# Protective Effects of *Ficus benghalensis* in Streptozotocin (STZ) Induced Diabetic Zebrafish (*Danio rerio*) Model

## Essa M Sabi<sup>1</sup>, Ahmed H Mujamammi<sup>1</sup>, Ziyad M Althafar<sup>2</sup>, Samia T Al-Shouli<sup>3</sup>, Lotfi S Bin Dahman<sup>4</sup>, Khalid M Sumaily<sup>1,\*</sup>

<sup>1</sup>Clinical Biochemistry Unit, Department of Pathology, College of Medicine, King Saud University, Riyadh, SAUDI ARABIA. <sup>2</sup>Department of Medical Laboratories Sciences, College of Applied Medical Sciences in Alquwayiyah, Shaqra University, Riyadh, SAUDI ARABIA.

<sup>3</sup>Immunology Unit, Pathology Department, College of Medicine, King Saud University, Riyadh, SAUDI ARABIA. <sup>4</sup>Department of Medical Biochemistry, College of Medicine and Health Sciences, Hadhramout University, Mukalla, YEMEN.

## ABSTRACT

Background: Diabetes Mellitus (DM) is a chronic metabolic disorder that is characterized by hyperglycemia. Severe complications such as cardiovascular disease, neuropathy, nephropathy, and retinopathy are associated with DM. Objectives: To explore a safer treatment option with no/less adverse effects, we evaluated the potential of Ficus benghalensis to alleviate certain diabetic conditions. Materials and Methods: Administering streptozotocin to zebrafish intraperitoneally induced diabetes. After induction, Ficus benghalensis hydroethanolic bark extract was administered to zebrafish in different concentrations (5, 10, and 15  $\mu$ g/mL). Biochemical blood parameters such as Blood Glucose, total cholesterol levels, triglycerides, and liver enzymes such as alkaline phosphatase, alanine aminotransferase, and aspartate aminotransferase were analyzed. Results: Treating the streptozotocin-induced group with Ficus benghalensis hydroethanolic bark had a hypoglycemic effect and reduced the cholesterol, triglycerides, and liver enzymes. The histopathological evaluation also showed improvement in the damaged pancreatic tissues. Conclusion: Our results suggest that Ficus benghalensis hydroethanolic bark extract can be used as a potent therapeutic candidate for treating diabetes.

**Keywords:** Diabetes mellitus, *Ficus benghalensis*, Alkaline phosphatase, Alanine aminotransferase, Aspartate aminotransferase, Zebrafish.

#### INTRODUCTION

Diabetes mellitus (DM) is an endocrine disorder that is a widespread public health problem. In DM, either insulin production by pancreatic beta cells isn't sufficient (Type-1 DM) or the produced insulin by peripheral tissues is defective (Type-2 DM) resulting in glucose elevation in the blood that interferes with macronutrient metabolism.<sup>1</sup> The main cause of mortality and morbidity in diabetic patients are vascular complications, which include micro and macrovascular systems (Cardiovascular system, diabetic neuropathy and retinopathy, and diabetic kidney disease).<sup>2</sup> The prevalence of DM is increasing and it's estimated that more than 10.5% adult population has this condition.<sup>3</sup> The most common treatment options that are currently available include insulin and oral antidiabetic drugs like biguanides, glinides, and sulfonylureas. These drugs have limitations due to their adverse effects,<sup>4-5</sup> the spike in the prevalence and the adverse effects of the existing drugs have resulted in the ongoing pursuit of drugs that will allow for more effective remedies. As many plants have vast phytochemicals of therapeutic importance, they can be immensely useful in treating various disorders. Many traditional plants have been studied and have been reported Submission Date: 17-04-2022; Revision Date: 20-05-2022; Accepted Date: 04-06-2022.

DOI: 10.5530/ijper.56.3.134 Correspondence: Dr. Khalid M Sumaily Clinical Biochemistry Unit, Department of Pathology, College of Medicine, King Saud University, Riyadh-11461, SAUDI ARABIA. E-mail: ksumaily@ksu. edu.sa



for better anti-diabetic potential with no adverse effects.<sup>6</sup> *Ficus benghalensis*, commonly known as the banyan tree or Figure is from the Moraceae family and it is a large tree high as 20-30 m.<sup>7</sup> Different parts of this plant such as aerial roots, leaf, bark, and fruit are well documented to have different therapeutic properties such as antioxidant, antimicrobial, anti-inflammatory, antimutagenic, anticancer, and antistress.<sup>8-10</sup> Its rich source of phytochemical constituents contribute to the properties mentioned above. Because of its various pharmacological properties, we designed the present experiment to determine the anti-diabetic activity of ethanolic *Ficus benghalensis* bark extract in *Danio rerio*.

### **MATERIALS AND METHODS**

### **Plant Extract Preparation**

Powdered bark of *Ficus benghalensis* was weighed and 30 g was mixed with 70% ethanol for 24 hr with occasional stirring. After stirring, the hydroethanolic solvent with the bark extract was filtered using a filter paper (Whatman - 125 mm). The ethanolic filtrate was dried at 40°C and the obtained powder was stored and used for further experiments.

#### Maintenance of Adult Danio rerio

Adult zebrafish with body size 2.0-3.0 cm weighing 250-300 mg were selected and used. These fishes were accustomed for 10 days in a 30 L tank of dechlorinated water with continuous aeration at  $25\pm2^{\circ}$ C. The fishes were maintained with a 14hr light/10hr dark photoperiod with the pH of dechlorinated water ranging from 7.0 - 7.4. Fishes were given commercial food twice a day.

## Toxicity test of *Ficus benghalensis* Hydroethanolic Bark Extract

A toxicity test of *Ficus benghalensis* hydroethanolic bark (FBHB) extract was carried out based on the recommendations of OECD guidelines. 55 fishes were randomly divided into 11 batches; each batch had 5 fishes in 5 L tanks. The first batch was the control batch which did not have any extract and the following 10 batches had various concentrations ranging from 10  $\mu$ g/mL to 100  $\mu$ g/mL FBHB extract. The testing period was for a week and during this time, the fishes were monitored regularly to analyze their swimming pattern, gills movements, and the presence of deaths.

# Streptozotocin-induced Diabetes and Experimental Design

5 Groups of disease-free zebrafish (n = 10) were placed in 5 L tanks. FBHB extracts were given in concentrations 5, 10 and 15 µg/mL. Streptozotocin (STZ) was utilized to induce diabetes. The rapid cooling ice-cold method was employed to anesthetize and then 10  $\mu$ l of streptozotocin was intraperitoneally (i.p.) administered using an insulin syringe. After administrating STZ, the fishes were kept in 2% sucrose for 24 hr, diabetes was induced and the fishes were transferred to the respective groups (Group 02 - 05) for the experiment

The treatment period was 7 days during which the fish were treated daily and after that, the zebrafish were euthanized and used for analysis. The experimental groups were as follows:

- Group 01: Control (Normal glucose level).
- Group 02: Induced group, Intraperitoneal administration of STZ.
- Group 03: Treated group 1, STZ-induced diabetes
  + 5 μg/mL of FBHB extract
- Group 04: Treated group 2, STZ-induced diabetes
  + 10 μg/mL of FBHB extract
- Group 05: Treated group 3, STZ-induced diabetes + 15 μg/mL of FBHB extract

#### **Estimation of Biochemical Blood Parameters**

Parameters such as Blood Glucose, total cholesterol levels, and triglycerides levels were analyzed. The blood glucose levels were measured to confirm diabetes induction. Fishes' blood was collected and measured using a OneTouch Select® commercially available glucometer. Total cholesterol and triglycerides were measured using an autoanalyzer.

#### Liver Homogenate Preparation

The fishes were dissected and the liver was isolated. The isolated liver was kept in ice-cold sucrose solution (0.25 M) and homogenized using a micropestle. The homogenized tissues were centrifuged at 7500 rpm at 4°C for 20 min. The supernatant was transferred to a new tube and used for analyzing liver enzymes such as alkaline phosphatase, alanine aminotransferase, and aspartate aminotransferase.

## Alkaline Phosphatase (ALP) Assay

ALP assay was performed by the method reported by Vaidyanathan *et al.*<sup>11</sup> Briefly, 1.5 ml of carbonatebicarbonate buffer along with 200  $\mu$ l magnesium chloride and 1.5 ml disodium phenyl phosphate were mixed and incubated at 37°C for 5 min. 100  $\mu$ l of the liver homogenate was added to the mixture and incubated for 20 min at 37°C. 1 ml trichloroacetic acid (10%) was added and the mixture was centrifuged at 5000 rpm for 10 min. 1 ml of Folin's phenol reagent, 2 ml of supernatant, and 2 ml of sodium carbonate (15%) were added and mixed. A blue color was formed and it was measured spectrophotometrically at 620 nm. ALP activity was expressed in  $\mu mol$  of phenol liberated/ mg of protein/min.

#### Alanine Aminotransferase (ALT) Assay

ALT assay was performed by following the protocol mentioned by Reitman and Franke *et al.*<sup>12</sup> Briefly, 200 µl of liver homogenate was added to 1 ml buffered substrate (a-Ketoglutarate (2 mM) and dl-alanine (200 mM)) and incubated for 1 hr at 37°C. After incubation, 1 ml of 2,4 -Dinitro Phenyl Hydrazine reagent was added to the mixture and incubated for 20 min at room temperature. 10 ml of NaOH (0.4 N) was added and kept for 5 min to stop the reaction. The absorbance was measured spectrophotometrically at 540 nm. ALT activity was expressed in µmoles of pyruvate liberated/hr/mg protein.

#### Aspartate Aminotransferase (AST) Assay

ALT assay was performed by following the protocol mentioned by Reitman and Franke *et al.*<sup>12</sup> Briefly, 200 µl of liver homogenate was added to 1 ml buffered substrate (a-Ketoglutarate (2 mM) and dl-aspartate, (200 mM)) and incubated for 1 hr at 37°C. After incubation, 1 ml of 2,4 -Dinitro Phenyl Hydrazine reagent was added to the mixture and incubated for 20 min at room temperature. 10 ml of NaOH (0.4 N) was added and kept for 5 min to stop the reaction. The absorbance was measured spectrophotometrically at 540 nm. ALT activity was expressed in µmoles of pyruvate liberated/ hr/mg protein.

#### **Statistical Analysis**

The experiments performed in the work were repeated in triplicates and the results were reported as mean  $\pm$  SD. A *P-value* less than 0.001 was considered to be significant. Statistical analysis (one-way ANOVA -Tukey multiple comparison tests) and the graphs were done using GraphPad Prism 5 software.

## RESULTS

## **Toxicity Test**

*Ficus benghalensis* hydroethanolic bark extract was tested and no toxic effects were observed in all the 10 concentrations given. When compared to the control group, the tested groups showed no observable effects. The toxicity test period was a week and the survival rate of adult zebrafish was 100% in FBHB extract. From our results, we conclude that the LC<sub>50</sub> for FBHB extract is greater than 100mg/L.

The serum glucose levels of STZ-induced zebrafish were 149.67  $\pm$  0.47 mg/dL which is comparatively higher than the control group (125  $\pm$  0.82 mg/dL). However, on treatment with FBHB extracts the glucose levels of zebrafishes showed a dose-dependent reduction with the mean values of 113.67  $\pm$  0.94, 67  $\pm$  0.82, and 62.33  $\pm$  0.94 mg/dL for doses 5, 10, and 15 µg/mL as shown in Figure 1.

## **Total Cholesterol Levels**

The fish in the control group had a cholesterol level of  $39.17 \pm 0.62 \text{ mg/dL}$ . Intraperitoneal administration of STZ caused a significant elevation of cholesterol levels to  $81.8 \pm 0.71 \text{ mg/dL}$ . But, on treating the induced groups with FBHB extracts suppressed cholesterol levels to  $56.43 \pm 0.42$ ,  $54.27 \pm 0.29$ , and  $46.23 \pm 0.76 \text{ mg/dL}$  for doses 5, 10, and 15 µg/mL in a dose-dependent fashion and it is depicted in Figure 2.

#### **Blood Triglycerides Levels**

The FBHB extracts significantly alleviated triglycerides levels in the treated groups compared to the induced group. The triglyceride levels of the induced group was  $158.67 \pm 0.47 \text{ mg/dL}$ , whereas in the control group it was  $136.33 \pm 0.47 \text{ mg/dL}$  and at doses 5, 10, and 15 µg/mL it was  $106.33 \pm 0.94$ ,  $96.5 \pm 0.41$ , and  $88.83 \pm 0.57 \text{ mg/dL}$ . (Figure 3).

## **Alkaline Phosphatase Activity**

ALP activity in the liver homogenate of the fishes in the induced group was  $66.17 \pm 1.84 \,\mu\text{mol/mg}$  protein/min which was significantly high compared to the control group (15.43  $\pm$  0.37  $\mu$ mol/mg protein/min). A dose-dependent suppression was seen when the induced

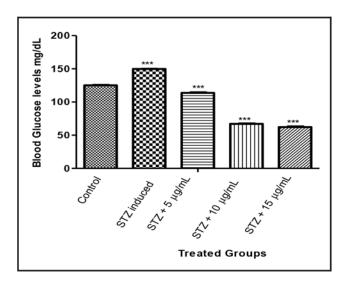


Figure 1: Effect of *Ficus benghalensis* hydroalcoholic bark extract on blood glucose level.

## **Blood Glucose Levels**

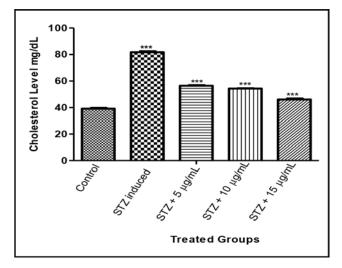


Figure 2: The changes in total cholesterol levels in the induced and *Ficus benghalensis* hydroalcoholic bark extract treated groups compared to the control group.

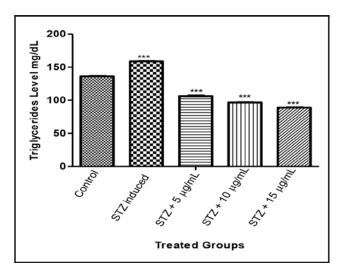


Figure 3: Effect of *Ficus benghalensis* hydroalcoholic bark extract on blood triglycerides level.

groups were supplemented with the FBHB extracts. The ALP activity were  $53.23 \pm 0.29$ ,  $46.97 \pm 0.12$ , and  $44.57 \pm 0.37 \,\mu\text{mol/mg}$  protein/min for doses 5, 10, and  $15 \,\mu\text{g/mL}$  respectively. (Figure 4).

#### Alanine Aminotransferase Activity

STZ significantly elevated ALT activity (Control group: 280  $\pm$  0.82; Induced group: 434  $\pm$  0.82 µmol/hr/ mg protein). Treating the induced group with *Ficus* benghalensis hydroethanolic bark extract had a significant reduction in the ALT activity with mean values of 319.33  $\pm$  0.47, 314.67  $\pm$  0.47, and 196.5  $\pm$  0.41 µmol/hr/mg protein for doses 5, 10, and 15 µg/mL respectively as illustrated in Figure 5.

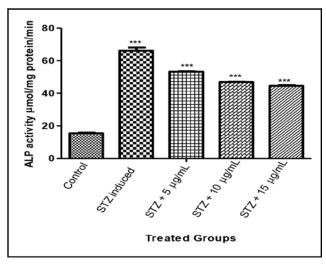


Figure 4: Effect of *Ficus benghalensis* hydroalcoholic bark extract on alkaline phosphatase (ALP).

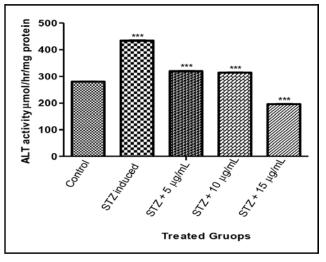


Figure 5: Effect of *Ficus benghalensis* hydroalcoholic bark extract on alanine aminotransferase (ALT).

#### Aspartate aminotransferase activity

The fishes in the induced group received STZ which caused a strong elevation in AST activity (119.53  $\pm$  0.45  $\mu$ mol/hr/mg protein) compared to the control group (93.73  $\pm$  0.49  $\mu$ mol/hr/mg protein). When treated with FBHB extract, the AST activity was significantly reduced. The AST activity was 107.07  $\pm$  0.49, 84.7  $\pm$  0.36, and 40.1  $\pm$  0.65  $\mu$ mol/hr/mg protein for doses 5, 10, and 15  $\mu$ g/mL respectively (Figure 6).

#### **Histopathological Evaluation of Pancrease**

The section of the pancreas from the zebrafish - control group showed normal histological features with a preserved architecture and intact epithelium (Figure 7A). STZ-induced groups had a severe loss in the structure and there is abrasion of epithelium (Figure 7B). A

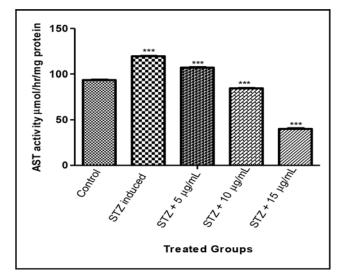
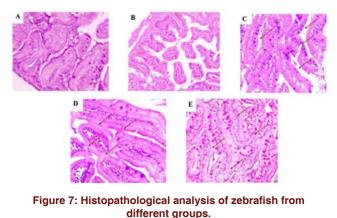


Figure 6: Effect of *Ficus benghalensis* hydroalcoholic bark extract on aspartate aminotransferase (AST).



A – Control group (Normal glucose levels), B – STZ-induced group, C – Induced group treated with 5 µg/mL FBHB extract, D - Induced group treated with 10 µg/mL FBHB extract, E - Induced group treated with 15 µg/mL FBHB extract.

gradual restorement of the pancreatic structure is seen as the dosage of FBHB increases. Infiltration of immune cells can be clearly visualized in the induced groups and a reduction of infiltrated immune cells is seen in higher doses of FBHB. Attenuation of the pancreatic tissue injury is visualized in the highest FBHB extract dose - $15 \mu g/mL$ . (Figure 7 C-E).

#### DISCUSSION

Several drugs that are used to treat diabetes have adverse effects. Plant-based drugs are widely preferred and they tend to have no or fewer adverse effect than synthetic drugs.<sup>13</sup> The major objective of the current study was to evaluate the remedial effects of *Ficus benghalensis* hydroethanolic bark extract against diabetes mellitus and some of the complications surrounding it. We preferred Zebrafish as it is a classic experimental model to study metabolic disorders due to the functional similarity in pancreas structure, lipid metabolism, glucose homeostasis, and adipose biology.14 Streptozotocin (STZ), a diabetogenic drug was our choice to induce diabetes as it is effective in the ablation of pancreatic  $\beta$ -cell that eventually leads to elevated blood glucose levels and suppresses insulin levels.<sup>15-16</sup> Intraperitoneal administration of STZ caused a spike in the blood glucose levels indicating diabetic conditions were induced. After treating the induced group with FBHB extract, hypoglycemic effects were observed. This observation may be because of better peripheral glucose uptake, restricted production of glucose by the liver, or the ability of the extract to restore the damaged pancreatic  $\beta$ -cell that causes the production of insulin. Dyslipidemia is characterized by increased cholesterol levels in the blood. Insulin resistance plays a critical role in the development of diabetic dyslipidemia.<sup>17</sup> This increase in cholesterol levels is harmful to diabetic patients as they are at risk of cardiovascular disease.<sup>18</sup> Cholesterol is vital for the proper functioning and survival of  $\beta$ -cells. If cholesterol irregularly accumulates in β-cells because of upregulated LDL-R expression, it can lead to cell dysfunction, the onset of many conditions such as insulin resistance.19-20 Therefore suppressing cholesterol levels may reverse insulin resistance and control the development of diabetes. Previous studies conducted with Ficus benghalensis suggest that the cholesterol-lowering ability may be due to the presence of beta-glucan, a soluble polysaccharide.<sup>21</sup> Similar to cholesterol, hypertriglyceridemia manifests itself as patients develop resistance to insulin. The risk of cardiovascular disease in Diabetic patients with hypertriglyceridemia is estimated to be 2-10 fold higher than the patients without diabetes.<sup>22-23</sup> Higher levels can lead to triglyceride-induced pancreatitis which will lead to acidosis and hyperglycemia.<sup>24</sup> Moderate elevation of triglycerides is a risk factor that is modifiable and triglycerides are a growing target in diabetes care.25 Hypotriglyceridemic and hypocholesterolemic effects can be seen when treating the induced groups with Ficus benghalensis hydroethanolic bark extract. This may be attributed to the induction of lipoprotein lipase enzymes. Alkaline phosphatase is widely used as a clinical marker to diagnose bone or hepatic disease.<sup>26</sup> Several studies show elevated ALP levels in diabetic patients.<sup>27</sup> ALP is a key contribution to vascular calcification and endothelial dysfunction which is linked to insulin resistance eventually leading to the development of diabetes.<sup>28</sup> ALP causes glomerular hyperfiltration and plays a role in the progression of diabetic complications such as diabetic nephropathy and end-stage renal

disease.<sup>29-30</sup> The ALP activity was reduced mildly at the highest dose suggesting that Ficus benghalensis bark extract at a much higher dose might greatly reduce the ALP levels. Liver dysfunction is one of the factors that cause the development of diabetes. Aminotransferases are generally considered biomarkers for hepatocellular health.<sup>31</sup> ALT is predominantly found in the liver and is associated with the accumulation of liver fat. The liver is the site of insulin clearance<sup>32</sup> and if insulin losses its ability to suppress glucose production in the liver, it causes a surge in liver glucose production leading to diabetic progression.33 Diabetes is most likely to cause non-alcoholic fatty liver disease (NAFLD). ALT is a classic marker for NAFLD.<sup>34</sup> Like ALT, Aspartate aminotransferase is also a live enzyme and abnormal levels indicate cholestatic or liver damage.<sup>35</sup> An increase in ALT and AST in the streptozotocin-induced groups suggest hepatic damage and treatment with FBHB extracts alleviates the increased aminotransferase levels and indicates its hepatoprotective roles even in a diabetic environment. Histopathological evaluation of Zebrafish kidneys was performed to evaluate the effect of streptozotocin and FBHB extracts. In the STZ-induced groups, the destruction in cellularity of the endocrine islet can be clearly visualized indicating a severed pancreas. Administering a dose of  $10 \,\mu\text{g/mL}$  of FBHB extracts reduced the infiltration of immune cells compared to a dose of 5  $\mu$ g/mL. At the highest dose  $(15 \,\mu g/mL)$ , the pancreatic cells seem to be recovering well.

## CONCLUSION

Induction of diabetes using streptozotocin was confirmed by the rise in blood glucose. The hydroethanolic bark extract of *Ficus benghalensis* has hypoglycemic effects as well as suppresses dyslipidemia and hypertriglyceridemia that is caused due to diabetes. Hepatic enzymes such as alanine aminotransferase (ALT), alkaline phosphatase (ALP), and aspartate aminotransferase (AST) are biomarkers of liver damage were also reduced in the FBHB extract-treated group. Overall, FBHB extract can act as a therapeutic agent and could be used for effectively managing diabetes mellitus. However, in order to understand the compound responsible for these effects and the mechanism of action, further studies are required.

## ACKNOWLEDGEMENT

The authors would like to acknowledge the College of Medicine Research Center, Deanship of Scientific research, King Saud University, Riyadh, Saudi Arabia for support.

## **CONFLICT OF INTEREST**

The authors declare that there is no conflict of interest.

### ABBREVIATIONS

**FBHB:** *Ficus benghalensis* Hydroethanolic Bark; **STZ**: Streptozotocin; **i.p:** Intraperitoneal; **ALP**: Alkaline phosphatase; **ALT**: Alanine aminotransferase; **NaOH**: Sodium Hydroxide; **AST**: Aspartate aminotransferase; **LDL-R**: Low Density Lipoprotein receptor; **ANOVA**: Analysis of Variance; **NAFLD**: Non-alcoholic fatty liver disease.

#### REFERENCES

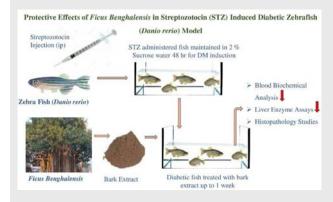
- Vieira R, Souto SB, Sánchez-López E, Machado AL, Severino P, Jose S, et al. Sugar-Lowering Drugs for Type 2 Diabetes Mellitus and Metabolic Syndrome-Review of Classical and New Compounds: Part-I. Pharmaceuticals (Basel). 2019;12(4). doi: 10.3390/ph12040152, PMID 31658729.
- Raghavan S, Vassy JL, Ho YL, Song RJ, Gagnon DR, Cho K, *et al.* Diabetes mellitus–related all-cause and cardiovascular mortality in a national cohort of adults. J Am Heart Assoc. 2019;8(4):e011295. doi: 10.1161/ JAHA.118.011295, PMID 30776949.
- Sun H, Saeedi P, Karuranga S, Pinkepank M, Ogurtsova K, Duncan BB, *et al.* IDF Diabetes Atlas: Global, regional and country-level diabetes prevalence estimates for 2021 and projections for 2045. Diabetes Res Clin Pract. 2022;183:109119. doi: 10.1016/j.diabres.2021.109119, PMID 34879977.
- Tan SY, Mei Wong JL, Sim YJ, Wong SS, Mohamed Elhassan SA, Tan SH, et al. Type 1 and 2 diabetes mellitus: A review on current treatment approach and gene therapy as potential intervention. Diabetes Metab Syndr. 2019;13(1):364-72. doi: 10.1016/j.dsx.2018.10.008, PMID 30641727.
- Simos YV, Spyrou K, Patila M, Karouta N, Stamatis H, Gournis D, *et al.* Trends of nanotechnology in type 2 diabetes mellitus treatment. Asian J Pharm Sci. 2021;16(1):62-76. doi: 10.1016/j.ajps.2020.05.001, PMID 33613730.
- Bindu Jacob, Narendhirakannan R T. Role of medicinal plants in the management of diabetes mellitus: A review. 3 Biotech. 2019;9(1):4. doi: 10.1007/s13205-018-1528-0, PMID 30555770.
- Nair A, Chattopadhyay D, Saha B. Plant-derived immunomodulators. New Look Phytomed Adv Herb Prod Novel Drug Leads. 2018. doi: 10.1016/B978-0-12-814619-4.00018-5.
- Murugesu S, Selamat J, Perumal V. Phytochemistry, pharmacological properties, and recent applications of *Ficus benghalensis* and *Ficus religiosa*. Plants (Basel). 2021;10(12):2749. doi: 10.3390/PLANTS10122749, PMID 34961220.
- Singh D, Singh B, Goel RK. Traditional uses, phytochemistry and pharmacology of *Ficus religiosa*: A review. J Ethnopharmacol. 2011;134(3):565-83. doi: 10.1016/j.jep.2011.01.046, PMID 21296646.
- Karmakar S, Paul S, Biswas NM, Khanam J, Kar SK, Mukherjee H, *et al.* A pharmacological audit and advancement on the recent trend of research on *Ficus benghalensis* L. including its *in vitro* hepatoprotective activity. Clin Phytosci. 2020;6(1). doi: 10.1186/s40816-020-00230-8.
- Vaidyanathan L, Deebiga TH. Screening and evaluation of antidiabetic potency of *Vernonia arborea* using zebrafish model. Int J Pharm Sci Res. doi: 10.13040/IJPSR.0975-8232.12(1).385-94.
- Reitman S, frankel S. A colorimetric method for the determination of serum glutamic oxalacetic and glutamic pyruvic transaminases. Am J Clin Pathol. 1957;28(1):56-63. doi: 10.1093/ajcp/28.1.56, PMID 13458125.

- Ahmad Khan MS, Herbal Medicine AI. Current trends and future prospects. New Look Phytomed Adv Herb Prod Novel Drug Leads. 2018. doi: 10.1016/ B978-0-12-814619-4.00001-X.
- Zang L, Maddison LA, Chen W. Zebrafish as a model for obesity and diabetes. Front Cell Dev Biol. 2018;6(AUG):91. doi: 10.3389/fcell.2018.00091, PMID 30177968.
- Moss JB, Koustubhan P, Greenman M, Parsons MJ, Walter I, Moss LG. Regeneration of the pancreas in adult zebrafish. Diabetes. 2009;58(8):1844-51. doi: 10.2337/db08-0628, PMID 19491207.
- Intine RV, Olsen AS, Sarras MP. A zebrafish model of diabetes mellitus and metabolic memory. J Vis Exp. 2013;72(72):e50232. doi: 10.3791/50232, PMID 23485929.
- Mooradian AD. Dyslipidemia in type 2 diabetes mellitus. Nat Clin Pract Endocrinol Metab. 2009;5(3):150-9. doi: 10.1038/ncpendmet1066, PMID 19229235.
- Fernandez ML, Andersen CJ. Effects of dietary cholesterol in diabetes and cardiovascular disease. Clin Lipidol. 2014;9(6):607-16. doi: 10.2217/ clp.14.40.
- Fryirs M, Barter PJ, Rye KA. Cholesterol metabolism and pancreatic β-cell function. Curr Opin Lipidol. 2009;20(3):159-64. doi: 10.1097/ MOL.0b013e32832ac180, PMID 19417651.
- Perego C, Da Dalt L, Pirillo A, Galli A, Catapano AL, Norata GD. Cholesterol metabolism, pancreatic β-cell function and diabetes. Biochim Biophys Acta Mol Basis Dis. 2019;1865(9):2149-56. doi: 10.1016/j.bbadis.2019.04.012, PMID 31029825.
- Pochhi M, Muddeshwar M. Hypoglycemic and antihyperlipidemic effect of aqueous leaves extract of *Ficus religiosa* in alloxan induced diabetic rats. Asian J Med Sci. 2017;8(2):50-5. doi: 10.3126/ajms.v8i2.16304.
- Singh AK, Singh R. Triglyceride and cardiovascular risk: A critical appraisal. Indian J Endocrinol Metab. 2016;20(4):418-28. doi: 10.4103/2230-8210.183460, PMID 27366705.
- Maahs DM, Daniels SR, De Ferranti SD, Dichek HL, Flynn J, Goldstein BI, et al. Cardiovascular disease risk factors in youth with diabetes mellitus: A scientific statement from the American Heart Association. Circulation. 2014;130(17):1532-58. doi: 10.1161/CIR.00000000000094, PMID 25170098.
- Hartz JC, De Ferranti S, Gidding S. Hypertriglyceridemia in diabetes mellitus: Implications for pediatric care. J Endocr Soc. 2018;2(6):497-512. doi: 10.1210/js.2018-00079, PMID 29850649.
- Alexopoulos AS, Qamar A, Hutchins K, Crowley MJ, Batch BC, Guyton JR. Triglycerides: Emerging targets in diabetes care? Review of moderate

hypertriglyceridemia in diabetes. Curr Diab Rep. 2019;19(4):13. doi: 10.1007/ s11892-019-1136-3, PMID 30806837.

- Harmey D, Hessle L, Narisawa S, Johnson KA, Terkeltaub R, Millán JL. Concerted regulation of inorganic pyrophosphate and osteopontin by Akp2, Enpp1, and ank: An integrated model of the pathogenesis of mineralization disorders. Am J Pathol. 2004;164(4):1199-209. doi: 10.1016/S0002-9440(10)63208-7, PMID 15039209.
- Chen SCC, Tsai SP, Jhao JY, Jiang WK, Tsao CK, Chang LY. Liver fat, hepatic enzymes, alkaline phosphatase and the risk of incident Type 2 diabetes: A prospective study of 132,377 adults [sci rep]. Sci Rep. 2017;7(1):4649. doi: 10.1038/s41598-017-04631-7, PMID 28680048.
- Kim DW, Hwang SY, Nam YJ, Kim D, Shin SJ, Yoon HE. The combined prognostic significance of alkaline phosphatase and vascular calcification in patients with end-stage kidney disease. Nutr Metab Cardiovasc Dis. 2020;30(9):1476-83. doi: 10.1016/j.numecd.2020.04.029, PMID 32586735.
- Zhao L, Li L, Ren H, Zou Y, Zhang R, Wang S, *et al.* Association between serum alkaline phosphatase and renal outcome in patients with type 2 diabetes mellitus. Ren Fail. 2020;42(1):818-28. doi: 10.1080/0886022X.2020.1804402, PMID 32781868.
- Fadini GP, Pauletto P, Avogaro A, Rattazzi M. The good and the bad in the link between insulin resistance and vascular calcification. Atherosclerosis. 2007;193(2):241-4. doi: 10.1016/j.atherosclerosis.2007.05.015, PMID 17606264.
- York MJ. Clinical pathology. A Compr guid to Toxicol nonclinical drug dev. Published online January 1, 2017:325-74. doi: 10.1016/B978-0-12-803620-4.00014-1.
- Najjar SM, Perdomo G. Hepatic insulin clearance: Mechanism and physiology. Physiology (Bethesda). 2019;34(3):198-215. doi: 10.1152/ physiol.00048.2018, PMID 30968756.
- Sharabi K, Tavares CDJ, Rines AK, Puigserver P. Molecular pathophysiology of hepatic glucose production. Mol Aspects Med. 2015;46:21-33. doi: 10.1016/j.mam.2015.09.003, PMID 26549348.
- Mahran HN, Saber LM, Alghaithy AA, Elareefy AA. The role of elevated alanine aminotransferase (ALT), FASL and atherogenic dyslipidemia in type II diabetes mellitus. J Taibah Univ Med Sci. 2017;12(1):8-13. doi: 10.1016/j. jtumed.2016.10.002, PMID 31435207.
- Giannini EG, Testa R, Savarino V. Liver enzyme alteration: A guide for clinicians. CMAJ. 2005;172(3):367-79. doi: 10.1503/cmaj.1040752, PMID 15684121.

#### **PICTORIAL ABSTRACT**



#### SUMMARY

- *Ficus benghalensis* have positive effects in alleviating experimentally induced diabetes.
- FBHB extracts has hypoglycemic, hypotriglyceridemic and hypocholesterolemic potency in STZ-induced diabetes.
- FBHB extracts reduced hepatocellular health biomarkers - ALP, AST, and ALT indicating hepatoprotective effects in diabetic mellitus.

## **About Authors**



**Dr. Essa M Sabi**, MPhil-PhD, is currently working as Assistant Professor and Consultant of Biochemical Genetics in the Departments of pathology and Laboratory Medicine - College of Medicine - King Saud University – Riyadh - Saudi Arabia. He has experience at academic, clinical and research levels in various disciplines related to the clinical biochemistry field. Additionally, he is supervising research groups of medical students. His area of interest includes clinical biochemistry, signaling, metabolomics, therapeutical tumour targeting, angiogenesis and cancer.



**Dr. Ahmed H Mujamammi,** MPhil-PhD, is currently working as Assistant Professor and Consultant of Biochemical Genetics in the Departments of Pathology and Laboratory Medicine - College of Medicine - King Saud University (KSU) – Riyadh - Saudi Arabia. He is the deputy of Medical Biochemistry Fellowship at KSU, and he has experience at academic, clinical and research levels in various disciplines related to the clinical biochemistry field. Additionally, he is supervising research groups of medical students. His area of interest includes clinical biochemistry, signaling, metabolomics, therapeutical tumour targeting, angiogenesis and cancer.



**Dr. Ziyad M Althafar,** PhD, is currently working as Assistant Professor and Senior Clinical Immunologist in the Department of Medical Laboratories - College of Applied Medical Sciences –Shaqra University – Riyadh - Saudi Arabia. He is the Vice Dean of the academic affairs at the College of Applied Medical Sciences – Shaqra University, and he has experience at academic, clinical and research levels in various disciplines related to the clinical immunology field. His area of interest includes clinical immunology, infection, inflammation, cell signaling and biochemistry.



**Dr. Samia T. Al-Shouli MD,** PGDip, MSc, PhD, Clinical Fellowship is currently working as Assistant Professor, physician and scientist in clinical immunology and Allergy, Pathology Department, College of Medicine, King Saud University, Riyadh, Saudi Arabia. Her major interest in management of Primary Immune Deficiency, Allergy and immunotherapy. She has initiated a number of research projects and works to promote research and education.

**Dr. Lotfi Saeed Bin Dahman MD**, Ph.D. Medical Biochemistry and Molecular Biology. He graduated from Assiut University with Ph.D. in Medical Biochemistry, collaboration with King Saud University (2008-2013). He was worked as a researcher at the Obesity Research Center and Chair for Biomarkers of Chronic Diseases, King Saud University (2008-2013). My current job as an associate professor of Medical Biochemistry at the College of Medicine and Health Sciences, Hadhramout University. Head of Medical Laboratory Sciences at the College of Medicine and Health Sciences, Hadhramout University (2018-till now). Director of International Cooperation, Hadhramout University (2018-till now). He is interested in diabetes, hypertension, obesity, cardiovascular disease, cell biology, and cancer biology research.



**Dr. Khalid M Sumaily, MD**, KSUFMB, IFCAP, FBCG, is currently working as Assistant Professor and Consultant of Medical Biochemistry and Biochemical Genetics in the Departments of Pathology and Laboratory Medicine - College of Medicine - King Saud University (KSU) – Riyadh – Saudi Arabia. He is the Director of Medical Biochemistry Fellowship at KSU, and he has experience at academic, clinical and research levels in various disciplines related to the clinical biochemistry field. Additionally, he is supervising research groups of medical students. His area of interest includes clinical biochemistry, signaling, metabolomics, therapeutical tumour targeting, angiogenesis and cancer.

**Cite this article:** Sabi EM, Mujamammi AH, Althafar ZM, Al-Shouli ST, Dahman LSB, Sumaily KM. Protective Effects of *Ficus benghalensis* in Streptozotocin (STZ) Induced Diabetic Zebrafish (*Danio rerio*) Model. Indian J of Pharmaceutical Education and Research. 2022;56(3):822-9.