

# Aromatase Inhibitors: Development and Current Perspectives

Nusrat B Sayyad, Prafulla M Sabale\*, Mohit D Umare, Komal K Bajaj

Department of Pharmaceutical Sciences, Rashtrasant Tukadoji Maharaj Nagpur University, Nagpur, Maharashtra, INDIA.

## ABSTRACT

**Aim:** Breast cancer is one of the most common cancers among women, consistently elevated estrogen level in the blood associated with persistently in breast cancer found in many studies. The mechanisms of carcinogenesis whereby cancer is associated involved in the conversion of androgen to estrogen in the presence of enzyme aromatase. **Materials and Methods:** Aromatase is a (CYP19), a cytochrome P450, which is the enzyme that synthesizes estrogen. Aromatase which is expressed in higher concentration in breast play a pivotal role in the origin and progression of breast cancer. *In situ* inhibition of aromatase enzyme accomplished by the prevention of aromatase expression in breast cancer. **Results:** The success of aromatase inhibition could result in the suppression and progression of breast cancer. One of the ways to reduce the incidence of breast cancer by inhibiting estrogen biosynthesis in breast tissue. Aromatase inhibitor therapy is considered for second-line treatment in patients who fail anti-estrogen therapy. **Conclusion:** Several potent and selective aromatase inhibitors have been developed and used to treat breast cancer. Some of them have passed clinical trial phase II. Development in the field has encouraged us to review work on the Aromatase inhibitor.

**Key words:** Non-Steroidal aromatase inhibitor, Aromatase, Cancer, Aromatase Inhibitors.

## INTRODUCTION

According to WHO women breast cancer is the most recurring cancer, hit almost 2.1 million women each year, and also the greatest root for cancer-related death. In 2018 it was estimated that 15% of all cancer death in women due to breast cancer which is approximately 627,000 women died from breast cancer. Familial and hereditary risk factors contribute to the occurrence of breast cancer. Other factors that increase risk in breast cancer are mutation in BRCA1, BRCA2, and p53 genes.<sup>1</sup> Patients with familial or non-familial gynecomastia Elevated expression of aromatase reported in breast epithelium, skin fibroblast, and peripheral lymphocytes.<sup>2,3</sup> Aromatase is an enzyme involved in the production of estrogen from the androgen and is responsible for catalyzing the last rate limiting/crucial/ key step for the biosynthesis of estrogen.<sup>4</sup> These are a group of steroid hormones that promote the

development and maintenance of female characteristics of the body.<sup>5</sup> Estrogen signalling pathways take part in diverse particularly cell survival and proliferation. In addition to the reproductive system, estrogen play important role in the cardiovascular system, musculoskeletal system and brain,<sup>6</sup> Non-steroidal aromatase inhibitor (aminoglutethimide) act by binding with the heme moiety (non-covalently and reversible) of the enzyme aromatase and prevent the binding of androgens. It was hypothesized that this may involve the interaction of the nitrogen atom of aminoglutethimide with the heme moiety.<sup>7</sup> The research was carried out and some imidazole antimycotic drugs were found to be effective in inhibiting aromatase but were not suitable for antitumor activity in breast cancer. Miconazole, clotrimazole, ketoconazole was found to have better inhibitory activity than aminoglutethimide

Submission Date: 17-10-2021;  
Revision Date: 29-12-2022;  
Accepted Date: 24-02-2022.

DOI: 10.5530/ijper.56.2.51

Correspondence:

Dr. Prafulla M Sabale  
Department of  
Pharmaceutical Sciences,  
RTM Nagpur University,  
Nagpur-440033,  
Maharashtra, INDIA.  
E-mail: prafullasable@  
yahoo.com



www.ijper.org

but are not used for antitumor activity. Azole derivatives like vorozole anastrozole and fadrazole all having 1,2,4-triazole ring were found to have excellent antitumor activity in the treatment of breast cancer by inhibiting aromatase enzyme.<sup>8-10</sup>

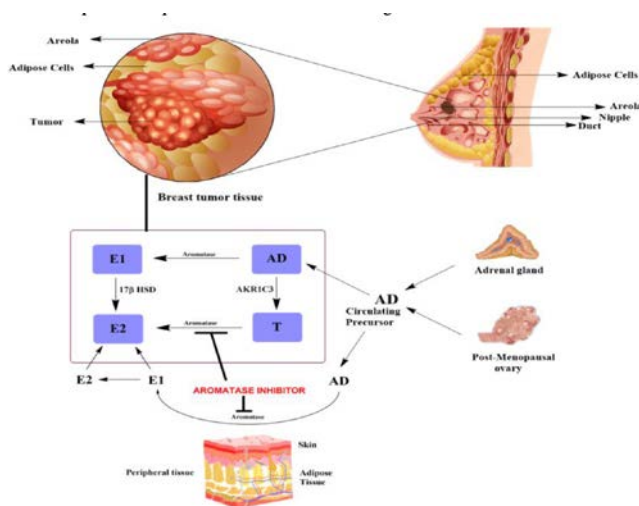
### Why aromatase inhibitors?

Aromatase reaction is the last step in the biosynthetic pathway of estrogen. The inhibition of this last step ensures that the bio-synthesis of other steroids classes remains unaffected.<sup>11</sup>

Aromatase inhibitor's role in post-menopausal: Primary estrogen source in post menopausal women is aromatase activity in breast, bone, vascular endothecium, and CNS. Also, gonadotropin regulation cannot regulate the aromatase level in post menopausal women and this avoids the complications which are due to feedback regulatory mechanism that increases LH and FSH after aromatase inhibition. Therefore, AIs are employed for the therapy of breast cancer in post-menopause women as shown in Figure 1.<sup>12-14</sup>

### Biosynthesis

Steroids producing cells are characterized by intracellular deposits. They contain mitochondria containing cholesterol side-chain enzyme (P450 SCC) and Esterified cholesterol. They also contain aromatase enzyme 3, 8 hydroxysteroid dehydrogenase (HSD) / isomerase 17,18-HSD enzymes, these enzymes are present in the smooth endoplasmic reticulum and Golgi bodies, involved in the formation of P storage granules.<sup>15,16</sup> The amount of smooth endoplasmic reticulum, mitochondria, Golgi bodies, and lipid in the granulosa cells are less in the smallest growing follicles.

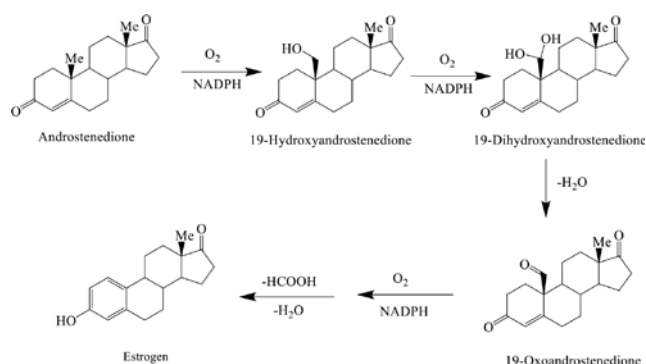


**Figure 1: Regulation and its Expression of aromatase in the breast cancer and peripheral system.**

A drastic change is seen in developing granulosa cells. This suggests that developing follicles increase the rate of Steroidogenesis. Steroidogenesis, stimulation of follicular growth, and steroidogenic enzyme mRNA are promoted by FSH and LH.<sup>17</sup> This leads to promotion in the growth of steroidogenic cells. Aromatase or estrogen synthase is an enzyme responsible for a key step in the biosynthesis of estrogen.<sup>18-20</sup> The gene that encodes an aromatic protein in humans is CYP19A1 and ranges approximately 123kb on chromosome 15q21.2. It consists of 93 Kb 5'-unsaturated region (UTR), 30kb coding region, and 3' end. The coding region starts with ATG, translation start site on Exon II as the coding region contains 9 exons (II-X) coding region.<sup>21</sup> Several unsaturated first exons are regulated by the tissue-specific promoter in 5'-UTR. There is 10 alternate tissue-specific promoter found in humans which includes I.1, I.2, I.2a in the placenta, I.4 in adipose tissue and skin, I.5 in fetal tissue in the brain, I.7 in bones, I.3 in adipose tissue PII in gonads and adipose tissue.<sup>22,23</sup> These promoters regulate aromatase expression in different tissues.<sup>22,24</sup> Cytochrome P450 is a superfamily of enzymes that catalyzes the estrogen biosynthesis.<sup>25</sup> The phenolic ring a characteristic feature of estrogen formed by binding of C19 steroid substrate and catalyzes the series of reactions with the help of heme protein. P450 is a flavoprotein, NADPH cytochrome P450 reductase.<sup>26</sup> These are ubiquitous proteins in the endoplasmic reticulum. When it comes in contact with NADPH it transfers reducing equivalents to any form of cytochrome P450. The reductase may be the product of a single gene. In superfamily of genes cytochrome P450, the aroma is the only member of gene family 19 denoted as CYP19. It is denoted as CYP19 based on the fact that oxygen attacks on the angular methyl group at C19. For the metabolism of C19, the aromatase reaction utilizes three molecules of NADPH and three molecules of oxygen. Various shreds of evidence suggest that all three oxygen molecules are used in the oxidation of the C19 angular methyl group which forms formic acid. This process occurs simultaneously with the aromatization of a ring to phenolics structures. This ring is characteristic of estrogen as illustrated in Figure 2.<sup>25,27,28</sup>

### Structure

At the junction of beta-sheet 3, B-C loop, and the I and F helices the catalytic site of the aromatase enzyme is located. The C19 is placed in proximity to 4.0 Å of an iron atom and beta face of Androstenedione towards Fe group of the enzyme. Binding pocket is formed of about 400 Å by moving I helix Backbone by 3.5 Å. Residue P308 plays an important role in the Catalytic activity of



**Figure 2: Mechanism Cascade in the biosynthesis of Aromatase Enzyme.**

aromatase as without P308 steric hindrance can decrease catalytic activity.<sup>27</sup> This P308 located on the distal side of I helix from androstenedione. It is not present in any of Cytochrome P450 family enzymes and can be used for site-directed mutagenesis studies.<sup>29</sup> Several hydrophobic Residues are piled against the alpha face Backbone. This long chain results in a deep spherical active site which increases specificity. Long-distance of steroids makes them more specific against aromatase.<sup>6,30,31</sup> The active site of aromatase is present in the spherical molecule illustrated in Figure 3 at the centre of the heme distal cavity. Various studies showed that androstenedione binds with C19 at 4.0 Å from the Fe atom and  $\beta$  face towards the heme group.<sup>27</sup> Various polypeptide segments present in the catalytic cleft of different loops and helix are Ile305, ASP309, Thr310, Ala306 from I helix. Trp224 and Phe221 from F-helix, Phe134 and Ile133 from B-C loop, Val370, Val373, Leu372 from K- $\beta$ 3 loop, Leu477 and Ser478 from  $\beta$ 8- $\beta$ 9 loop and Met 374 from  $\beta$ 3. The cavity of complementary to androstenedione's shape is formed by tight packing against the steroid backbone of hydrophobic residue and porphyrin ring of heme.<sup>32</sup> As shown in Figure 3 a and b different combinations of side-chain residue make interaction and attachment on androstenedione.<sup>33-35</sup> Vander wall forces approaching substrate from the alpha face and follows contour, some make contact at the edges. All these residue and polypeptide forms a pocket of approx. 400 Å with tightly encloses androstadienone. The presence of two hydrogen bonds and their geometrics are arranged in such a way that 3-Keto O1 and a water Oxygen atom are in the carboxylate plane, this provides strong hydrogen bonding interaction. In ASP 309 sidechain protonation and deprotonation are required of the carboxylate group for substrate binding interaction and catalysis. This acts as a proton relay network which helps in androgen to estrogen conversion in which water molecule and polar



**Figure 3: Ribbon diagram showing the structure of the human placental aromatase with ligand androstenedione.<sup>30</sup>**

side chain act as a proton source. The only hole in the binding pocket is located where 3 water molecules are present. This channel opens in the exterior part of the protein surface. The flexibility of tertiary structure allows smooth passage of steroidal structure.<sup>36,37</sup>

## Generation

### First Generation

#### Aminoglutethimide

Aminoglutethimide is manufactured by Novartis and marketed under the name of Cytadren, for anticonvulsant activity in 1966 and discontinued because it has more effects on the adrenal gland. It was approved by the FDA on 10/29/1980 for the treatment of breast cancer and other hormone-sensitive cancers in 1980.<sup>38</sup> It was discontinued later, its chemical name is 3-(4-aminophenyl)-3-ethyl-2,6-piperidinedione. The drug inhibits the conversion of cholesterol to  $\Delta$ 5-pregnenolone leads to a decrease in the production of adrenal glucocorticoids, mineralocorticoids, estrogens, and androgens. Simultaneous administration of hydrocortisone is necessary to suppress the increased levels of ACTH. Cytadren is administered orally of 250 mg tablets. Use of aminoglutethimide (1000mg conventional dose) shows early side effects which are mild and frequent, while late side effects are infrequent. About 40% of the patients develop lethargy and ataxia (usually disappear over 4 weeks). About one-third of the patients show a morbilliform skin rash along with fever. It is moderate and rarely serious (erythroderma syndrome). Other side effects occur infrequently but are clinically relevant, such as hypothyroidism, hematologic toxicity (prolonged thrombocytopenia, agranulocytosis, pancytopenia), pulmonary haemorrhage

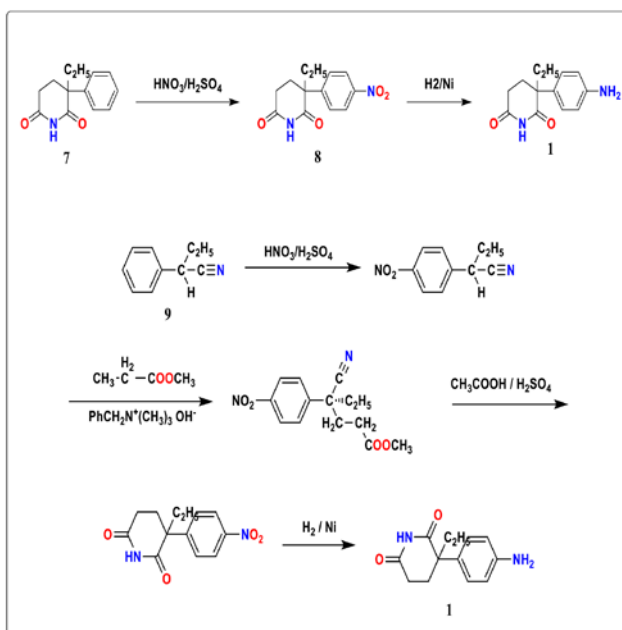


Figure 4: Synthesis of Aminogluthemide.

due to alveolar damage, cholestasis, and systemic lupus erythematosus.<sup>39,40</sup> Synthesis of 1 (Figure 4) requires commercially available Glutethimide 7 which is nitrated to form 2-(4-nitrophenyl)-2-ethylglutarimide 8 reductions of the nitro group using nickel catalyst to give aminogluthemide 1. As glutethimide is expensive another method for synthesis is using commercially available 2-phenylbutyronitrile. 2-phenylbutyronitrile 9 is nitrated to form 2-(4-nitrophenyl)butyronitrile under Michael addition reaction conditions in the presence of benzyl trimethyl, ammonia hydroxide is added to methyl acrylate, the product obtained undergo acid hydrolyses using a mixture of acetic acid and sulphuric acid, a cyclization occurs to 2-(4-nitrophenyl)-2-ethylglutarimide, the product is reduced by hydrogen to give the desired product aminogluthemide 1.<sup>41,42</sup>

### Testolactone

The first steroid used for breast cancer treatment is Testolactone. Testolactone is manufactured by Bristol-Myers Squib and marketed under the name of Teslac, which was approved by the FDA in 1970. It is a synthetic antineoplastic agent with a six-membered lactone ring in place of the usual five-membered carbocyclic D-ring of androgen steroid nucleus.<sup>43</sup> Testolactone is chemically designated as 13-hydroxy-3-oxo-13,17-secoandrosta-1,4-dien-17-oic acid  $\delta$ -lactone. It is available in the form of tablets 50mg for oral administration, an increase in a dose up to 1000mg leads to increase renal tubular reabsorption of calcium but no serum calcium concentration observed. Now

it is discontinued by the FDA. Exact mechanism of antineoplastic activity of Testolactone has not been established. The principal mechanism is suggested to be a reduction in estrone synthesis from adrenal androstenedione by inhibiting the aromatase enzyme.<sup>42,44</sup> Although it is having some similarities with testosterone but not having any androgenic effects. The drug shows non-competitive, irreversible inhibition. This inhibition leads to suppression of tumour growth. Testolactone demonstrated a very low rate of toxic effects, consisting of vomiting, nausea, and diarrhoea, with no side effects of an endocrine nature. The synthetic approach involved the initial formation of the lactone ring of ring D by Baeyer-Villiger Oxidation using peracetic acid followed the formation of the intermediate by addition of double bond in ring A by using selenium dioxide oxidation.<sup>39,45,46</sup>

### Second Generation

#### Formestane

Formestane was marketed under the name of Lentrone by Ciba-Geigy Pharmaceuticals was not approved by FDA.<sup>47,48</sup> intramuscular injection form Lentrone was approved in Europe has been withdrawn. Formestane was the first selective, second-generation, type I, steroidal aromatase inhibitor used in the treatment of estrogen-receptor-positive breast cancer in postmenopausal women. Formestane suppresses estrogen production from anabolic steroids or prohormones.<sup>49</sup> Its chemical name is 4-Hydroxyandrost-4-ene-3,17-dione. Major systemic side effects include hot flushes and vaginal spotting, lethargy, rash, nausea, and dizziness.<sup>50</sup> Formestane (3) in Figure 5 was synthesized from testosterone (9). The oxidation of testosterone (9) with Jones reagent gives androst-4-ene-3,17-dione which on

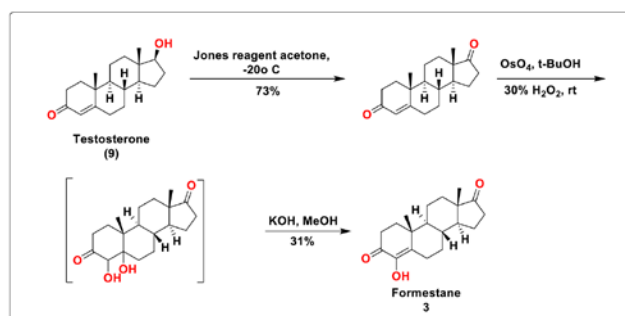


Figure 5: Synthesis of Formestane.

Hydroxylation with  $OsO_4/H_2O_2$  followed by alkaline dehydration of the resultant diols gave formestane (3).<sup>51</sup>

### Third Generation

#### Exemestane

Exemestane (Aromasin) (Pfizer) is a US FDA approved drug in 1999 it is a steroid aromatase inhibitor chemically

known as 6-methylideneandrosta-1,4-diene-3,17-dione (IUPAC name (1S,2R,10R,11S,15S)-2,15-dimethyl-8-methylidene-tetracyclo[8.7.0.0.2,7.0.11,15]heptadeca-3,6-diene-5,14-dione) used as an adjunct therapy in postmenopausal women for advanced breast cancer which is estrogen receptor-positive. It is also used in advanced breast cancer who received tamoxifen for about two to three years and then switch to exemestane to complete 5 years of therapy. A single 25 mg dose of Aromasin was found to be effective in both types of cancer it shows a long effect in reducing estrogen synthesis. Exemestane is a steroidal aromatase inhibitor that binds irreversibly to the aromatase enzyme. The structure of androstenedione which is a natural substrate and exemestane have similarities which show fake substrate binding to the aromatase enzyme. This long-duration action can be due to irreversible reaction on aromatase inhibitor as compared to the pharmacokinetic properties of the drug. Adverse drug reaction seen in early breast cancer for hot flashes, arthralgia, fatigue, increased sweating and headache and in advanced breast cancer, it was observed that hot flashes, fatigue, increased sweating were common additionally increase in appetite was seen. Exemestane (4) in Figure 6 is synthesized in various steps. First, a Vilsmeier-Haack reagent is synthesized for which paraformaldehyde and dimethylamine hydrochloride in isopentanol was refluxed using a Dean-Stark separator. Subsequently addition of androsta-1,4-dien-17 $\beta$ -ol-3-one (10) in the mixture after cooling to 10 to 15°C and then refluxed for 15 hr which give 6-methylene derivative (11). Further Jones oxidation in acetone and

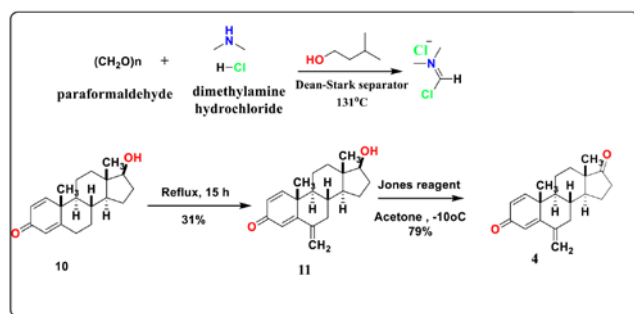


Figure 6: Synthesis of Exemestane.

recrystallization by ethanol and water (65:35) yields exemestane (4).

#### Fourth Generation

##### Anastrozole

Anastrozole (Arimidex® Astra Zeneca) is a US FDA approved in 1995 fourth-generation non-steroidal

aromatase inhibitor indicated for postmenopausal women with early hormone receptor-positive breast cancer, locally advanced or metastatic and in advanced breast cancer if the disease is progressed after initial tamoxifen therapy.<sup>52,53</sup> Chemically it is a nonsteroidal benzotriazole derivative. IUPAC name 2,2-[5-(1H-1,2,4-triazol-1-ylmethyl)-1,3-phenylene]bis(2-methyl-propiononitrile)). It is an orally active drug prescribed 1 mg tablet once daily. It acts by inhibiting aromatase enzyme by binding reversibly to the heme ion of CYP450 which reduces biosynthesis of estrogen. The adverse reaction seen in early breast cancer is hot flashes, asthenia, arthralgia, hypertension, nausea and vomiting, and arthritis. In the latest finding embryofetal toxicity is seen in animal studies and clinical trials in pregnant women.<sup>54-56</sup> Displacement of bromine in 3,5-bis(bromomethyl)toluene (12) with a nitrile group by SN2 mechanism forms 3,5-bis(nitrile)toluene (13) in the presence of potassium nitrate, tetrabutyl ammonium bromide and phase transfer catalyst. The bis nitrile product formed in DMF in excess of methyl iodide was deprotonated with the help of sodium hydride giving a bis dimethylated product (14). This bis dimethyl product undergoes Wohl-Ziegler reaction in the presence of benzoyl peroxide and n-bromosuccinimide. Finally,

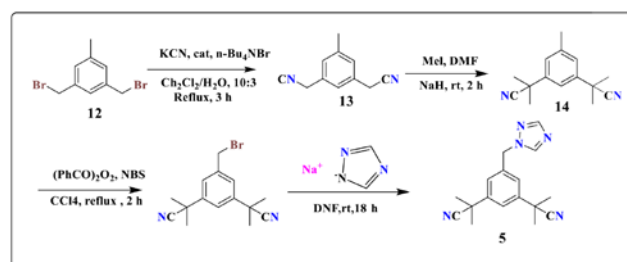
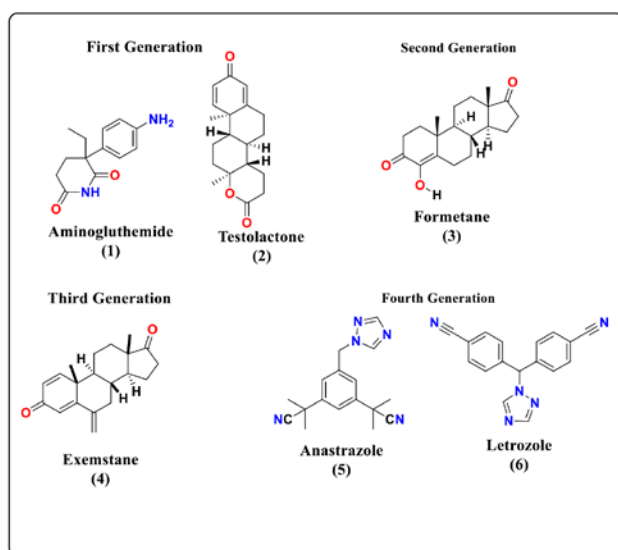


Figure 7: Synthesis of Anastrozole.

anastrozole as illustrated in Figure 7 is formed by the replacement of benzyl bromide with sodium triazole by an SN2 mechanism.<sup>57</sup>

##### Letrozole

Letrozole (Femara® Novartis) a US FDA 1997 and EU 1996 approved is a non-steroidal aromatase inhibitor prescribed for advanced breast cancer and as adjuvant therapy in postmenopausal women.<sup>58,59</sup> Chemically it is 4,40-[(1H-1,2,4-triazol-1-yl) methylene] bis-benzonitrile as shown in Figure 8. In the structure cyanobenzene moiety acts similarly as a steroidal backbone in non-steroidal aromatase inhibitor, this led to competitive inhibition.<sup>60</sup> The dose recommended is 2.5 mg tablet once a day. The duration of treatment is not confirmed but



**Figure 8: Generation of Aromatase Inhibitors.**

should be continued until tumour growth is reduced.<sup>61</sup> Letrozole is a competitive inhibitor of the aromatase enzyme. It acts by binding to heme ion of cytochrome P450 and reduces the level of estrogen, estradiol in serum. Arthralgia and arthritis were the major side effect of letrozole.<sup>62</sup> Other side effects were osteoporosis and hypercholesterolemia. The synthesis starts with SN2 substitution with imidazole in 4- bromomethylbenzonitrile as a starting material in the presence of methylene chloride. Subsequent deprotonation of the adduct in the presence of DMF, potassium- t- butoxide, and para fluobenzonitrile forms letrozole. This snar reaction forms as oil. Oil is converted into crystal form by converting into hemisuccinate. Its melting point is 149 to 150°C.<sup>57,63</sup>

### Resistance

Aromatase inhibitors function by blocking the aromatase enzyme so that less estrogen (endogenous) is available to stimulate the growth and proliferation of ER+ breast cancer cells.<sup>64</sup> The use of AI also reduces the risk of recurrence. Therapeutic resistance is common with AI and is responsible for limiting the use of AI for endocrine therapy.<sup>65,66</sup> In the treatment with AI if the therapy is not enough effective to decrease the size of the tumour and there is inadequate growth inhibition by AI is recognized as clinical resistance to AI.<sup>67</sup> In contrast to this, molecular and pathological changes are seen in the clinically resistant tumour by AI treatment.<sup>65,68</sup> Two types of resistance are known and recognized clinically while treating with endocrine therapy. Primary resistance (*denovo resistance*), ER+ tumours that are inadequate to respond to the endocrine treatment or the ER+ tumour is not sufficiently responding to endocrine

manipulations is recognized as primary resistance to AI. Secondary resistance (*acquired resistance*), ER+ tumour showing response to the therapy initially but later becomes resistant towards the therapy. Acquired resistance suggests that, while giving treatment with AI inductive changes or clonal selection occurs that changes the effectiveness of therapy and resulting in resistant cellular phenotype. Primary and secondary types of resistance might have common mechanisms of resistance.<sup>69,70</sup> Cross-resistance and non-cross resistance. Total cross-resistance, some breast cancer may show total cross-resistance to endocrine therapy. Some Breast tumours may show resistance not only to AI but also to other forms of endocrine therapy.<sup>71</sup> Non-cross resistance, Some breast tumours show resistance to AI but show effective response or sensitivity towards other endocrine therapy,<sup>72</sup> Specific non-cross resistance, Few tumours may show resistance for one AI or one class of AI but shoe response to another AI.<sup>73</sup>

### Mechanism of resistance

#### Ineffective or compromised inhibition of aromatase enzyme

Treatment with AI may sometimes be not sufficient or compromised. This is due to a lack of potency. Complete blockage of estrogen synthesis was not achieved by early generations of AI and the residual estrogen maintains the proliferation of tumour cells in the case of ER+ tumours.<sup>65,74</sup> Poor Pharmacokinetics, Aminoglutethimide catalyzes its metabolism by inducing liver cytochrome P450 enzymes.<sup>75</sup> Drug interaction: AI shows drug interaction with tamoxifen.<sup>76</sup> Concomitant administration of anastrozole or lenestrazole with tamoxifen leads to a decrease in the availability of letrozole by 30-40 % and anastrozole by 20-30% in plasma.<sup>59,77</sup> Compensatory endocrine loop: Ineffective inhibition of AI is also due to the high level of aromatase enzyme as in the case of premenopausal women. In premenopausal women, compensatory endocrine loops result in increased levels of gonadotropins which in turn stimulates the production of androgens and aromatase in the ovaries.<sup>78,79</sup> For use of AI in premenopausal women, concomitant use of luteinizing hormone-releasing hormone (LHRH)agonist which can block the rise in levels of gonadotropins. Mutation: Production of Mutant aromatase molecules that are resistant to AI or showing different sensitivity for different AI is possible.<sup>80</sup>

#### Exogenous estrogens

AI does not have any effect on exogenous estrogens like synthetic estrogens, industrial pollutants, and

phytoestrogens. Some adrenal androgens that can interact with ER.<sup>81</sup>

### Inherent estrogen sensitivity

AI inhibits the synthesis of endogenous estrogen and its action on estrogen receptors. This shows that ais can only be used in the case of ER+ tumours. ER+ tumours may not respond effectively while treating with AI. This may be due to a mutation in the ER.<sup>82</sup> In some ER+ tumours, hereditary material encoding for abnormal or mutant ER is reported and such receptors cannot transmit normal signals on binding with estrogen or estrogen antibodies. These tumours with mutant ER are inherently insensitive to hormone stimulation and therefore do not respond to AI therapy. Abnormality in the signalling pathway of ER like an abnormality in co-regulators may also result in insensitive ER+ tumours.<sup>83,84</sup>

### Activation of signalling pathways

Messenger systems are present that can activate ER signalling by low levels of estrogen or in the absence of estrogen. Example: HER Human Epidermal Factor receptor signalling can show phosphorylation of ER even without estrogen and activates the signalling pathway.<sup>85</sup> Other intracellular kinases capable of activating and supersensitized ER signalling include mapks and IGFR/ AKT.<sup>86</sup>

### Proliferation and growth of the tumour by estrogen-independent pathway

In some tumours, proliferation and growth of tumour cells are due to stimulation by estrogen-independent pathway. In this type of tumour, ER is normal and regulating estrogen-dependent process but tumour growth is under the influence of the ER independent pathway.<sup>87</sup> Such type of tumours is not affected by AI as they can only cause inhibition of estrogen-regulated pathway.<sup>88</sup>

### Cell Survival

Acquired resistance pattern is responsible for the development of cellular clones which can make the therapy ineffective against these clonal cells and they have a survival advantage over cells which are sensitive towards the treatment. Acquired resistant tumours are usually ER+ve and clones have ER.<sup>65,89</sup>

### CONCLUSION

Although there are aromatase inhibitors which are available in market for breast cancer, it also be used for ovulation induction in female infertility and

gynaecomastia. The success of aromatase inhibitor, developing small biologically active molecule with lower side effects and better tolerability are in process. One of the ways to block synthesis of estrogen is inhibition of aromatase enzyme, as it plays key role in rate limiting step. Since some drugs are in clinical trial pipeline for the treatment of breast cancer, many chemical entities are synthesized as selective inhibitor of aromatase enzyme. Bothersome side effects associated with aromatase inhibitor with long term therapy likely to be their effect on bone density. Although it can be minimised with concomitant use of potent bisphosphonate therapy to reduce bone thinning and osteoporosis. This review aims are facilitating more readers about development of selective, novel, potential chemical entities. Administration of aromatase inhibitors associated with increased level of follicle stimulating hormone (FSH), testosterone and LH. Lowering estradiol levels, therefore increase levels of testosterone in men with low testosterone levels.

### ACKNOWLEDGEMENT

The authors are thankful to the Head, Department of Pharmaceutical Sciences, RTM Nagpur university, Nagpur, Maharashtra, India for providing research facilities to carry out this research work.

### CONFLICT OF INTEREST

The authors declare that there is no conflict of interest.

### REFERENCES

1. Yadav MR, Barmade MA, Tamboli RS, Murumkar PR. Developing steroidal aromatase inhibitors-an effective armament to win the battle against breast cancer. *Eur J Med Chem.* 2015 Nov;105:1-38. doi: 10.1016/j.ejmech.2015.09.038, PMID 26469743.
2. Bulard J, Mowszowicz I, Schaison G. Increased aromatase activity in pubic skin fibroblasts from patients with isolated gynecomastia. *J Clin Endocrinol Metab.* 1987 Mar;64(3):618-23. doi: 10.1210/jcem-64-3-618, PMID 3818893.
3. Berkovitz GD, Guerami A, Brown TR, MacDonald PC, Migeon CJ. Familial gynecomastia with increased extraglandular aromatization of plasma carbon19-steroids. *J Clin Invest.* 1985 Jun 1;75(6):1763-9. doi: 10.1172/JCI111888, PMID 3924954.
4. Ji JZ, Lao KJ, Hu J, Pang T, Jiang ZZ, Yuan HL, Miao JS, Chen X, Ning SS, Xiang H, Guo YM, Yan M, Zhang LY. Discovery of novel aromatase inhibitors using a homogeneous time-resolved fluorescence assay. *Acta Pharmacol Sin.* 2014 Aug;35(8):1082-92. doi: 10.1038/aps.2014.53, PMID 25047514.
5. Zubeldia-Brenner L, Roselli CE, Recabarren SE, Gonzalez Deniselle MC, Lara HE. Developmental and functional effects of steroid hormones on the neuroendocrine axis and spinal cord. *J Neuroendocrinol.* 2016;28(7). doi: 10.1111/jne.12401, PMID 27262161.
6. Chumsri S, Howes T, Bao T, Sabnis G, Brodie A. Aromatase, aromatase inhibitors, and breast cancer. *J Steroid Biochem Mol Biol.* 2011 May;125(1-2): 13-22. doi: 10.1016/j.jsbmb.2011.02.001, PMID 21335088.
7. Ghodsi R, Hemmateenejad B. QSAR study of diarylalkylimidazole and diarylalkyltriazole aromatase inhibitors. *Med Chem Res.* 2016 May;25(5):834-42. doi: 10.1007/s00044-016-1530-1.

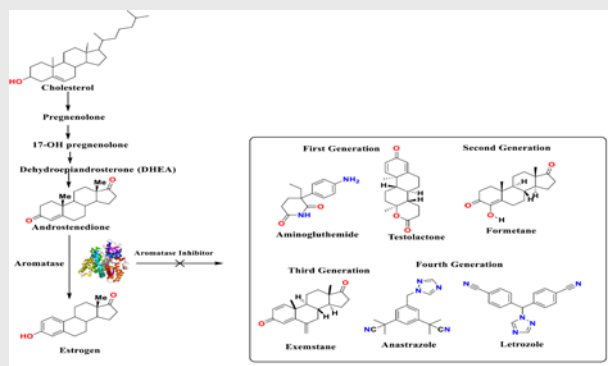
8. Bae SH, Park JH, Choi HG, Kim H, Kim SH. Imidazole antifungal drugs inhibit the cell proliferation and invasion of human breast cancer cells. *Biomol Ther (Seoul)*. 2018 Sep 1;26(5):494-502. doi: 10.4062/biomolther.2018.042, PMID 30092625.
9. Ballard SA, Lodola A, Tarbit MH. A comparative study of 1-substituted imidazole and 1,2,4-triazole antifungal compounds as inhibitors of testosterone hydroxylations catalysed by mouse hepatic microsomal cytochromes P-450. *Biochem Pharmacol*. 1988 Dec;37(24):4643-51. doi: 10.1016/0006-2952(88)90333-4, PMID 3202901.
10. Trösken ER, Fischer K, Völkel W, Lutz WK. Inhibition of human CYP19 by azoles used as antifungal agents and aromatase inhibitors, using a new LC-MS/MS method for the analysis of estradiol product formation. *Toxicology*. 2006 Feb;219(1-3):33-40. doi: 10.1016/j.tox.2005.10.020, PMID 16330141.
11. Avendaño C, Menéndez JC. Anticancer drugs that modulate hormone action. In: *Medicinal chemistry of anticancer drugs*. Elsevier; 2015 Jun. p. 81-131.
12. Mokbel K. The evolving role of aromatase inhibitors in breast cancer. *Int J Clin Oncol*. 2002 Oct;7(5):279-83. doi: 10.1007/s101470200040, PMID 12402060.
13. Mouridsen HT, Robert NJ. The role of aromatase inhibitors as adjuvant therapy for early breast cancer in postmenopausal women. *Eur J Cancer*. 2005 Aug;41(12):1678-89. doi: 10.1016/j.ejca.2004.10.020, PMID 16098456.
14. Nakata T, Takashima S, Shiotsu Y, Murakata C, Ishida H, Akinaga S, *et al.* Role of steroid sulfatase in local formation of estrogen in post-menopausal breast cancer patients. *J Steroid Biochem Mol Biol*. 2003 Sep;86(3-5):455-60. doi: 10.1016/S0960-0760(03)00357-1, PMID 14623544.
15. Balam FH, Ahmadi ZS, Ghorbani A. Inhibitory effect of chrysin on estrogen biosynthesis by suppression of enzyme aromatase (CYP19): A systematic review. *Heliyon*. 2020 Mar;6(3):e03557. doi: 10.1016/j.heliyon.2020.e03557, PMID 32181408.
16. Ratre P, Mishra K, Dubey A, Vyas A, Jain A, Thareja S. Aromatase inhibitors for the treatment of breast cancer: A journey from the scratch. *Anticancer Agents Med Chem*. 2020 Jun;20(17):1994-2004. doi: 10.2174/187152062066200627204105, PMID 32593281.
17. Andersen CY, Ezcurra D. Human steroidogenesis: Implications for controlled ovarian stimulation with exogenous gonadotropins. *Reprod Biol Endocrinol*. 2014 Dec 28;12:128. doi: 10.1186/1477-7827-12-128, PMID 25543693.
18. Shoham Z, Schachter M. Estrogen biosynthesis—regulation, action, remote effects, and value of monitoring in ovarian stimulation cycles. *Fertil Steril*. 1996 Apr;65(4):687-701. doi: 10.1016/S0015-0282(16)58197-7, PMID 8654622.
19. Williams GP, Darbre PD. Low-dose environmental endocrine disruptors, increase aromatase activity, estradiol biosynthesis and cell proliferation in human breast cells. *Mol Cell Endocrinol*. 2019 Apr;486:55-64. doi: 10.1016/j.mce.2019.02.016, PMID 30817981.
20. Boon WC, Chow JDY, Simpson ER. The multiple roles of estrogens and the enzyme aromatase. *Prog Brain Res*. 2010p;181:209-32. doi: 10.1016/S0079-6123(08)81012-6, PMID 20478440.
21. Ghosh D, Egbuta C, Kanyo JE, Lam TT. Phosphorylation of human placental aromatase CYP19A1. *Biochem J*. 2019 Nov 15;476(21):3313-31. doi: 10.1042/BCJ20190633, PMID 31652308.
22. Bulun SE, Chen D, Moy I, Brooks DC, Zhao H. Aromatase, breast cancer and obesity: a complex interaction. *Trends Endocrinol Metab*. 2012 Feb;23(2):83-9. doi: 10.1016/j.tem.2011.10.003, PMID 22169755.
23. Bulun SE, Lin Z, Imir G, Amin S, Demura M, Yilmaz B, *et al.* Regulation of aromatase expression in estrogen-responsive breast and uterine disease: From bench to treatment. *Pharmacol Rev*. 2005 Sep;57(3):359-83. doi: 10.1124/pr.57.3.6, PMID 16109840.
24. Zhao H, Zhou L, Shangguan AJ, Bulun SE. Aromatase expression and regulation in breast and endometrial cancer. *J Mol Endocrinol*. 2016 Jul;57(1):R19-33. doi: 10.1530/JME-15-0310, PMID 27067638.
25. Simpson ER, Mahendroo MS, Means GD, Kilgore MW, Hinshelwood MM, Graham-Lorence S, *et al.* Aromatase Cytochrome P450, The Enzyme Responsible for Estrogen. *Biosynthesis*. 1994;15(3):14.
26. Simpson ER, Mahendroo MS, Means GD, Kilgore MW, Corbin CJ, Mendelson CR. Tissue-specific regulation of aromatase cytochrome P450 (CYP19) expression. In: Berlin, Heidelberg: Springer. p. 611-25; 1993p. *Cytochrome P450 [internet]* Schenkman JB, Greim H, editors. (Handbook of Experimental Pharmacology; vol. 105).
27. Ghosh D, Griswold J, Erman M, Pangborn W. Structural basis for androgen specificity and oestrogen synthesis in human aromatase. *Nature*. 2009 Jan;457(7226):219-23. doi: 10.1038/nature07614, PMID 19129847.
28. Werck-Reichhart D, Feyereisen R. Cytochromes P450: A success story. *Genome Biol*. 2000;1(6):REVIEWS3003. doi: 10.1186/gb-2000-1-6-reviews3003, PMID 11178272.
29. Tsugaya M, Harada N, Tozawa K, Yamada Y, Hayashi Y, Tanaka S, *et al.* Aromatase mRNA levels in benign prostatic hyperplasia and prostate cancer. *Int J Urol*. 1996 Jul;3(4):292-6. doi: 10.1111/j.1442-2042.1996.tb00537.x, PMID 8844286.
30. Ghosh D, Griswold J, Erman M, Pangborn W. X-ray structure of human aromatase reveals an androgen-specific active site. *J Steroid Biochem Mol Biol*. 2010 Feb;118(4-5):197-202. doi: 10.1016/j.jsbmb.2009.09.012, PMID 19808095.
31. Hong Y, Li H, Yuan YC, Chen S. Molecular characterization of aromatase. *Ann N Y Acad Sci*. 2009 Feb;1155:112-20. doi: 10.1111/j.1749-6632.2009.03703.x, PMID 19250198.
32. Chan HJ, Petrossian K, Chen S. Structural and functional characterization of aromatase, estrogen receptor, and their genes in endocrine-responsive and -resistant breast cancer cells. *J Steroid Biochem Mol Biol*. 2016;161:73-83. doi: 10.1016/j.jsbmb.2015.07.018, PMID 26277097.
33. Narashimamurthy J, Rao AR, Sastry GN. Aromatase inhibitors: A new paradigm in breast cancer treatment. *Curr Med Chem Anticancer Agents*. 2004 Nov 1;4(6):523-34. doi: 10.2174/1568011043352669, PMID 15579017.
34. Chlebowski R, Cuzick J, Amakye D, Bauerfeind I, Buzdar A, Chia S, *et al.* Clinical perspectives on the utility of aromatase inhibitors for the adjuvant treatment of breast cancer. *Breast*. 2009 Aug;18(Suppl 2):S1-11. doi: 10.1016/S0960-9776(09)70002-5, PMID 19712865.
35. Dowsett M, Cuzick J, Ingle J, Coates A, Forbes J, Bliss J, *et al.* Meta-analysis of breast cancer outcomes in adjuvant trials of aromatase inhibitors versus tamoxifen. *J Clin Oncol*. 2010 Jan 20;28(3):509-18. doi: 10.1200/JCO.2009.23.1274, PMID 19949017.
36. Cojocaru V, Winn PJ, Wade RC. The ins and outs of cytochrome P450s. *Biochim Biophys Acta*. 2007 Mar;1770(3):390-401. doi: 10.1016/j.bbagen.2006.07.005, PMID 16920266.
37. Ghosh D, Lo J, Egbuta C. Structure, function and inhibition of aromatase. *Resistance to Targeted Anti-Cancer Therapeutics*. 2015:33-61. doi: 10.1007/978-3-319-17972-8\_3. (Resistance to Targeted Anti-Cancer Therapeutics; vol. 8).
38. Li JJ. Imide containing synthetic drugs. Elsevier; 2019p. p. 353-66.
39. Cocconi G. First generation aromatase inhibitors-aminoglutethimide and testololactone. *Breast Cancer Res Treat*. 1994;30(1):57-80. doi: 10.1007/BF00682741, PMID 7949205.
40. Ghuge RB, Murumkar PR, Choudhary KM, Joshi KD, Chauhan M, Barot RR, *et al.* Development of steroidal aromatase inhibitors as potential anti-breast cancer agents. *Curr Enzyme Inhib*. 2020 May 4;16(1):45-62. doi: 10.2174/1573408016666200212094804.
41. Vardanyan RS, Hruby VJ. Soporific agents (hypnotics and sedative drugs). In: *Synthesis of essential drugs*. Elsevier; 2006p. p. 57-68.
42. Vardanyan RS, Hruby VJ. Antineoplastics: Synthesis of essential drugs. Elsevier; 2006p. p. 389-418.
43. Cepa MMDS, Tavares da Silva EJ, Correia-da-Silva G, Roleira FMF, Teixeira NAA. Structure-activity relationships of new A, D-ring modified steroids as aromatase inhibitors: design, synthesis, and biological activity evaluation. *J Med Chem*. 2005 Oct;48(20):6379-85. doi: 10.1021/jm050129p, PMID 16190763.
44. Miller WL, Achermann JC, Flück CE. The adrenal cortex and its disorders. *J Pediatr Endocrinol*. 2008p:444-511.
45. Fried J, Thoma RW, Klingsberg A. Oxidation of steroids by microorganisms. III. Side chain degradation, ring d-cleavage and dehydrogenation in ring A. *J Am Chem Soc*. 1953 Nov;75(22):5764-5. doi: 10.1021/ja01118a530.
46. Zinczuk J, Bacigaluppo JA, Colombo MI, Cravero RM, González-Sierra M, Rúveda EA. An efficient and environmentally benign chemical synthesis of testolactone. *J Braz Chem Soc*. 2003 Dec;14(6):19-4. doi: 10.1590/S0103-50532003000600013.
47. Jordan VC, Brodie AMH. Development and evolution of therapies targeted to the estrogen receptor for the treatment and prevention of breast cancer.



- Steroids. 2007 Jan;72(1):7-25. doi: 10.1016/j.steroids.2006.10.009, PMID 17169390.
48. Bhatnagar AS. The early days of letrozole. *Breast Cancer Res Treat.* 2007 Oct;105;Suppl 1:3-5. doi: 10.1007/s10549-007-9699-0, PMID 17912632.
  49. Pérez Carrión R, Alberola Candel V, Calabresi F, Michel RT, Santos R, Delozier T, et al. Comparison of the selective aromatase inhibitor formestane with tamoxifen as first-line hormonal therapy in postmenopausal women with advanced breast cancer. *Ann Oncol.* 1994;5;Suppl 7:S19-24. PMID 7873457.
  50. Wiseman LR, McTavish D. Formestane. A review of its pharmacodynamic and pharmacokinetic properties and therapeutic potential in the management of breast cancer and prostatic cancer. *Drugs.* 1993 Jan;45(1):66-84. doi: 10.2165/00003495-199345010-00007, PMID 7680986.
  51. Martin GDA, Narvaez J, Marti A. Synthesis and bioconversions of formestane. *J Nat Prod.* 2013 Oct 25;76(10):1966-9. doi: 10.1021/np400585t, PMID 24074257.
  52. Buzdar AU. Anastrozole (Arimidex) in clinical practice versus the old 'gold standard', tamoxifen. *Expert Rev Anticancer Ther.* 2002 Dec;2(6):623-9. doi: 10.1586/14737140.2.6.623, PMID 12503208.
  53. Buzdar A, Howell A. Advances in aromatase inhibition: Clinical efficacy and tolerability in the treatment of breast cancer. *Clin Cancer Res.* 2001;7(9):2620-35. PMID 11555572.
  54. Kabelik D, Kelly AM, Goodson JL. Dopaminergic regulation of mate competition aggression and aromatase-Fos colocalization in vasotocin neurons. *Neuropharmacology.* 2010 Jan;58(1):117-25. doi: 10.1016/j.neuropharm.2009.06.009, PMID 19540858.
  55. Kelly CM, Buzdar AU. Aromatase inhibitors alone or in sequence with tamoxifen - Clinical Evaluation of the BIG 1-98 trial: Evaluation of 'letrozole alone or in sequence with tamoxifen for postmenopausal women with breast cancer', *N Engl J Med.* *Expert Opin Pharmacother* 2010 Feb. 2009;36(11):489-92:766-76.
  56. Sanford M, Plosker GL. Anastrozole: A review of its use in postmenopausal women with early-stage breast cancer. *Drugs.* 2008;68(9):1319-40. doi: 10.2165/00003495-200868090-00007, PMID 18547136.
  57. Johnson DS, Li JJ, editors. *Front matter.* In: Hoboken, NJ: John Wiley and Sons, Inc. p. i-xv; 2007. The art of drug synthesis [internet] [cited Aug 23 2020]. Available from: <http://doi.wiley.com/10.1002/9780470134979.fmatter>.
  58. Lamb HM, Adkins JC. Letrozole. A review of its use in postmenopausal women with advanced breast cancer. *Drugs.* 1998;56(6):1125-40. doi: 10.2165/00003495-199856060-00020, PMID 9878997.
  59. Breast International Group (BIG) 1-98 Collaborative Group, Thürlimann B, Keshaviah A, Coates AS, Mouridsen H, Mauriac L, Forbes JF, et al. A comparison of letrozole and tamoxifen in postmenopausal women with early breast cancer. *N Engl J Med.* 2005 Dec 29;353(26):2747-57. doi: 10.1056/NEJMoa052258, PMID 16382061.
  60. Miller WR, Telford J, Love C, Leonard RCF, Hillier S, Gundacker H, et al. Effects of letrozole as primary medical therapy on *in situ* oestrogen synthesis and endogenous oestrogen levels within the breast. *Breast.* 1998 Oct;7(5):273-6. doi: 10.1016/S0960-9776(98)90095-9.
  61. Dellapasqua S, Colleoni M. Letrozole. *Expert Opin Drug Metab Toxicol.* 2010 Feb;6(2):251-9. doi: 10.1517/17425250903540246, PMID 20095792.
  62. Goss PE, Ingle JN, Martino S, Robert NJ, Muss HB, Piccart MJ, et al. A randomized trial of letrozole in postmenopausal women after five years of tamoxifen therapy for early-stage breast cancer. *N Engl J Med.* 2003 Nov 6;349(19):1793-802. doi: 10.1056/NEJMoa032312, PMID 14551341.
  63. Johnson DS, Li JJ, editors. *The art of drug synthesis.* New York: Wiley Interscience; 2007. 276 p.
  64. Santen RJ, Yue W, Naftolin F, Mor G, Berstein L. The potential of aromatase inhibitors in breast cancer prevention. *Endocr Relat Cancer.* 1999 Jun;6(2):235-43. doi: 10.1677/erc.0.0060235, PMID 10731115.
  65. Miller WR. Aromatase inhibitors: Prediction of response and nature of resistance. *Expert Opin Pharmacother.* 2010 Aug;11(11):1873-87. doi: 10.1517/14656566.2010.487863, PMID 20497094.
  66. Ma CX, Reinert T, Chmielewska I, Ellis MJ. Mechanisms of aromatase inhibitor resistance. *Nat Rev Cancer.* 2015 May;15(5):261-75. doi: 10.1038/nrc3920, PMID 25907219.
  67. Fribbens C, Garcia Murillas I, Beaney M, Hrebien S, O'Leary B, Kilburn L, et al. Tracking evolution of aromatase inhibitor resistance with circulating tumour DNA analysis in metastatic breast cancer. *Ann Oncol.* 2018 Jan;29(1):145-53. doi: 10.1093/annonc/mdx483, PMID 29045530.
  68. Miller WR, Larionov AA. Understanding the mechanisms of aromatase inhibitor resistance. *Breast Cancer Res.* 2012 Feb;14(1):201. doi: 10.1186/bcr2931, PMID 22277572.
  69. Clarke R, Liu MC, Bouker KB, Gu Z, Lee RY, Zhu Y, et al. Antiestrogen resistance in breast cancer and the role of estrogen receptor signaling. *Oncogene.* 2003 Oct;22(47):7316-39. doi: 10.1038/sj.onc.1206937, PMID 14576841.
  70. Milano A, Dal Lago LD, Sotiriou C, Piccart M, Cardoso F. What clinicians need to know about antioestrogen resistance in breast cancer therapy. *Eur J Cancer.* 2006 Nov;42(16):2692-705. doi: 10.1016/j.ejca.2006.06.022, PMID 16963260.
  71. Geisler J, Lønning PE. Resistance to endocrine therapy of breast cancer: Recent advances and tomorrow's challenges. *Clin Breast Cancer.* 2001 Jan;1(4):297-308; discussion 309. doi: 10.3816/CBC.2001.n.004, PMID 11899352.
  72. Johnston SRD, Martin LA, Leary A, Head J, Dowsett M. Clinical strategies for rationale combinations of aromatase inhibitors with novel therapies for breast cancer. *J Steroid Biochem Mol Biol.* 2007 Aug;106(1-5):180-6. doi: 10.1016/j.jsbmb.2007.05.019, PMID 17624764.
  73. Miller WR, Bartlett J, Brodie AMH, Brueggemeier RW, Di Salle E, Lønning PE, et al. Aromatase inhibitors: Are there differences between steroidal and non-steroidal aromatase inhibitors and do they matter? *Oncologist.* 2008 Aug;13(8):829-37. doi: 10.1634/theoncologist.2008-0055, PMID 18695261.
  74. Lønning PE. Pharmacology of new aromatase inhibitors. *The Breast.* 1996;5(3):202-8. doi: 10.1016/S0960-9776(96)90094-6.
  75. Goldhirsch A, Leuvenberger U, Hormonrezeptoren CF, Mammakarzinom, Rundsch G-G. 1981;21(1):18-23.
  76. Wong C, Chen S. The development, application and limitations of breast cancer cell lines to study tamoxifen and aromatase inhibitor resistance. *J Steroid Biochem Mol Biol.* 2012 Sep;131(3-5):83-92. doi: 10.1016/j.jsbmb.2011.12.005, PMID 22265958.
  77. Fabian CJ. The what, why and how of aromatase inhibitors: hormonal agents for treatment and prevention of breast cancer. *Int J Clin Pract.* 2007 Dec;61(12):2051-63. doi: 10.1111/j.1742-1241.2007.01587.x, PMID 17892469.
  78. Osborne CK. Aromatase inhibitors in relation to other forms of endocrine therapy for breast cancer. *Endocr Relat Cancer.* 1999 Jun;6(2):271-6. doi: 10.1677/erc.0.0060271, PMID 10731120.
  79. Ingle JN. Endocrine therapy trials of aromatase inhibitors for breast cancer in the adjuvant and prevention settings. *Clin Cancer Res.* 2005;11(2 Pt 2):900s-5s. PMID 15701884.
  80. Pistelli M, Mora AD, Ballatore Z, Berardi R. Aromatase inhibitors in premenopausal women with breast cancer: The state of the art and future prospects. *Curr Oncol.* 2018;25(2):e168-75. doi: 10.3747/co.25.3735, PMID 29719441.
  81. Anastrozole alone or in combination with tamoxifen versus tamoxifen alone for adjuvant treatment of postmenopausal women with early breast cancer: first results of the ATAC randomised trial. *Lancet.* 2002 Jun;359(9324):2131-9. doi: 10.1016/S0140-6736(02)09088-8.
  82. Lumachi F, Brunello A, Maruzzo M, Basso U, Basso SM. Treatment of estrogen receptor-positive breast cancer. *Curr Med Chem.* 2013 Jan 16;20(5):596-604. doi: 10.2174/092986713804999303, PMID 23278394.
  83. Johnston SRD, Dowsett M. Aromatase inhibitors for breast cancer: lessons from the laboratory. *Nat Rev Cancer.* 2003 Nov;3(11):821-31. doi: 10.1038/nrc1211, PMID 14668813.
  84. Fuqua SA, Cui Y. Estrogen and progesterone receptor isoforms: Clinical significance in breast cancer. *Breast Cancer Res Treat.* 2004;87;Suppl 1:S3-10. doi: 10.1007/s10549-004-1577-4, PMID 15597215.
  85. Lal P, Tan LK, Chen B. Correlation of HER-2 status with estrogen and progesterone receptors and histologic features in 3,655 invasive breast carcinomas. *Am J Clin Pathol.* 2005 Apr;123(4):541-6. doi: 10.1309/YMJ3-A83T-B39M-RUT9, PMID 15743737.
  86. Johnston SRD, Martin LA, Head J, Smith I, Dowsett M. Aromatase inhibitors: Combinations with fulvestrant or signal transduction inhibitors as a strategy to

- overcome endocrine resistance. *J Steroid Biochem Mol Biol.* 2005 May;95(1-5):173-81. doi: 10.1016/j.jsmb.2005.04.004, PMID 15996863.
87. Rothenberger NJ, Somasundaram A, Stabile LP. The role of the estrogen pathway in the tumor microenvironment. *Int J Mol Sci.* 2018 Feb 19;19(2). doi: 10.3390/ijms19020611, PMID 29463044.
88. Koyuturk M, Ersoz M, Altioek N. Simvastatin induces apoptosis in human breast cancer cells: p53 and estrogen receptor independent pathway requiring signalling through JNK. *Cancer Lett.* 2007 Jun;250(2):220-8. doi: 10.1016/j.canlet.2006.10.009, PMID 17125918.
89. Miller WR, Larionov A. Changes in expression of oestrogen regulated and proliferation genes with neoadjuvant treatment highlight heterogeneity of clinical resistance to the aromatase inhibitor, letrozole. *Breast Cancer Res.* 2010;12(4):52. doi: 10.1186/bcr2611.

## PICTORIAL ABSTRACT



## SUMMARY

- One of the best ways to block synthesis of estrogen is inhibition of aromatase enzyme, as it plays key role in rate limiting step.
- It also be used for ovulation induction in female infertility and gynaecomastia.
- The success of aromatase inhibitor, developing small biologically active molecule with lower side effects and better tolerability are in process.
- This review aims are facilitating more readers about development of selective, novel, potential chemical entities.

## About Authors



**Dr. Prafulla M. Sabale** is a Professor at Department of Pharmaceutical Sciences and currently working as The Director, Board of Examination and Evaluation, Rashtrasant Tukadoji Maharaj Nagpur University, Nagpur. He is having total 23 years of experience in teaching, research and administration. Dr Sabale is a Central Council member of Pharmacy Council of India, New Delhi, nominated u/s. 3(a) by the University Grants Commission. He has completed his M. Pharm and Ph.D. from Pharmacy Department, The M. S. University of Baroda, Vadodara. He has received grants from AICTE, UGC, GUJCOST and URPS. He was the recipient of Young Gandhian Technological Award in the year 2011. To his credit Dr Sabale has several research papers in various journals of national and international repute. He has been granted one Indian Patent and filed four more patents. He is life member of several professional bodies such as FIC, APTI, IPA, ISTE, SPER, ISCA, NSI etc.



**Ms. Nusrat B Sayyad** has completed her master's degree from National Institute of Pharmaceutical Education and Research, Hyderabad. Currently she is pursuing Ph D at Department of Pharmaceutical Sciences, RTM Nagpur University, Nagpur, Maharashtra, India. she has attended national and international conferences and also published research and review articles in the field of medicinal chemistry. Her area of research interest is Heterocyclic Chemistry, green synthesis and molecular modelling studies.



**Mr. Mohit D Umare** has completed his master's degree from Department of Pharmaceutical Sciences, RTM Nagpur University, Nagpur, Maharashtra, India. Presently, he is working as senior process associate in Tata consultancy services, Pune. He has attended national and international conferences and also published research articles in the field of medicinal chemistry.



**Ms. Komal K Bajaj** has completed her master's degree from Department of Pharmaceutical Sciences, RTM Nagpur University, Nagpur, Maharashtra, India. Presently, she is working as senior process associate in Tata consultancy services, Pune. She has attended national and international conferences and also published research articles in the field of medicinal chemistry.

**Cite this article:** Sayyad NB, Sabale PM, Umare MD, Bajaj KK. Aromatase Inhibitors: Development and Current Perspectives. *Indian J of Pharmaceutical Education and Research.* 2022;56(2):311-20.