciences*. 2014;1(6):521-4.
- Sahu R, Saxena J. Screening of total phenolic and flavonoid content in conventional and non-conventional species of curcuma. *Journal of Pharmacognosy and Phytochemistry*. 2013;2(1):176-9.
- Lakhanpal P, Rai DK. Quercetin: a versatile flavonoid. *Internet Journal of Medical Update*. 2007;2(2):22-37.
- Watson RR, Preedy VR. *Bioactive foods in promoting health: fruits and vegetables*. 1<sup>st</sup> ed. New York: Academic Press. 2009.
- Grumzeescu A. *Nutraceuticals*. 1<sup>st</sup> ed. New York: Academic Press. 2016.
- Narayana KR, Reddy MS, Chaluvadi MR, Krishna DR. Bioflavonoids classification, pharmacological, biochemical effects and therapeutic potential. *Indian Journal of Pharmacology*. 2001;33(1):2-16.
- Ohadoma SC, Akah PA, Okolo CE. Isolation and Characterization of Flavonol Glycosides from Leaves Extract of *Lupinus arboreus* Sims. *UK Journal of Pharmaceutical and Biosciences*. 2016;4(3):6-9.
- Nabi NG, Shrivastava M. Estimation of Total Flavonoids and Antioxidant Activity of *Spilanthes acmella* Leaves. *UK Journal of Pharmaceutical and Biosciences*. 2016;4(6):29-34.
- Rasha R, Latif AE, Mansour RMA. New flavone glycoside from *Stellaria pallid* (Dumort.) and biological activities. *International Journal of Pharmaceutical Sciences Review and Research*. 2016;36(2):77-8.
- Yao LH, Jiang YM, Shi J, Tomas-Barberan FA, Datta N, Singanusong R, et al. Flavonoids in food and their health benefits. *Plant Foods for Human Nutrition*. 2004;59(3):113-22.
- Dixon RA, Dey PM, Lamb CJ. Phytoalexins: enzymology and molecular biology. *Advances in Enzymology and Related Areas of Molecular Biology*. 1983;55(1):69.
- Aherne SA, O'Brien NM. Dietary flavonols: chemistry, food content and metabolism. *Nutrition*. 2002;18(1):75-81.
- Watson RR. *Complementary and Alternative Therapies and the Aging Population: An Evidence-Based Approach*. 1<sup>st</sup> ed. New York: Academic Press. 2011.
- Moon YJ, Wang X, Morris ME. Dietary flavonoids: effects on xenobiotic and carcinogen metabolism. *Toxicology in vitro*. 2006;20(2):187-210.
- Watson RR, Preedy VR, Zibadi S. *Polyphenols in human health and disease*. New York: Academic press. 2013.
- Thilakarathna SH, Rupasinghe HP. Flavonoid bioavailability and attempts for bioavailability enhancement. *Nutrients*. 2013;5(9):3367-87.
- Landete JM. Updated knowledge about polyphenols: functions, bioavailability, metabolism and health. *Critical Reviews in Food Science and Nutrition*. 2012;52(10):936-48.
- Scalbert A, Morand C, Manach C, Rémésy C. Absorption and metabolism of polyphenols in the gut and impact on health. *Biomedicine and Pharmacotherapy*. 2002;56(6):276-82.
- Nijveldt RJ, Van NEL, Van HDE, Boelens PG, Van NK, Leeuwen PAV. Flavonoids: a review of probable mechanisms of action and potential applications. *The American Journal of Clinical Nutrition*. 2001;74(4):418-25.
- De Groot H. Reactive oxygen species in tissue injury. *Hepato-gastroenterology*. 1994;41(4):328-32.
- Grace PA. Ischaemia-reperfusion injury. *British Journal of Surgery*. 1994;81(5):637-47.

44. Sivamani RK, Jagdeo JR, Elsner P, Maibach HI, editors. *Cosmeceuticals and active cosmetics*. 3<sup>rd</sup> ed. US: CRC Press. 2015.
45. Gupta J, Gupta A, Gupta AK. Flavonoids: its working mechanism and various protective roles. *International Journal of Chemical Studies*. 2016;4:190-8.
46. Korkina LG, Afanas'ev I. Antioxidant and chelating properties of flavonoids. In: Sies H, editor. *Advances in Pharmacology* New York: Academic Press. 1996;:151-63.
47. Hanasaki Y, Ogawa S, Fukui S. The correlation between active oxygens scavenging and antioxidative effects of flavonoids. *Free Radical Biology and Medicine*. 1994;16(6):845-50.
48. Kerry NL, Abbey M. Red wine and fractionated phenolic compounds prepared from red wine inhibit low density lipoprotein oxidation *in vitro*. *Atherosclerosis*. 1997;135(1):93-102.
49. Horáková L. Flavonoids in prevention of diseases with respect to modulation of Ca-pump function. *Interdisciplinary Toxicology*. 2011;4(3):114-24.
50. Shoskes DA. Effect of bioflavonoids quercetin and curcumin on ischemic renal injury: a new class of renoprotective agents. *Transplantation*. 1998;66(2):147-52.
51. Huk I, Brovkovich V, Nanobash VJ, Weigel G, Neumayer C, Partyka L, *et al*. Bioflavonoid quercetin scavenges superoxide and increases nitric oxide concentration in ischaemia-reperfusion injury: an experimental study. *British Journal of Surgery*. 1998;85(8):1080-5.
52. Assreuy J, Cunha FQ, Epperlein M, Noronha-Dutra A, O'Donnell CA, Liew FY, *et al*. Production of nitric oxide and superoxide by activated macrophages and killing of *Leishmania major*. *European Journal of Immunology*. 1994;24(3):672-6.
53. Vijayabaskaran M, Venkateswaramurthy N, Babu G, Perumal P, Jayakar B. *In vitro* antioxidant evaluation of *Pseudearthria viscida* Linn. *International Journal of Current Pharmaceutical Research*. 2010;2:21-3.
54. Abdel-Monem N, Abdel-Azeem AM, El Ashry ES, Ghareeb DA, Nabil-Adam A. Assessment of secondary metabolites from marine-derived fungi as antioxidant. *Open Journal of Medicinal Chemistry*. 2013;3(3):60-73.
55. Shutenko Z, Henry Y, Pinard E, Seylaz J, Potier P, Berthet F, *et al*. Influence of the antioxidant quercetin *in vivo* on the level of nitric oxide determined by electron paramagnetic resonance in rat brain during global ischemia and reperfusion. *Biochemical Pharmacology*. 1999;57(2):199-208.
56. Mruthunjaya K, Hukkeri V. *In vitro* antioxidant and free radical scavenging potential of *Parkinsonia aculeata* Linn. *Pharmacognosy Magazine*. 2008;4(13):42-51.
57. García-Mediavilla V, Crespo I, Collado PS, Esteller A, Sánchez-Campos S, Tuñón MJ, *et al*. The anti-inflammatory flavones quercetin and kaempferol cause inhibition of inducible nitric oxide synthase, cyclooxygenase-2 and reactive C-protein and down-regulation of the nuclear factor kappaB pathway in Chang Liver cells. *European Journal of Pharmacology*. 2007;557(2-3):221-9.
58. Vanacker SA, Tromp MN, Haenen GR, Vandervijgh WJ, Bast A. Flavonoids as scavengers of nitric oxide radical. *Biochemical and Biophysical Research Communications*. 1995;214(3):755-9.
59. Dehmlow C, Erhard J, de Groot HE. Inhibition of Kupffer cell functions as an explanation for the hepatoprotective properties of silibinin. *Hepatology*. 1996;23(4):749-54.
60. Kalogeris T, Baines CP, Krenz M, Korthuis RJ. Ischemia/Reperfusion. *Comprehensive Physiology*. 2011;7(1):113-70.
61. Sanhuesa J, Valdes J, Campos R, Garrido A, Valenzuela A. Changes in the xanthine dehydrogenase/xanthine oxidase ratio in the rat kidney subjected to ischemia-reperfusion stress: preventive effect of some flavonoids. *Research Communications in Chemical Pathology and Pharmacology*. 1992;78(2):211-8.
62. Chang WS, Lee YJ, Lu FJ, Chiang HC. Inhibitory effects of flavonoids on xanthine oxidase. *Anticancer Research*. 1993;13(6A):2165-70.
63. Iio M, Ono Y, Kai S, Fukumoto M. Effects of flavonoids on xanthine oxidation as well as on cytochrome c reduction by milk xanthine oxidase. *Journal of Nutritional Science and Vitaminology*. 1986;32(6):635-42.
64. Cos P, Ying L, Calomme M, Hu JP, Cimanga K, Van Poel B, *et al*. Structure-activity relationship and classification of flavonoids as inhibitors of xanthine oxidase and superoxide scavengers. *Journal of Natural Products*. 1998;61(1):71-6.
65. Friesenecker B, Tsai AG, Allegra C, Intaglietta M. Oral administration of purified micronized flavonoid fraction suppresses leukocyte adhesion in ischemia-reperfusion injury: *in vivo* observations in the hamster skin fold. *Journal of Vascular Research*. 1994;14(1-2):50-5.
66. Friesenecker B, Tsai AG. Cellular basis of inflammation, edema and the activity of Daflon 500 mg. *International Journal of Microcirculation*. 1995;15(Suppl 1):17-21.
67. Ferrándiz ML, Gil B, Sanz MJ, Ubeda A, Erazo S, González E, *et al*. Effect of bakuchiol on leukocyte functions and some inflammatory responses in mice. *Journal of Pharmacy and Pharmacology*. 1996;48(9):975-80.
68. Bennett JP, Gomperts BD, Wollenweber E. Inhibitory effects of natural flavonoids on secretion from mast cells and neutrophils. *Arzneimittelforschung*. 1981;31(3):433-7.
69. Es-Safi NE, Ghidouche S, Ducrot PH. Flavonoids: hemisynthesis, reactivity, characterization and free radical scavenging activity. *Molecules*. 2007;12(9):2228-58.
70. Nelson CW, Wei EP, Povlishock JT, Kontos HA, Moskowitz MA. Oxygen radicals in cerebral ischemia. *American Journal of Physiology-Heart and Circulatory Physiology*. 1992;263(5):H1356-62.
71. Ferrali M, Signorini C, Caciotti B, Sugherini L, Ciccoli L, Giachetti D, *et al*. Protection against oxidative damage of erythrocyte membrane by the flavonoid quercetin and its relation to iron chelating activity. *FEBS Letters*. 1997;416(2):123-9.
72. Sorata Y, Takahama U, Kimura M. Protective effect of quercetin and rutin on photosensitized lysis of human erythrocytes in the presence of hematoporphyrin. *Biochimica et Biophysica Acta (BBA)-General Subjects*. 1984;799(3):313-7.
73. Lago JH, Toledo-Arruda AC, Mernak M, Barrosa KH, Martins MA, Tibério IF, *et al*. Structure-activity association of flavonoids in lung diseases. *Molecules*. 2014;19(3):3570-95.
74. Hussain T, Tan B, Yin Y, Blachier F, Tossou MC, Rahu N. Oxidative stress and inflammation: what polyphenols can do for us?. *Oxidative Medicine and Cellular Longevity*. 2016;1-10.
75. Middleton JE, Kandaswami C. Effects of flavonoids on immune and inflammatory cell functions. *Biochemical Pharmacology*. 1992;43(6):1167-79.
76. Ferrandiz ML, Alcaraz MJ. Anti-inflammatory activity and inhibition of arachidonic acid metabolism by flavonoids. *Agents and Actions*. 1991;32(3-4):283-8.
77. Kesarkar S, Bhandage A, Deshmukh S, Shevkar K, Abhyankar M. Flavonoids: an overview. *Journal of Pharmacy Research*. 2009;2(6):1148-54.
78. Tripathi KD. *Antiparkinsonian Drugs*. In: *Essentials of medical pharmacology*. 7<sup>th</sup> ed. New Delhi: JP Medical Ltd. 2013;425.
79. Chu Y, Kordower JH. The use of aged monkeys to study PD: important roles in pathogenesis and experimental therapeutics. *Parkinson's Disease*. 2008;77-85.
80. Simonian NA, Coyle JT. Oxidative stress in neurodegenerative diseases. *Annual review of Pharmacology and Toxicology*. 1996;36(1):83-106.
81. Jenner P. Oxidative mechanisms in nigral cell death in Parkinson's disease. *Movement Disorders: Official Journal of the Movement Disorder Society*. 1998;13:24-34.
82. Olanow CW, Tatton WG. Etiology and pathogenesis of Parkinson's disease. *Annual Review of Neuroscience*. 1999;22(1):123-44.
83. Datla KP, Christidou M, Widmer WW, Rooprai HK, Dexter DT. Tissue distribution and neuroprotective effects of citrus flavonoid tangeretin in a rat model of Parkinson's disease. *Neuroreport*. 2001;12(17):3871-5.
84. Dexter DT, Wells FR, Agid F, Agid Y, Lees AJ, Jenner P, *et al*. Increased nigral iron content in postmortem parkinsonian brain. *The Lancet*. 1987;330(8569):1219-20.
85. Dexter DT, Wells FR, Lee AJ, Agid F, Agid Y, Jenner P, *et al*. Increased nigral iron content and alterations in other metal ions occurring in brain in Parkinson's disease. *Journal of Neurochemistry*. 1989;52(6):1830-6.
86. Sofic E, Paulus W, Jellinger K, Riederer P, Youdim MB. Selective increase of iron in substantia nigra zona compacta of parkinsonian brains. *Journal of Neurochemistry*. 1991;56(3):978-82.
87. Sanchez-Pamos J. A marker of oxyradical-mediated DNA damage (oxo8dG) is increased in nigrostriatum of Parkinson's disease brain. *Neurodegeneration (Experimental Neurology)*. 1994;3:197-204.

88. Alam ZI, Jenner A, Daniel SE, Lees AJ, Cairns N, Marsden CD, et al. Oxidative DNA damage in the parkinsonian brain: an apparent selective increase in 8-hydroxyguanine levels in substantia nigra. *Journal of Neurochemistry*. 1997;69(3):1196-203.
89. Dexter DT, Carter CJ, Wells FR, Javoy-Agid F, Agid Y, Lees A, et al. Basal lipid peroxidation in substantia nigra is increased in Parkinson's disease. *Journal of Neurochemistry*. 1989;52(2):381-9.
90. Sofic E, Lange KW, Jellinger K, Riederer P. Reduced and oxidized glutathione in the substantia nigra of patients with Parkinson's disease. *Neuroscience Letters*. 1992;142(2):128-30.
91. Sian J, Dexter DT, Lees AJ, Daniel S, Jenner P, Marsden CD. Glutathion-related enzymes in brain in Parkinson's disease. *Annals of Neurology: Official Journal of the American Neurological Association and the Child Neurology Society*. 1994;36(3):356-61.
92. Yoshida E, Mokuno K, Aoki SI, Takahashi A, Riku S, Murayama T, et al. Cerebrospinal fluid levels of superoxide dismutases in neurological diseases detected by sensitive enzyme immunoassays. *Journal of the Neurological Sciences*. 1994;124(1):25-31.
93. Yoritaka A, Hattori N, Mori H, Kato K, Mizuno Y. An immunohistochemical study on manganese superoxide dismutase in Parkinson's disease. *Journal of the Neurological Sciences*. 1997;148(2):181-6.
94. Li C, Zhou HM. The role of manganese superoxide dismutase in inflammation defense. *Enzyme Research*. 2011;1:7.
95. Floyd RA, Carney JM. Free radical damage to protein and DNA: mechanisms involved and relevant observations on brain undergoing oxidative stress. *Annals of Neurology*. 1992;32(S1):S22-7.
96. Magalingam KB, Radhakrishnan AK, Haleagrahara N. Protective mechanisms of flavonoids in Parkinson's disease. *Oxidative Medicine and Cellular Longevity*. 2015;1:15.
97. Bulua AC, Simon A, Maddipati R, Pelletier M, Park H, Kim KY, et al. Mitochondrial reactive oxygen species promote production of proinflammatory cytokines and are elevated in TNFR1-associated periodic syndrome (TRAPS). *Journal of Experimental Medicine*. 2011;208(3):519-33.
98. Jia Z, Babu PV, Si H, Nallasamy P, Zhu H, Zhen W, et al. Genistein inhibits TNF- $\alpha$ -induced endothelial inflammation through the protein kinase pathway A and improves vascular inflammation in C57BL/6 mice. *International Journal of Cardiology*. 2013;168(3):2637-45.
99. Qureshi AA, Guan XQ, Reis JC, Papasian CJ, Jabre S, Morrison DC, et al. Inhibition of nitric oxide and inflammatory cytokines in LPS-stimulated murine macrophages by resveratrol, a potent proteasome inhibitor. *Lipids in Health and Disease*. 2012;11(1):1-17.
100. Chen X, Yang X, Liu T, Guan M, Feng X, Dong W, et al. Kaempferol regulates MAPKs and NF- $\kappa$ B signaling pathways to attenuate LPS-induced acute lung injury in mice. *International Immunopharmacology*. 2012;14(2):209-16.
101. Zhu X, Zeng K, Qiu Y, Yan F, Lin C. Therapeutic effect of emodin on collagen-induced arthritis in mice. *Inflammation*. 2013;36(6):1253-9.
102. Park HY, Kim GY, Choi YH. Naringenin attenuates the release of pro-inflammatory mediators from lipopolysaccharide-stimulated BV2 microglia by inactivating nuclear factor- $\kappa$ B and inhibiting mitogen-activated protein kinases. *International Journal of Molecular Medicine*. 2012;30(1):204-10.
103. Tripathi KD. *Drugs for Peptic Ulcer and Gastroesophageal Reflux Disease*. In: *Essentials of medical pharmacology*. 7<sup>th</sup> ed. New Delhi: JP Medical Ltd. 2013;647.
104. Bigoniya P, Singh K. Ulcer protective potential of standardized hesperidin, a citrus flavonoid isolated from *Citrus sinensis*. *Revista Brasileira de Farmacognosia*. 2014;24(3):330-40.
105. Motilva V, Alarc de la LCÓ, Mart CMÍ, Torreblanca J. Effects of naringenin and quercetin on experimental chronic gastric ulcer in rat. Studies on the histological findings. *Phytotherapy Research*. 1992;6(3):168-70.
106. Konturek SJ, Radecki T, Brzozowski T, Drozdowicz D, Piastucki I, Muramatsu M, et al. Antiulcer and gastroprotective effects of solon, a synthetic flavonoid derivative of sophoradin. Role of endogenous prostaglandins. *European Journal of Pharmacology*. 1986;125(2):185-92.
107. Carlo GD, Mascolo N, Izzo AA, Capasso F, Autore G. Effects of quercetin on the gastrointestinal tract in rats and mice. *Phytotherapy Research*. 1994;8(1):42-5.
108. Konturek SJ, Kitler ME, Brzozowski T, Radecki T. Gastric protection by mercandanol, a new synthetic flavonoid inhibiting histidine decarboxylase. *Gastroenterology*. 1985;88:1452-7.
109. Hayes NA, Foreman JC. The activity of compounds extracted from feverfew on histamine release from rat mast cells. *Journal of Pharmacy and Pharmacology*. 1987;39(6):466-70.
110. Izzo AA, Carlo GD, Mascolo N, Capasso F, Autore G. Antiulcer effect of flavonoids. Role of endogenous PAF. *Phytotherapy Research*. 1994;8(3):179-81.
111. Almasaudi SB, El-Shitany NA, Abbas AT, Abdel-dayem UA, Ali SS, Al Jaouni SK, et al. Antioxidant, anti-inflammatory and antiulcer potential of manuka honey against gastric ulcer in rats. *Oxidative Medicine and Cellular Longevity*. 2016;1:10.
112. Cnubben NH, Rietjens IM, Wortelboer H, Zanden JV, Bladeren PJV. The interplay of glutathione-related processes in antioxidant defense. *Environmental Toxicology and Pharmacology*. 2001;10(4):141-52.
113. El Souda SS, Ahmed KM, Grace MH, Elkherassy EE, Farrag AR, Abdelwahab SM. Flavonoids and gastroprotective effect of *Matricaria chamomilla* against indomethacin-induced ulcer in rats. *Journal of Herbs, Spices and Medicinal Plants*. 2015;21(2):111-7.
114. Tripathi KD. *Cholinergic System and Drugs*. In: *Essentials of medical pharmacology*. 7<sup>th</sup> ed. New Delhi: JP Medical Ltd. 2013;101.
115. Ghayur MN, Khan H, Gilani AH. Antispasmodic, bronchodilator and vasodilator activities of (+)-catechin, a naturally occurring flavonoid. *Archives of Pharmacal Research*. 2007;30(8):970-5.
116. Duarte J, Pérez FV, Utrilla P, Jiménez J, Tamargo J, Zarzuelo A. Vasodilatory effects of flavonoids in rat aortic smooth muscle. Structure-activity relationships. *General Pharmacology*. 1993;24(4):857-62.
117. Broucke COVD, Lemli JA. Spasmolytic activity of the flavonoids from *Thymus vulgaris*. *Pharmaceutisch Weekblad*. 1983;5(1):9-14.
118. Vasconcelos LH, Correia AC, de Souza IL, da Cc Silva M, Paredes-Gamero EJ, Bárbara VD, et al. Flavonoid galeitin 3, 6-dimethyl ether attenuates guinea pig ileum contraction through K<sup>+</sup> channel activation and decrease in cytosolic calcium concentration. *European Journal of Pharmacology*. 2015;767:52-60.
119. Tripathi KD. *Drugs used in Mental Illness: Antidepressant and Antianxiety Drugs*. In: *Essentials of medical pharmacology*. 7<sup>th</sup> ed. New Delhi: JP Medical Ltd. 2013;454.
120. Chimenti F, Cottiglia F, Bonsignore L, Casu L, Casu M, Floris C, et al. Quercetin as the Active Principle of *Hypericum hircinum* Exerts a Selective Inhibitory Activity against MAO-A: Extraction, Biological Analysis and Computational Study. *Journal of Natural Products*. 2006;69(6):945-9.
121. Saaby L, Rasmussen HB, Jäger AK. MAO-A inhibitory activity of quercetin from *Calluna vulgaris* (L.) Hull. *Journal of Ethnopharmacology*. 2009;121(1):178-81.
122. Yamada M, Yasuhara H. Clinical pharmacology of MAO inhibitors: safety and future. *Neurotoxicology*. 2004;25(1-2):215-21.
123. Naidu PS, Singh A, Kulkarni SK. Reversal of reserpine-induced orofacial dyskinesia and cognitive dysfunction by quercetin. *Pharmacology*. 2004;70(2):59-67.
124. Subakanmani S, Murugan S, Devi PU. Evaluation of Antidepressant like Effects of Ethanolic *Hypericum hookerianum* and its Glycosidic Flavonoid Enriched Extract in Reserpine Induced Swiss Albino Mice. *Asian Journal of Biochemistry*. 2016;11(1):1-3.
125. Haenen GR, Bast A. [50] Nitric oxide radical scavenging of flavonoids. In: *Methods in Enzymology*. New York: Academic Press. 1990;301:490-503.
126. El-Marasy SA, Abdallah HM, El-Shenawy SM, El-Khatib AS, El-Shabrawy OA, Kenawy SA. Anti-depressant effect of hesperidin in diabetic rats. *Canadian Journal of Physiology and Pharmacology*. 2014;92(11):945-52.
127. Donato F, de Gomes MG, Goes AT, Borges FC, Fabbro LD, Antunes MS, et al. Hesperidin exerts antidepressant-like effects in acute and chronic treatments in mice: Possible role of L-arginine-NO-cGMP pathway and BDNF levels. *Brain Research Bulletin*. 2014;104:19-26.
128. Can ÖD, Özkay ÜD, Üçel Üİ. Anti-depressant-like effect of vitexin in BALB/c mice and evidence for the involvement of monoaminergic mechanisms. *European Journal of Pharmacology*. 2013;699(1-3):250-7.
129. Zhen L, Zhu J, Zhao X, Huang W, An Y, Li S, et al. The antidepressant-like effect of fisetin involves the serotonergic and noradrenergic system. *Behavioural Brain Research*. 2012;228(2):359-66.

130. Demir EA, Gergerlioglu HS, Oz M. Antidepressant-like effects of quercetin in diabetic rats are independent of hypothalamic–pituitary–adrenal axis. *Acta Neuropsychiatrica*. 2016;28(1):23-30.
131. Yi LT, Li CF, Zhan X, Cui CC, Xiao F, Zhou LP, *et al.* Involvement of monoaminergic system in the antidepressant-like effect of the flavonoid naringenin in mice. *Progress in Neuro-Psychopharmacology and Biological Psychiatry*. 2010;34(7):1223-8.
132. Yi LT, Li J, Li HC, Su DX, Quan XB, He XC, *et al.* Antidepressant-like behavioral, neurochemical and neuroendocrine effects of naringenin in the mouse repeated tail suspension test. *Progress in Neuro-Psychopharmacology and Biological Psychiatry*. 2012;39(1):175-81.
133. Yi LT, Xu HL, Feng J, Zhan X, Zhou LP, Cui CC. Involvement of monoaminergic systems in the antidepressant-like effect of nobiletin. *Physiology and Behavior*. 2011;102(1):1-6.
134. De la Peña JB, Kim CA, Lee HL, Yoon SY, Kim HJ, Hong EY, *et al.* Luteolin mediates the antidepressant-like effects of *Cirsium japonicum* in mice, possibly through modulation of the GABA A receptor. *Archives of Pharmacal Research*. 2014;37(2):263-9.
135. Cassani J, Dorantes-Barrón AM, Novales LM, Real GA, Estrada-Reyes R. Anti-depressant-like effect of kaempferitrin isolated from *Justicia spicigera* Schitld (Acanthaceae) in two behavior models in mice: evidence for the involvement of the serotonergic system. *Molecules*. 2014;19(12):21442-61.
136. Kohanski MA, Dwyer DJ, Collins JJ. How antibiotics kill bacteria: from targets to networks. *Nature Reviews Microbiology*. 2010;8(6):423.
137. Basile A, Giordano S, López-Sáez JA, Cobianchi RC. Antibacterial activity of pure flavonoids isolated from mosses. *Phytochemistry*. 1999;52(8):1479-82.
138. Mahboubi A, Asgarpanah J, Sadaghiyani PN, Faizi M. Total phenolic and flavonoid content and antibacterial activity of *Punica granatum* L. var. pleniflora flowers (Golnar) against bacterial strains causing foodborne diseases. *BMC Complementary and Alternative Medicine*. 2015;15(1):1-7.
139. Perez-Vizcaino F, Duarte J, Jimenez R, Santos-Buelga C, Osuna A. Antihypertensive effects of the flavonoid quercetin. *Pharmacological Reports*. 2009;61(1):67-75.
140. Middleton E, Kandaswami C, Theoharides TC. The effects of plant flavonoids on mammalian cells: implications for inflammation, heart disease and cancer. *Pharmacological Reviews*. 2000;52(4):673-751.
141. Sanchez M, Galisteo M, Vera R, Villar IC, Zarzuelo A, Tamargo J, *et al.* Quercetin down regulates NADPH oxidase, increases eNOS activity and prevents endothelial dysfunction in spontaneously hypertensive rats. *Journal of Hypertension*. 2006;24(1):75-84.
142. Duarte J, Pérez-Palencia R, Vargas F, Ocete MA, Pérez-Vizcaino F, Zarzuelo A, *et al.* Antihypertensive effects of the flavonoid quercetin in spontaneously hypertensive rats. *British Journal of Pharmacology*. 2001;133(1):117-24.
143. Clark JL, Zahradka P, Taylor CG. Efficacy of flavonoids in the management of high blood pressure. *Nutrition Reviews*. 2015;73(12):799-822.
144. El-Bassossy HM, Abo-Warda SM, Fahmy A. Chrysin and luteolin attenuate diabetes-induced impairment in endothelial-dependent relaxation: effect on lipid profile, AGEs and NO generation. *Phytotherapy Research*. 2013;27(11):1678-84.
145. Leeya Y, Mulvany MJ, Queiroz EF, Marston A, Hostettmann K, Jansakul C. Hypotensive activity of an n-butanol extract and their purified compounds from leaves of *Phyllanthus acidus* (L.) Skeels in rats. *European Journal of Pharmacology*. 2010;649(1-3):301-13.
146. Gómez-Guzmán M, Jiménez R, Sánchez M, Zarzuelo MJ, Galindo P, Quintela AM, *et al.* Epicatechin lowers blood pressure, restores endothelial function and decreases oxidative stress and endothelin-1 and NADPH oxidase activity in DOCA-salt hypertension. *Free Radical Biology and Medicine*. 2012;52(1):70-9.
147. Godse S, Mohan M, Kasture V, Kasture S. Effect of myricetin on blood pressure and metabolic alterations in fructose hypertensive rats. *Pharmaceutical Biology*. 2010;48(5):494-8.
148. Morand C, Dubray C, Milenkovic D, Lioger D, Martin JF, Scalbert A, *et al.* Hesperidin contributes to the vascular protective effects of orange juice: a randomized crossover study in healthy volunteers. *The American Journal of Clinical Nutrition*. 2010;93(1):73-80.
149. Alam MA, Kauter K, Brown L. Naringin improves diet-induced cardiovascular dysfunction and obesity in high carbohydrate, high fat diet-fed rats. *Nutrients*. 2013;5(3):637-50.
150. Olaleye MT, Crown OO, Akinmoladun AC, Akindahunsi AA. Rutin and quercetin show greater efficacy than nifedipin in ameliorating hemodynamic, redox and metabolite imbalances in sodium chloride-induced hypertensive rats. *Human and Experimental Toxicology*. 2014;33(6):602-8.
151. Rivera L, Morón R, Sánchez M, Zarzuelo A, Galisteo M. Quercetin ameliorates metabolic syndrome and improves the inflammatory status in obese Zucker rats. *Obesity*. 2008;16(9):2081-7.
152. Santangelo C, Vari R, Scazzocchio B, Di Benedetto R, Filesi C, Masella R. Polyphenols, intracellular signalling and inflammation. *Annali-istituto Superiore Di Sanita*. 2007;43(4):394-405.
153. Middleton E, Kandaswami C, Theoharides TC. The effects of plant flavonoids on mammalian cells: implications for inflammation, heart disease and cancer. *Pharmacological Reviews*. 2000;52(4):673-751.
154. Yoon JH, Baek SJ. Molecular targets of dietary polyphenols with anti-inflammatory properties. *Yonsei Medical Journal*. 2005;46(5):585-96.
155. Serafini M, Peluso I, Raguzzini A. Flavonoids as anti-inflammatory agents. *Proceedings of the Nutrition Society*. 2010;69(3):273-8.
156. Noreen Y, Serrano G, Perera P, Bohlin L. Flavan-3-ols isolated from some medicinal plants inhibiting COX-1 and COX-2 catalysed prostaglandin biosynthesis. *Planta Medica*. 1998;64(06):520-4.
157. Kim OK, Murakami A, Nakamura Y, Ohigashi H. Screening of edible Japanese plants for nitric oxide generation inhibitory activities in RAW 264.7 cells. *Cancer Letters*. 1998;125(1-2):199-207.
158. Cho JY, Kim PS, Park J, Yoo ES, Baik KU, Kim YK, *et al.* Inhibitor of tumor necrosis factor- $\alpha$  production in lipopolysaccharide-stimulated RAW264. 7 cells from *Amorpha fruticosa*. *Journal of Ethnopharmacology*. 2000;70(2):127-33.
159. Wadsworth TL, McDonald TL, Koop DR. Effects of Ginkgo biloba extract (EGb 761) and quercetin on lipopolysaccharide-induced signaling pathways involved in the release of tumor necrosis factor- $\alpha$ . *Biochemical Pharmacology*. 2001;62(7):963-74.
160. Wadsworth TL, Koop DR. Effects of Ginkgo biloba extract (EGb 761) and quercetin on lipopolysaccharide-induced release of nitric oxide. *Chemico-biological Interactions*. 2001;137(1):43-58.
161. Tripathi KD. Insulin, Oral Hypoglycaemic Drugs and Glucagon. In: *Essentials of medical pharmacology*. 7<sup>th</sup> ed. New Delhi: JP Medical Ltd. 2013;258.
162. Brahmachari G. Bio-flavonoids with promising antidiabetic potentials: A critical survey. *Research Signpost*. 2011;661(2):187-212.
163. Bhatthana SJ, Velasquez MT. Beneficial role of dietary phytoestrogens in obesity and diabetes. *The American Journal of Clinical Nutrition*. 2002;76(6):1191-201.
164. Li JM, Che CT, Lau CB, Leung PS, Cheng CH. Inhibition of intestinal and renal Na<sup>+</sup>-glucose cotransporter by naringenin. *The International Journal of Biochemistry and Cell Biology*. 2006;38(5-6):985-95.
165. Shimizu M, Kobayashi Y, Suzuki M, Satsu H, Miyamoto Y. Regulation of intestinal glucose transport by tea catechins. *Biofactors*. 2000;13(1-4):61-5.
166. Johnston K, Sharp P, Clifford M, Morgan L. Dietary polyphenols decrease glucose uptake by human intestinal Caco-2 cells. *FEBS letters*. 2005;579(7):1653-7.
167. Zhao H, Yakar S, Gavrilova O, Sun H, Zhang Y, Kim H, *et al.* Phloridzin improves hyperglycemia but not hepatic insulin resistance in a transgenic mouse model of type 2 diabetes. *Diabetes*. 2004;53(11):2901-9.
168. Matsui T, Kobayashi M, Hayashida S, Matsumoto K. Luteolin, a flavone, does not suppress postprandial glucose absorption through an inhibition of  $\alpha$ -glucosidase action. *Bioscience, Biotechnology and Biochemistry*. 2002;66(3):689-92.
169. Kim JS, Kwon CS, SoN KH. Inhibition of alpha-glucosidase and amylase by luteolin, a flavonoid. *Bioscience, Biotechnology and Biochemistry*. 2000;64(11):2458-61.
170. Djamil R, Winarti W, Zaidan S, Abdillah S. Antidiabetic Activity of Flavonoid from Binahong Leaves (*Anredera cordifolia*) Extract in Alloxan Induced Mice. *Journal of Pharmacognosy and Natural Products*. 2017;3(2):1-4.
171. Jadhav R, Puchchakayala G. Hypoglycemic and antidiabetic activity of flavonoids: boswellic acid, ellagic acid, quercetin, rutin on streptozotocin-nicotinamide induced type 2 diabetic rats. *International Journal of Pharmacy and Pharmaceutical Sciences*. 2012;4(2):251-6.
172. Pari L, Srinivasan S. Antihyperglycemic effect of diosmin on hepatic key enzymes of carbohydrate metabolism in streptozotocin-nicotinamide-induced diabetic rats. *Biomedicine and Pharmacotherapy*. 2010;64(7):477-81.

173. Srinivasan S, Pari L. Antihyperlipidemic effect of diosmin: A citrus flavonoid on lipid metabolism in experimental diabetic rats. *Journal of Functional Foods*. 2013;5(1):484-92.
174. Srinivasan S, Pari L. Ameliorative effect of diosmin, a citrus flavonoid against streptozotocin-nicotinamide generated oxidative stress induced diabetic rats. *Chemico-Biological Interactions*. 2012;195(1):43-51.
175. Demonty I, Lamarche B, Deshaies Y, Jacques H. Role of soy isoflavones in the hypotriglyceridemic effect of soy protein in the rat. *The Journal of Nutritional Biochemistry*. 2002;13(11):671-7.
176. Babu PV, Si H, Fu Z, Zhen W, Liu D. Genistein Prevents Hyperglycemia-Induced Monocyte Adhesion to Human Aortic Endothelial Cells through Preservation of the cAMP Signaling Pathway and Ameliorates Vascular Inflammation in Obese Diabetic Mice-3. *The Journal of Nutrition*. 2012;142(4):724-30.
177. Fu Z, Gilbert ER, Pfeiffer L, Zhang Y, Fu Y, Liu D. Genistein ameliorates hyperglycemia in a mouse model of nongenetic type 2 diabetes. *Applied Physiology, Nutrition and Metabolism*. 2012;37(3):480-8.
178. Gupta SK, Dongare S, Mathur R, Mohanty IR, Srivastava S, Mathur S, et al. Genistein ameliorates cardiac inflammation and oxidative stress in streptozotocin-induced diabetic cardiomyopathy in rats. *Molecular and Cellular Biochemistry*. 2015;408(1-2):63-72.
179. Hwang JT, Park IJ, Shin JI, Lee YK, Lee SK, Baik HW, et al. Genistein, EGCG and capsaicin inhibit adipocyte differentiation process via activating AMP-activated protein kinase. *Biochemical and Biophysical Research Communications*. 2005;338(2):694-9.
180. Wiseman S, Mulder T, Rietveld A. Tea flavonoids: bioavailability in vivo and effects on cell signaling pathways in vitro. *Antioxidants and Redox Signaling*. 2001;3(6):1009-21.
181. Akiyama T, Ishida J, Nakagawa S, Ogawara H, Watanabe SI, Itoh N, et al. Genistein, a specific inhibitor of tyrosine-specific protein kinases. *Journal of Biological Chemistry*. 1987;262(12):5592-5.
182. So FV, Guthrie N, Chambers AF, Moussa M, Carroll KK. Inhibition of human breast cancer cell proliferation and delay of mammary tumorigenesis by flavonoids and citrus juices. *Nutrition and Cancer*. 1996;26:167-81.
183. Plaumann B, Fritsche M, Rimpler H, Brandner G, Hess RD. Flavonoids activate wild-type p53. *Oncogene*. 1996;13(8):1605-14.
184. Messina MJ, Persky V, Setchell KD, Barnes S. Soy intake and cancer risk: a review of the *in vitro* and *in vivo* data. *Nutrition and Cancer*. 1994;21(2):113-31.
185. Wu AH, Ziegler RG, Horn-Ross PL, Nomura AM, West DW, Kolonel LN, et al. Tofu and risk of breast cancer in Asian-Americans. *Cancer Epidemiology and Prevention Biomarkers*. 1996;5(11):901-6.
186. Severson RK, Nomura AM, Grove JS, Stemmermann GN. A prospective study of demographics, diet and prostate cancer among men of Japanese ancestry in Hawaii. *Cancer Research*. 1989;49(7):1857-60.
187. Jacobsen BK, Knutsen SF, Fraser GE. Does high soy milk intake reduce prostate cancer incidence? The Adventist Health Study (United States). *Cancer Causes and Control*. 1998;9(6):553-7.
188. Yang CS, Landau JM, Huang MT, Newmark HL. Inhibition of carcinogenesis by dietary polyphenolic compounds. *Annual Review of Nutrition*. 2001;21(1):381-406.
189. Le Marchand L. Cancer preventive effects of flavonoids- a review. *Biomedicine and Pharmacotherapy*. 2002;56(6):296-301.
190. Martínez-Pérez C, Ward C, Turnbull AK, Mullen P, Cook G, Meehan J, et al. Antitumour activity of the novel flavonoid Oncamex in preclinical breast cancer models. *British Journal of Cancer*. 2016;114(8):905-16.
191. Wei H, Tye L, Bresnick E, Birt DF. Inhibitory effect of apigenin, a plant flavonoid, on epidermal ornithine decarboxylase and skin tumor promotion in mice. *Cancer Research*. 1990;50(3):499-502.
192. Seelinger G, Merfort I, Wölflle U, Schempp CM. Anti-carcinogenic effects of the flavonoid luteolin. *Molecules*. 2008;13(10):2628-51.
193. Seufi AM, Ibrahim SS, Elmaghaby TK, Hafez EE. Preventive effect of the flavonoid, quercetin, on hepatic cancer in rats via oxidant/antioxidant activity: molecular and histological evidences. *Journal of Experimental and Clinical Cancer Research*. 2009;28(1):1-8.
194. Luo H, Daddysman MK, Rankin GO, Jiang BH, Chen YC. Kaempferol enhances cisplatin's effect on ovarian cancer cells through promoting apoptosis caused by down regulation of cMyc. *Cancer cell international*. 2010;10(1):1-9.
195. Romanouskaya TV, Grinev VV. Cytotoxic effect of flavonoids on leukemia cells and normal cells of human blood. *Bulletin of Experimental Biology and Medicine*. 2009;148(1):57-9.
196. Giuliani C, Noguchi Y, Harii N, Napolitano G, Tatone D, Bucci I, et al. The flavonoid quercetin regulates growth and gene expression in rat FRTL-5 thyroid cells. *Endocrinology*. 2007;149(1):84-92.
197. Phan T, Yu XM, Kunnimalaiyaan M, Chen H. Antiproliferative effect of chrysin on anaplastic thyroid cancer. *Journal of Surgical Research*. 2011;170(1):84-8.
198. Qin Y, Li Z, Chen Y, Hui H, Sun Y, Yang H, et al. III-10, a newly synthesized flavonoid, induced differentiation of human U937 leukemia cells via PKC $\delta$  activation. *European Journal of Pharmaceutical Sciences*. 2012;45(5):648-56.
199. Batra P, Sharma AK. Anti-cancer potential of flavonoids: recent trends and future perspectives. *3 Biotech*. 2013;3(6):439-59.
200. George VC, Dellaire G, Rupasinghe HV. Plant flavonoids in cancer chemoprevention: role in genome stability. *The Journal of Nutritional Biochemistry*. 2017;45:1-4.
201. Yeh CC, Yang JI, Lee JC, Tseng CN, Chan YC, Hseu YC, et al. Anti-proliferative effect of methanolic extract of *Gracilaria tenuistipitata* on oral cancer cells involves apoptosis, DNA damage and oxidative stress. *BMC Complementary and Alternative Medicine*. 2012;12:1-9.
202. Cao Y, Cao R. Angiogenesis inhibited by drinking tea. *Nature*. 1999;398(6726):381.
203. Mojzic J, Varinska L, Mojziso G, Kostova I, Mirossay L. Antiangiogenic effects of flavonoids and chalcones. *Pharmacological Research*. 2008;57(4):259-65.
204. Ravishankar D, Rajora AK, Greco F, Osborn HM. Flavonoids as prospective compounds for anti-cancer therapy. *The International Journal of Biochemistry and Cell Biology*. 2013;45(12):2821-31.

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