

Seasonal Changes of Phagocytic Activity Enhanced by an Acoholic Mint Extract in Cultured Rainbow Trout

Spînu Marina, Pall Eموke, Niculae Mihaela, Vasiu Aurel, Brudaşcă Gheorghe Florinel, Cerbu Constantin Gheorghe, Negrutiu Vlad, Vasiu Constantin, Silvana Popescu, Sandru Carmen Dana

Department of Clinical Sciences - Infectious diseases, University of Agricultural Sciences and Veterinary Medicine, Str. Manastur no.3-5, Cluj-Napoca, ROMANIA.

ABSTRACT

Objective: We hypothesized that leukocyte numbers and their *in vitro* phagocytic activity in farmed rainbow trout are subject to negative influence of extreme temperatures for fish and that mint extract could alleviate these effects. **Materials and Methods:** Blood samples from farmed rainbow trout collected during winter and summer were subjected to leukocyte subpopulation counts (%) and carbon particle inclusion test to estimate the *in vitro* phagocytic activity after 15 and 30 min of incubation at 4 and 22°C, with or without alcoholic mint extract (In). **Results and Discussion:** There was a significant increase in lymphocytes ($p < 0.05$) and monocytes ($p < 0.01$) in summer versus winter, while the heterophils were dominant during winter ($p < 0.01$). *In vitro* spontaneous phagocytosis increased with the seasonal increase in temperature (0.425 ± 0.094 , winter and 0.835 ± 0.102 , summer, $p < 0.05$) and during the first reading period (0 to 15 min, $p < 0.01$). The mint extract increased phagocytosis for the overall reading period from 2x (winter, 0.281 ± 0.221) up to 5x (summer, 1.032 ± 0.221) ($p < 0.01$). **Conclusion:** The positive activity of the alcoholic mint extract during phagocytosis, more intense during summer than during winter, proved to be a temperature driven process, plant active principles playing a dual stimulating role for both monocytes and heterophiles.

Key words: Rainbow trout, Season, Leukocytes, Phagocytosis, Mint extract.

INTRODUCTION

Fish are some of the most primitive vertebrates and for the development of the immune system represent an important link between invertebrates and evolved vertebrates.¹ In phylogeny, the fish are the first to show both innate and adaptive cellular and humoral responses, capable to respond to microorganisms, toxins or malignant cells. These two systems work together to destroy invaders or to trigger defence processes.²⁻⁴ Fish non-specific defence mechanisms are like those of other vertebrates, the phagocytic activity being fulfilled by heterophiles and macrophages.⁵ The innate immune response is defined by lack of specificity towards antigens, thus being independent

on previous contacts with those. Various non-self-particles can trigger the innate cellular and molecular effectors as the first line of defence in very short time scale.⁶⁻⁷ The anatomical and functional integrity of the immunological defence mechanisms are extremely important and depend on the dynamics of host-aggressor interrelations.⁸ The magnitude of the fish immune response is influenced by their habitat and its intrinsic factors, but also by their poikilothermic condition.⁵ Immunostimulants such as bacterial derivatives, animal and plant extracts increase resistance to infectious disease by enhancing non-specific immune defence mechanisms.⁹

Submission Date: 30-08-2017;

Revision Date: 17-11-2017;

Accepted Date: 23-11-2017

DOI: 10.5530/ijper.52.4s.80

Correspondence:

Marina Spinu,
University of Agricultural Sciences and Veterinary Medicine, Faculty of Veterinary Medicine, Str. Manastur 3-5, Cluj-Napoca, ROMANIA.
Phone: +40264596384
E-mail: marina.spinu@gmail.com



www.ijper.org

Meanwhile, numerous medicinal plants or their parts, single or in combinations, as traditional preparations or commercial products have been tested in diverse aquatic animals.¹⁰

Natural stimulating compounds are sought after by immunologists as cheap and readily available alternatives for different species of farmed animals, including fish, therefore medicinal plants could represent an appropriate source. The aim of the study was to investigate seasonal variation of phagocytosis as an indicator of innate immunity and the *in vitro* potential of an alcoholic mint (*Menta piperita*) extract in enhancing phagocytosis in cultured rainbow trout (*Oncorhynchus mykiss*).

MATERIAL AND METHODS

Blood was sampled on heparine (50 IU/ml) by caudal vein puncture from conventionally farmed rainbow trout during winter and summer seasons and subjected to leukocyte subpopulation counts (Dia-Quick Panoptic method, CliniLab, Romania) and carbon particle inclusion test¹¹ to estimate the *in vitro* phagocytic activity after 15 and 30 min of incubation both at 4 and 22°C. The phagocytosis variants were represented by 0.5 ml of blood either un-supplemented (control), or added with 20 µl of 70° alcohol (solvent control) or 20 µl of an alcoholic mint extract (experimental variant), respectively. Phagocytic cells engulf inert particles such as carbon due to the defensive capacity of these cells. Two microliters of supernatant of India ink, obtained by centrifugation at 6000 rpm for 45 min were added to each variant. 150 µl of the mixture were transferred immediately to 2 ml of saline, and the rest was incubated for a total of 30 min. Similar aliquots of each variant were removed to 2 ml of saline every 15 min. The final tubes, containing saline, India ink and blood mixtures were centrifuged at 800 rpm and the supernatants were read spectrophotometrically at a wavelength of 535 nm; d=1 cm (SUMAL PE2, Carl Zeiss, Jena). There was a decrease in absorbance with time, as carbon was phagocytized. The phagocytic activity index was read in optical density units (ODU) and then expressed as the difference between the natural logarithms (ln) of the optical densities of the phagocytosis at 0-15 min, 15-30 and 0-30 min divided by time. Student's t-test was used to evaluate the statistical significance of the differences between the seasons and variants.

RESULTS AND DISCUSSION

In fish, nonspecific immunity is the first line of defence and is a considerable part of the immune response. Certain parameters of innate immune response, such as intensive respiratory burst, lysozyme activity, complement

concentration and total IgM, are associated with disease resistance.¹² Non-specific immunity, similar to that of evolved vertebrates, depends on the activity of monocytes/macrophages, melanoma-macrophage centers, granulocytes and platelets.¹³ Research in teleost fish described host defence strategies that involved intracellular (nitric oxide, phagolysosome fusion) and extracellular (heterophile degranulation and extracellular traps) that play an important role in resistance to disease.¹⁴ The most important cells involved in non-specific defence are the phagocytes.⁶⁻⁷ Macrophages and neutrophil granulocytes are the two types of cells with significant phagocytic capacity in rainbow trout.¹⁵ They are the first cells to encounter invading microorganisms and therefore play a very important role in the initial stage of infection.²

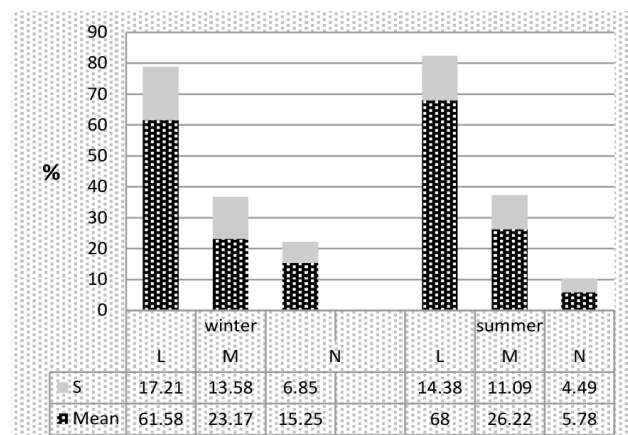


Figure 1: Leukocyte subpopulations in the two group of trout (winter and summer samplings).

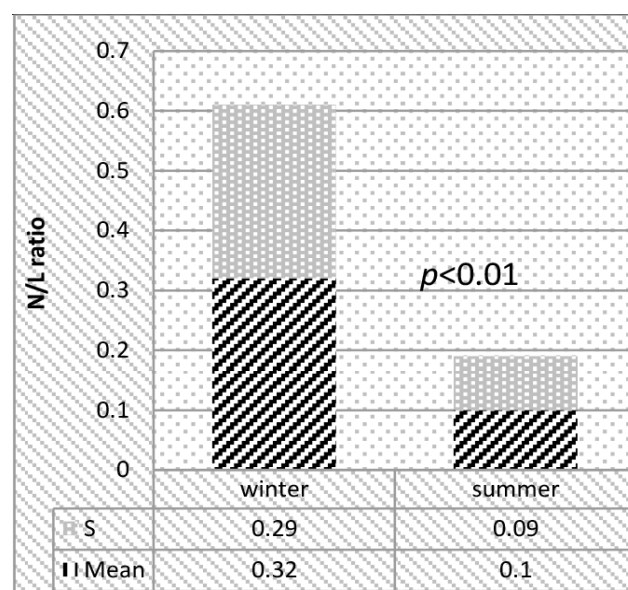


Figure 2: N/L ratio as a stress indicator: a more pronounced stress level was recorded during winter.

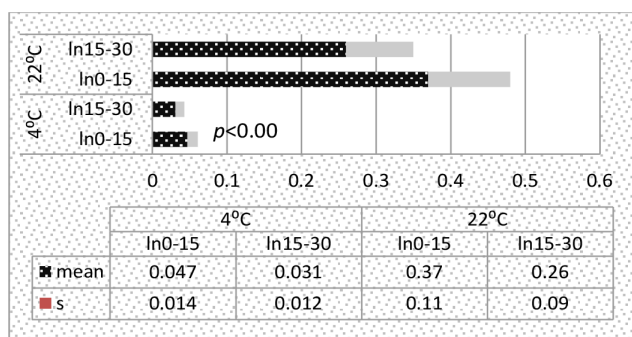


Figure 3: *In vitro* phagocytosis in farmed rainbow trout as influenced by the incubation temperature (x±s).

It has been reported that ambient temperature influences both antibody production and resistance to infection in cold blooded animals.¹⁶

The results obtained in this study indicated an increase of the lymphocyte and monocytes levels in summer versus winter, while the heterophils were dominant during the winter (Figure 1). Both the seasonal variance of the environmental temperature (Figure 2, and 3) and the incubation temperature during *in vitro* evaluation of phagocytosis statistically significantly influenced the

Table 1: Phagocytic activity in farmed rainbow trout during the winter season, under alcoholic mint extract influence (x±s).

Winter	Variants								
	Control			Alcohol			Mint extract		
	In0-In15	In15-In30	In0-In30	In0-In15	In15-In30	In0-In30	In0-In15	In15-In30	In0-In30
Mean	0.425	0.117	0.445	0.014	0.049	0.025	0.174	0.107	0.281
S	0.304	0.067	0.120	0.009	0.033	0.015	0.121	0.051	0.081

Table 2: Phagocytic activity in summer season in farmed rainbow trout. Alcoholic mint extract influence *in vitro* (x±s).

Summer	Variants								
	Control			Alcohol			Mint extract		
	In0-In15	In15-In30	In0-In30	In0-In15	In15-In30	In0-In30	In0-In15	In15-In30	In0-In30
Mean	0.835	0.261	0.574	0.721	0.272	0.993	0.804	0.227	1.032
S	0.994	0.767	0.460	0.321	0.543	0.475	0.431	0.521	0.221

N/L stress index, higher in winter than during summer and lowered the phagocytosis at 4°C.

Phagocytic activity is an important immunological parameter to define in investigating the bacterial infections in fish, since it is related to the nutritional status, age and breed of the animal, as well as prevention and therapies. Vegetal extracts can exert stimulating effects on immunocompetent cells.⁷ Under the conditions of the experiment, the extract proved to be stimulating, suggesting the possibility of its use in improving immunity in farmed rainbow trout (Table 1 and 2).

The alcoholic mint extract improved phagocytosis from 2x (winter, 0.281±0.081) up to 5x (summer, 1.032±0.221) for the overall reading period (0 to 30 min) (p<0.001).

CONCLUSION

The changes in the leukocyte numbers differed depending on the temperature of the season and subpopulation. The phagocytosis stimulating activity of the alcoholic mint extract proved to be a temperature driven process

due to the active principles from mint, which probably stimulated both the monocytes and the heterophiles, more intensely during summer than during winter.

ACKNOWLEDGMENT

We acknowledge the partial financial support provided by the Ministry of Education (UEFISCDI) grant no PN II 61/2012 Delatbiotox.

CONFLICT OF INTEREST

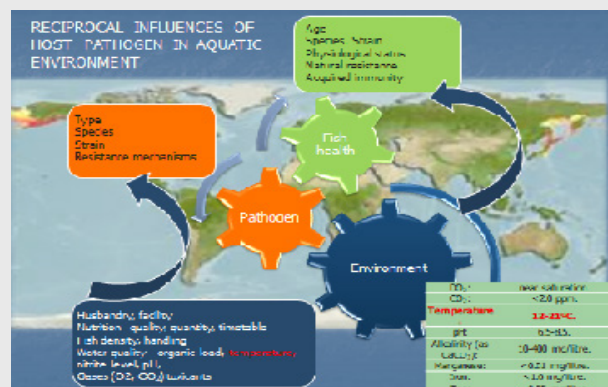
The authors certify that they have no affiliations with or involvement in any organization or entity with any financial interest, or non-financial interest in the subject matter or materials discussed in this manuscript.

REFERENCES

1. Zhu LY, Nie L, Zhu G, Xiang LX, Shao JZ. Advances in research of fish immune-relevant genes: A comparative overview of innate and adaptive immunity. *Developmental and Comparative Immunology*. 2013;39(1-2):39-62.

- Bayne CJ, Gerwick L. The acute phase response and innate immunity of fish. *Dev Comp Immunol.* 2001;25(8):725-43.
- Ellis AE. Immunity to bacteria in fish. *Fish Shellfish Immunol.* 1999;9(4):291-308.
- Magnadottir B, Audunsdottir SS, Bragason BTH, Gisladdottir B, Jonsson ZO, Gudmundsdottir S. The acute phase response of Atlantic cod (*Gadus morhua*): Humoral and cellular responses. *Fish Shellfish Immunol.* 2011;30(4):1124-30.
- Tort L, Balasch JC, Mackenzie S. Fish immune system. A crossroads between innate and adaptive responses. *Immunologia.* 2003;22(3):277-86.
- Magnadottir B. Innate immunity of fish (overview). *Fish and Shellfish Immunology.* 2006;20(2):137-51.
- Uribe C, Folch H, Enriquez R, Moran G. Innate and adaptive immunity in teleost fish: a review. *Veterinari Medicina.* 2011;56(10):486-503.
- Cerbu CG, Brudașcă GF, Giupană RM, Guranda S, Niculae M, Paștiu AI, et al. Theoretical approach of the immune system of Danube huchen (*Hucho hucho*). *Lucrări Științifice Medicină Veterinară.* 2015;XVIII Timișoara.2015;2:49-54.
- Kum C, Sekkin S. The Immune System Drugs in Fish: Immune Function, Immunoassay, Drugs. *Recent Advances in Fish Farms, Dr. Faruk Aral (Ed.), InTech.* 2011;11:169-216. DOI: 10.5772/26869.
- Hai NV. The use of medicinal plants as immune stimulants in aquaculture: A review. *Aquaculture.* 2015;446:88-96.
- Spinu M, Degen A. Effect of cold stress on performance and immune responses of Bedouin and White Leghorn hens. *British Poultry Science.* 1993;34(1):177-85
- Wiegertjes GF, Stet RJ, Parmentier HK, van Muiswinkel WB. Immunogenetics of disease resistance in fish: a comparative approach. *Dev Comp Immunol.* 1996;20(6):365-81.
- Adeyemo OK, Agbede SA, Olaniyan AO, Shoaga OA. The haematological response of clarias gariepinus to changes in acclimation temperature. *African Journal of Biomedical Research.* 2003;6(2):105-8.
- Rieger AM, Barreda DR. Antimicrobial mechanisms of fish leukocytes. *Developmental and Comparative Immunology.* 2011;35(12):1238-45.
- Afonso A, Lousada S, Silva J, Ellis AE, Silva MT. Neutrophil and macrophage responses to inflammation in the peritoneal cavity of rainbow trout *Oncorhynchus mykiss*. A light and electron microscopic cytochemical study. *Dis Aquat Organ.* 1998;34(1):27-37.
- Bowden TJ. Modulation of the immune system of fish by their environment. *Fish and Shellfish Immunology.* 2008;25(4):373-83.

PICTORIAL ABSTRACT



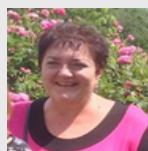
SUMMARY

- This study investigated the effect of seasonal variation of environmental temperature on numbers and phagocytic function of leucocytes and the potential modulating influence of an alcoholic mint extract on those
- Leukocyte subpopulations were counted and N/L ratios calculated, while a carbon particle inclusion test was used to evaluate the phagocytosis.
- Lymphocyte subpopulations reacted differently to seasonal changes of the environmental temperatures.
- The active principles from mint probably stimulated both the monocytes and the heterophiles more intensely during summer than during winter, thus enhancing phagocytosis.

About Authors



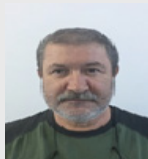
Marina Spinu, DVM, PhD: currently as Professor of Infectious diseases, Head of Department of Clinical Sciences, USAMV, Faculty of Veterinary Medicine, Cluj-Napoca, Romania. Areas of interest: immunology, infectious diseases, antibiotic resistance, alternative therapies (over 500 national and international publications). Under her guidance 17 scholars have been awarded with their PhD degree.



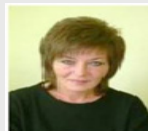
Carmen Dana Sandru, DVM, PhD: currently Associate Professor of Preventive medicine and Veterinary Legislation and ethics, at the USAMV, Faculty of Veterinary Medicine, Cluj-Napoca, Romania. Areas of interest: infectious diseases, preventive medicine, alternative therapies, environment health (over 250 national and international publications).



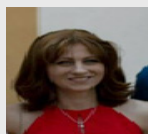
Constantin VasIU, DVM, PhD: currently Infectious diseases, Head of Department of Clinical Sciences, USAMV, Faculty of Veterinary Medicine, Cluj-Napoca, Romania. Areas of interest: infectious diseases, preventive medicine, alternative therapies, environment health (over 250 national and international publications).



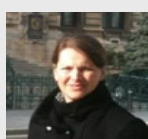
Florinel Brudasca, DVM, PhD: currently as Professor of Infectious diseases, USAMV, Faculty of Veterinary Medicine, Cluj-Napoca, Romania. Areas of interest: game and exotic animals pathology, infectious diseases, immune stimulation, vaccinology (over 250 national and international publications). Under his guidance 2 scholars have been awarded with their PhD degree, and another one submitted his thesis.



Silvana Popescu, DVM, PhD: Currently Associate Professor of Animal Hygiene at the USAMV, Faculty of Veterinary Medicine, Cluj-Napoca, Romania. Areas of interest: environment hygiene, environment health, preventive medicine, animal welfare, alternative therapies, (over 180 national and international publications).



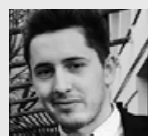
Mihaela Niculae, DVM, PhD: Currently Lecturer in Infectious diseases, at the USAMV, Faculty of Veterinary Medicine, Cluj-Napoca, Romania. Areas of interest: infectious diseases, alternative therapies, microbiology, environment health (over 170 national and international publications).



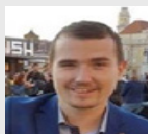
Emoke Pall, DVM, PhD: Currently researcher with 11 years of experience, at the USAMV, Faculty of Veterinary Medicine, Cluj-Napoca, Romania. Areas of interest: infectious diseases, cell cultures, stem cells, microbiology/PCR (over 170 national and international publications).



Aurel VasIU, DVM, PhD: Currently Lecturer in Infectious diseases, at the USAMV, Faculty of Veterinary Medicine, Cluj-Napoca, Romania. Areas of interest: infectious diseases, farmed animal pathology, microbiology (over 100 national and international publications).



Constantin Cerbu, DVM, PhD Student: currently Assistant with 3 years of teaching experience in Infectious diseases, at the USAMV, Faculty of Veterinary Medicine, Cluj-Napoca, Romania. Areas of interest: infectious diseases, microbiology of the environment, farm animal/fish pathology.



Vlad Negrutiu, DVM: Currently PhD student at the USAMV, Faculty of Veterinary Medicine, Cluj-Napoca, Romania. Areas of interest: infectious diseases, microbiology, fish pathology, alternative therapies.

Cite this article: Spinu M, Pall E, Niculae M, VasIU A, Brudasca GF, Cerbu CG, *et al.* Seasonal Changes of Phagocytic Activity Enhanced by an Acoholic Mint Extract in Cultured Rainbow Trout. Indian J of Pharmaceutical Education and Research. 2018;52(4S):S86-S90.