Antiobesity Potential of Fresh Cow Urine and its Distillate - A Biomedicine for Tomorrow

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ABSTRACT
Background: The cow urine has been used traditionally for the management of many diseases and also used as bio-enhancer to improve their therapeutic effect. Objective: To prepare and evaluate the anti-obesity potential of distillate cow urine and compare with fresh cow urine against high fat diet induced obesity in Wistar rats. Materials and Methods: Qualitative analysis of fresh cow urine and its distillate was done and used for the evaluation of assessment of anti-obesity parameters like BMI, abdominal circumference, obesity index, atherogenic index, lipid profile analysis and histopathological evaluation after two months daily oral treatment against high fat diet induced Obesity. Results: The treated groups with fresh cow urine and its distillate has reduced BMI, abdominal circumference, obesity index, atherogenic index, total cholesterol, triglycerides, LDL-C and VLDL-C Significantly while increased the levels of HDL-C as compared with control group \((P<0.05)\). The Histopathological evaluation revealed the size of visceral white adipose tissue in treated groups was reduced as compared with control Group. Conclusion: In the present research conclude that fresh cow urine and its distillate both are having significant anti-obesity activity against high fat diet induced obesity in Wistar rats.

key words: Cow urine, Distillated Cow urine, Anti-obesity, BMI, Atherogenic inde.

INTRODUCTION

As per World Health Organization (WHO), obesity is a comprehensive hazardous disorder worldwide, with respect to the data analysis of body mass index (BMI). The incidence of obesity increased at a tormenting mark and is the alarming situation for a major public health concern. However, obesity develops metabolic disorders like diabetes mellites, high blood pressure as well as cardiac diseases along with some chronic problems such as sleep apnoea, osteoarthritis, tumour’s, stroke in addition to inflammation-based pathologies.1 In the present scenario, researchers focused on the therapeutic potential of natural products use for treatment and counteract obesity along with its complications with negligible adverse effects. From past several years various medicines have been using to manage obesity. Though, maximum anti-obesity drugs which were permitted and promoted have now been inhibited due to serious side effects.2

From the olden time, Cow’s Urine have been using as a remedy. In Ayurveda, Cow’s Urine is having its unique role in treatment of various diseases. It’s having an immeasurable therapeutic value as well as described in ‘Ashtanga Sangraha’ and ‘Sushruta Samhita’ as the utmost useful ingredient/secretion of animal source. It has been known as the water of life or beverages of immortality, the nectar of the God. In India, using of Cow’s Urine in drink has been adept for several years. Ayurvedic medicine practitioners regularly use cow’s urine as a medicine and the medications prepared from it are used as therapy for numerous illnesses. Perfections have been revealed and described...
in those suffering from flu, aging, tuberculosis, allergies, leucorrhoea, colds, skin infections, rheumatoid arthritis, chicken pox, bacterial/viral infections, hepatitis, leprosy, ulcer, cardiac disease, asthma, chemical intoxication etc. Cow’s Urine can be used in many therapy for treatment of numerous treatable and allegedly hopeless illnesses and cow sourced products have multiple roles and benefits.\textsuperscript{3,4} Cow’s urine can be used to kill the lots of drugs resistant bacteria as well as virus. As per previous literature the cow’s urine has been approved US patent (6896907 and 6410059) for its therapeutic properties, for its use along with antibiotics against bacterial infection along with fights against several types of cancer.\textsuperscript{5,6} nowadays several AIDS patients are attracting towards cow’s urine therapy due to its positive outcomes.

The analytical data of Cow’s Urine has revealed that it comprises Vitamin A, B, C, D, E, minerals, carboacid, chlorine, nitrogen, manganese, creatine, hormones, magnesium, sulphur, iron, tartaric acid, phosphate, sodium, silicon, citric acid, succinic acid, calcium salts, lactose, enzymes, and gold. In deficiency or excess of these constituents inside the body can make an ill to the individual. Cow’s urine comprises these substances that are naturally existing in the human body. Thus, ingesting of cow’s urine sustains the equilibrium of these substances and this supports to cure incurable illnesses.\textsuperscript{7,8}

Based on extensive literature survey and patent search state that there is no scientific data explored on the anti-obesity activity of cow’s urine and its distillate, hence, the study delineated with the anti-obesity property of cow’s urine and its distillate in experimentally induced obesity in Wistar rats.

**MATERIALS AND METHODS**

**Animals**

Young Wistar rats of either sex, weighing approximately 120–150 g was housed in room temperature of $25 \pm 1^\circ C$ and 12 h light and dark cycles. Animals were fed on a standard chow diet (Nutrivet Life sciences, Pune, India) and water ad libitum. The experimental protocol was approved by the Institutional Animal Ethics Committee (IAEC) and carried out according to the guidelines given by Committee for Control and Supervision of Experiments on Animals (CPCSEA), New Delhi, India CPCSEA Approval No: SPTM-IAEC/2016-2017/01/11, (Registration No: 1300/ac/09/CPCSEA).

**Drugs and Chemicals**

Diagnostic kits for Total cholesterol, HDL, Triglycerides purchased from Jinendra scientific, Jalgaon, Maharashtra, India and Lova Chemicals, Mumbai, Maharashtra, India.

**Collection of FCU**

FCU was collected personally from Maa Kanakeshwari Goshala, Bombay-Agra Road, and Shirpur-425 405.

**Preparation of DCU**

DCU was prepared by the simple distillation process. FCU was boiled in a round bottom flask to which a vapor condensing device was attached. The vapors through tube were collected in a pot put over cold water. The DCU was kept in a closed amber colored container and stored in cool place.

**Elemental analysis of FCU and DCU**

**Preparation of Lassaigne’s Solution (Sodium Filtrate)**

Introduced a freshly cut, clean and dry piece of sodium metal into an ignition tube, which was held in vertical position with a pair of the thong. The lower part of the tube was heated gently till sodium melted to a shining globule. Removed the ignition tube from the flame and added a small amount of FCU. Heated the tube on the flame till the reaction ceased and the tube became red hot. Now plunged the tube into about 20 ml of distilled water in a china dish and broken the bottom of the tube by striking against the dish. The contents were heated to boiling for the few min. It was then cooled and filtered to get a colorless alkaline filtrate (Lassaigne’s solution). This solution was also prepared for DCU.

**Test for nitrogen in FCU and DCU**

The Lassaigne’s solution (2 ml) was taken in a test tube and added some drops of freshly prepared aqueous ferrous sulfate solution. Formation of the dark green color of ferrous hydroxide was formed indicated the presence of nitrogen. The mixture was heated to boiling, cooled and acidified with Sulphuric acid. Formation of Prussian blue color shows the existence of nitrogen.

**Test for sulfur in FCU and DCU**

The Lassaigne’s solution (2 ml) added 2-3 drops of freshly prepared sodium nitroprusside solution. The violet colour indicates the presence of sulfur.

**Test for chloride in FCU and DCU**

The Lassaigne’s solution was acidified with dilute nitric acid and added a few drops of silver nitrate solution, the formation of white precipitate indicates the presence of nitrogen.

**Qualitative Examination of Inorganic and Organic Matters in FCU and DCU**\textsuperscript{9,11,12}

It involved qualitative examinations of inorganic (those, which are free from carbon) and organic matters (may be secondary metabolites products in the cow’s urine). The role of these components for the medicinal
purpose is important, hence detected by simple chemical analysis. The test methods of main components are as given in Table 1.

**Determination of pH and Specific Gravity of FCU and DCU**

The procedure of pH Determination

The glass electrode was washed with distilled water gently before the operation and calibrated with standard solutions of different pH (4, 7 and 9) and the pH of samples of FCU and DCU were recorded.

The procedure of Specific Gravity Determination

The weighing bottle was weighed and then a fixed amount of sample of FCU and DCU were filled in weighing bottle and weighed again. After this, the empty bottle was again weighed and then the specific gravity was calculated.

**Acute toxicity studies (as per OECD guidelines)**

The study was carried out as per OECD guidelines and received draft 423 guidelines from the committee CPCSEA, Government of India.

**High-Fat Diet Formula (HFD)**

It consists of fat 58%, protein 25% and carbohydrate 17%, lard 13%, cholesterol 1%, vitamin, and minerals 0.6% as a percentage of total kcal ad libitum, the animals selected for the study were fed with this composition for a period of two months.

**Pharmacological assessment**

**Experimental design**

After one week of acclimatization, the animals were separated into six groups each containing six animals,

**Group I:** Received only normal diet without any treatment.

**Group II:** Received HFD without any treatment.

**Group III:** Received HFD and daily treatment of 1 ml/kg of body weight with FCU.

**Group IV:** Received HFD and daily treatment of 2 ml/kg of body weight with FCU.

**Group V:** Received HFD and daily treatment of 1 ml/kg of body weight with DCU.

**Group VI:** Received HFD and daily treatment of 2 ml/kg of body weight with DCU.

The various parameters like BMI, AC, OI, TC, TG, HDL-C, LDL-C, VLDL-C and AI were evaluated at the starting and after completion of 2 months’ treatment of FCU and DCU according to the groups mentioned above.

**Morphological Parameters to measure obesity**

**Evaluation of BMI, Abdominal Circumference and Obesity index**

The BMI, AC and Obesity index are the major parameters to be evaluated in the assessment of anti-obesity drug. In the present study, BMI was evaluated by following formula

\[
\text{BMI} = \frac{\text{body weight (g)}}{\text{length}^2 (\text{cm}^2)}
\]

Abdominal circumference was measured using a measuring tape. The obesity index was determined by formula,

\[
\text{Obesity index} = \text{Cube root of body weight of rat (g)/nasoanal length (mm)} \times 10^4
\]

**Evaluation of Lipid Profile**

On the 1st day and 60th day of the experiment, the blood samples were withdrawn from retro-orbital plexus of all the animals into sterilized dry Eppendorf tubes then allowed to stand for 30 to 40 min at room temperature. The clear serum was separated at 3000 rpm for 15 min by centrifugation. The levels of serum TC, HDL-C, TG were determined using semi-auto analyser (Erba), according to the procedure given in respective diagnostic kits. To calculate serum LDL-C and VLDL-C values Friedewald formula was used.

\[
\text{LDL-C} = \text{TC} - (\text{HDL-C} + \text{TG}/5)
\]

\[
\text{VLDL-C} = \text{TG}/5
\]

Atherogenic index (AI).

The AI was calculated by the following formula.

\[
\text{AI} = \frac{\text{(Total cholesterol – HDL-C)}}{\text{HDL-C}}
\]

**Histopathological evaluation**

At the termination of the experimental period, rats were sacrificed by spinal dislocation. The white adipose tissues were fixed in 10% formalin solution and embedded in paraffin were subjected to histopathological evaluation. Standard sections of 5 μm thickness were cut, stained using hematoxylin and eosin and examined under an optical microscope (40X).

**Statistical Analysis**

All outcomes are expressed as the mean ± SEM. The results were evaluated for statistical significance using ANOVA and Dunnett’s test. P values <0.05 were measured as significant (Using prism software 5.2 version).

**RESULTS**

**Elemental analysis of FCU and DCU**

The elemental analysis was performed as per the procedures mentioned in above section of materials and methods and the results are expressed in Table 2.

**Qualitative analysis of FCU and DCU**
Table 1: Qualitative test for chemical composition of FCU and DCU

<table>
<thead>
<tr>
<th>COMPONENT</th>
<th>TEST</th>
<th>OBSERVATION</th>
</tr>
</thead>
<tbody>
<tr>
<td>Urea</td>
<td>(UREASE TEST) Sample and added Soya bean meal with Phenol red</td>
<td>Formation of red color indicate presence of urea.</td>
</tr>
<tr>
<td>Potassium</td>
<td>3 ml sample and added few drops of sodium cobalt nitrite solutions</td>
<td>Formation of yellow precipitate indicate presence of potassium.</td>
</tr>
<tr>
<td>Calcium</td>
<td>Sample and added Amino oxalate</td>
<td>Formation of white precipitate indicate presence of calcium.</td>
</tr>
<tr>
<td>Iron</td>
<td>5 ml of test solution and added few drops of 2% potassium Ferro cyanide</td>
<td>Formation of dark blue coloration indicate presence of calcium.</td>
</tr>
<tr>
<td>Phosphorus</td>
<td>Sample and added Conc. Nitric acid</td>
<td>Formation of white precipitate indicate presence of phosphorus.</td>
</tr>
<tr>
<td>Bi-carbonate</td>
<td>3 ml of urine and added dilute HCL</td>
<td>Effervescence produce which confirms presence of bi-carbonates</td>
</tr>
<tr>
<td>Sulphate</td>
<td>Sample and added BaCl₂</td>
<td>Formation of white precipitate indicate presence of sulphate</td>
</tr>
<tr>
<td>Bile pigment</td>
<td>5 ml of sample and added Dissolved crystal of sucrose, added 3 ml of conc. Sulphuric acid</td>
<td>Formation of reddish purple ring indicate presence of bile pigment.</td>
</tr>
<tr>
<td>Carbohydrate</td>
<td>(MOLISH TEST) Sample and added molish reagent, added conc. sulphuric acid</td>
<td>Formation of violet ring formed at the junction indicate presence carbohydrate.</td>
</tr>
<tr>
<td>Protein</td>
<td>(HELLEr'S TEST) 3 ml of sample and added 3 ml conc. Nitric acid</td>
<td>Formation of white precipitate formed at the junction indicate presence of protein.</td>
</tr>
<tr>
<td>Ketone bodies</td>
<td>(POTHER'S TEST) 5 ml of sample, added Saturated with solid ammonium sulphate and added 2-3 drop of 5% solution of sodium nitroprusside, added 2 ml conc. Ammonia</td>
<td>No permanent colour was formed which indicate presence of ketene bodies.</td>
</tr>
<tr>
<td>Creatinine</td>
<td>(JAFFE'S TEST) 5 ml of sample and added 2 ml saturated picric acid with 10% NaOH</td>
<td>Formation of Deep orange colour indicate presence of creatinine</td>
</tr>
<tr>
<td>Ammonia</td>
<td>Take 5 ml of FCU or DCU added on red litmus paper</td>
<td>Litmus paper turned to blue indicate presence of ammonia.</td>
</tr>
<tr>
<td>Uric acid</td>
<td>(SCHIFF'S TEST) Moisten a strip of filter paper with Silver nitrate solution and added to it a drop of urine</td>
<td>Formation of black or yellow brown strain indicate presence of uric acid.</td>
</tr>
<tr>
<td>Tartaric acid</td>
<td>2-3 ml of test solution, added one drop dilute NH₄OH and excess 5% AgNO₃ solution, boiled for 15 min. on water bath</td>
<td>Formation of white gelatinous precipitate indicate presence of tartaric acid.</td>
</tr>
<tr>
<td>Oxalic acid</td>
<td>2 ml sample and added few drops 5% lead acetate</td>
<td>Formation of white precipitate indicate presence of oxalic acid.</td>
</tr>
</tbody>
</table>

Table 2: Results of element detection tests for FCU and DCU

<table>
<thead>
<tr>
<th>Preparations</th>
<th>Test for elements</th>
<th>Observation</th>
<th>Inference</th>
</tr>
</thead>
<tbody>
<tr>
<td>FCU</td>
<td>Nitrogenurine(Garnvati)ion</td>
<td>Blue color</td>
<td>(+)</td>
</tr>
<tr>
<td></td>
<td>Sulphur</td>
<td>violet color</td>
<td>(+)</td>
</tr>
<tr>
<td></td>
<td>Chloride</td>
<td>White precipitate</td>
<td>(+)</td>
</tr>
<tr>
<td>DCU</td>
<td>Nitrogenurine(Garnvati)ion</td>
<td>Blue color</td>
<td>(+)</td>
</tr>
<tr>
<td></td>
<td>Sulphur</td>
<td>violet color</td>
<td>(+)</td>
</tr>
<tr>
<td></td>
<td>Chloride</td>
<td>No White precipitate</td>
<td>(-)</td>
</tr>
</tbody>
</table>
The pH and specific gravity of FCU and DCU were determined and expressed in the Table 4.

**Acute Toxicity Study**

The acute toxicity study was done as per OECD guidelines (423). The FCU and DCU were administered orally to test animals at different doses maximum up to 10 ml/kg and observed for 24 h and no mortality, abnormal sign and symptoms were seen for one week.

**Evaluation of BMI, Abdominal Circumference, and Obesity index before the treatment (initial)**

The BMI, abdominal circumference and obesity index were evaluated before starting the treatment on the initial day. All the values of these parameters of normal and treated groups were statistically insignificant as compared with the control group, except BMI and obesity index of the treated group. However, the values of con-
trol groups were less and values of treated groups were high, it was indicating that the BMI and obesity index of the control group were smaller than other treated groups. Results are given in Table 5.

**Evaluation of Lipid Profile and Atherogenic index (AI) before the treatment (initial)**

The serum lipid profile (TC, TG, HDL-C, LDL-C, and VLDL-C) and atherogenic index were evaluated before starting the treatment on the initial day. All the values of these parameters of normal and treated groups were statistically insignificant as compared with the control group. Results are given in Table 6.

**Evaluation of BMI, Abdominal Circumference, and Obesity index after two months treatment**

The BMI, abdominal circumference and obesity index were evaluated after two months’ treatment on the 60th day. The results of this study revealed that BMI, abdominal circumference, and obesity index of the control group were increased to a maximum level at the same time treatment of FCU and DCU decreased the values of same parameters significantly, except obesity index of the group treated with low dose of FCU. Results are given in Table 7.

**Evaluation of Lipid Profile and Atherogenic index (AI) after two months**

The serum lipid profile (TC, TG, HDL-C, LDL-C, and VLDL-C) and atherogenic index were evaluated after two months’ treatment on the 60th day. All the values of these parameters of normal and treated groups were statistically significant as compared with the control group. Results are given in Table 8.

**Histopathological Evaluation**

Histological examinations of this study have been revealed that the sizes of the white adipocytes were significantly reduced in FCU and DCU treated groups when compared with control group Figure 1. However, the effect of the DCU treated group is greater than FCU treated group.

**DISCUSSION**

As in view of the inadequate accessibility of drugs having anti-obesity effect and considering their adverse effects,
the search remains to discovery active natural product based medicines to fight against obesity. The occurrence of obesity is rapidly rising and the consequences are directly or indirectly leads to the serious and life-threatening metabolic disorders. It has been reported that rats fed a diet containing high fat resulted in distinctive visceral adiposity, dyslipidemia and oxidative stress which are typically associated with obesity.21,22 The HFD induced obesity is known to be the most significant model for the assessment of anti-obesity activity because of its close resemblance of copycatting the usual route of obesity episodes in human beings.23,24

Thus, in the present study high-fat diet was given for 2 months to the Wistar rats for induction of obesity to develop the experimental model to be utilized for the assessment of anti-obesity activity. It is well established that over-consumption food along with the high concentration of fat lead to obesity in several animal's models including rats due to metabolic complications as well as liver dysfunction. It was well documented in many studies that obesity is also related with a dyslipidemia with an increase in the levels of TG, LDL-C, VLDL-c and decrease in the levels of HDL-C.25,26,27 The results and observations of this research showed that rats exposed to a diet containing high fat for 2 months increased the body weight significantly, thus it was confirmed the obese status of the rats fed with HFD.28 The difference in the body weight was observed between the groups fed a HFD and normal diet, however, no substantial difference was detected in the daily food intake of animals. This observation revealed the fact that an increase in body weight was not dependent on the extent of food consumed by the rats but it was a result of consumption of HFD. As stated earlier high-fat diet resulted in lipid abnormalities associated to obesity include an raised serum concentration of fatty acids, TG, LDL-C, VLDL-C, TG and reduction in serum HDL-C.29,30 results of the evaluation of lipid profile from control group of this study revealed similar observations thus here confirmed the establishment of the experimental model for the assessment of anti-obesity activity. This suggest that the fresh cow urine and its distillate may reduce pancreatic lipase activity thus reducing the absorption of dietary fat and regulates lipid profile. In epidemiological studies, BMI is widely considered as a prominent marker for assessment of obesity hence evaluation of the BMI

**Table 7: Evaluation of BMI, Abdominal Circumference and Obesity index (60th day)**

<table>
<thead>
<tr>
<th>Groups</th>
<th>BMI</th>
<th>AC</th>
<th>OI</th>
</tr>
</thead>
<tbody>
<tr>
<td>I Normal</td>
<td>0.63±0.0169</td>
<td>14.58±0.239</td>
<td>0.03±0.00270</td>
</tr>
<tr>
<td>II Control</td>
<td>0.80±0.0263</td>
<td>20.08±0.507</td>
<td>0.03±0.00322</td>
</tr>
<tr>
<td>III FCU (1 ml/kg)</td>
<td>0.67±0.0349**</td>
<td>13.41±0.436**</td>
<td>0.03±0.00624</td>
</tr>
<tr>
<td>IV FCU (2 ml/kg)</td>
<td>0.54±0.0269**</td>
<td>13.33±0.211**</td>
<td>0.03±0.00583**</td>
</tr>
<tr>
<td>V DCU (1 ml/kg)</td>
<td>0.58±0.0112**</td>
<td>14.08±0.300**</td>
<td>0.03±0.00194**</td>
</tr>
<tr>
<td>VI DCU (2 ml/kg)</td>
<td>0.48±0.0234**</td>
<td>13.33±0.279**</td>
<td>0.028±0.000478**</td>
</tr>
</tbody>
</table>

** p<0.05 (significant) when compared the other group with control group by using ANOVA followed by Dunnett’s test. (N=6 values Mean± SEM)

**Table 8: Evaluation of Lipid Profile and Atherogenic index (AI) (60th day)**

<table>
<thead>
<tr>
<th>Groups</th>
<th>Total CH</th>
<th>TG</th>
<th>HDL</th>
<th>VLDL</th>
<th>LDL</th>
<th>AI</th>
</tr>
</thead>
<tbody>
<tr>
<td>I Normal</td>
<td>81.50±1.77</td>
<td>81.58±4.96</td>
<td>51.59±2.5</td>
<td>16.31±0.992</td>
<td>13.60±2.14</td>
<td>0.59±0.0637</td>
</tr>
<tr>
<td>II Control</td>
<td>131.42±4.37</td>
<td>166.21±14.5</td>
<td>32.43±3.61</td>
<td>33.24±2.89</td>
<td>65.74±2.14</td>
<td>3.34±0.526</td>
</tr>
<tr>
<td>III FCU (1 ml/kg)</td>
<td>107.42±4.06**</td>
<td>125.29±5.37**</td>
<td>51.33±3.34**</td>
<td>25.05±1.07**</td>
<td>31.03±3.87**</td>
<td>1.13±0.166**</td>
</tr>
<tr>
<td>IV FCU (2 ml/kg)</td>
<td>91.46±4.06**</td>
<td>108.45±4.39**</td>
<td>54.71±1.81**</td>
<td>21.69±0.878**</td>
<td>15.06±4.65**</td>
<td>0.68±0.0997**</td>
</tr>
<tr>
<td>V DCU (1 ml/kg)</td>
<td>105.98±3.66**</td>
<td>118.57±2.97**</td>
<td>48.25±2.82**</td>
<td>23.71±0.594**</td>
<td>34.01±4.37**</td>
<td>1.23±0.141**</td>
</tr>
<tr>
<td>VI DCU (2 ml/kg)</td>
<td>87.85±1.99**</td>
<td>97.18±4.92**</td>
<td>55.55±1.82**</td>
<td>19.43±0.984**</td>
<td>12.86±1.4**</td>
<td>0.58±0.0322**</td>
</tr>
</tbody>
</table>

** p<0.05 (significant) when compared the other group with control group by using ANOVA followed by Dunnett’s test. (N=6 values Mean± SEM)
was done. There was a significant reduction in BMI, AC and OI of FCU and DCU treated groups were observed as compared to the control group. In addition to its weight reducing the effect, FCU and DCU treated HFD rats showed significantly lowered levels of serum TC, TG, LDL-C, VLDL-C and increased levels of HDL-C. It is stated that obesity, particularly abdominal obesity, is related with dyslipidemia, characterized by raised TG and decreased HDL-C concentrations. TGs are involved in the ectopic gathering of lipid stores in the hepatic part and are linked with a various disorder like metabolic syndrome. VLDL-C carries cholesterol and TG to the tissues, whereas HDL-C is supportive in carrying extra cholesterol to the liver for elimination in the bile. Elevated level of TC and LDL-C in addition to low level of HDL-C are risk factors for coronary artery disease (CAD). Therefore, the atherogenic index was assessed and it was expressively decreased in FCU and DCU treated groups as compared to control group. Extreme growing of adipose tissue results in obesity that includes hyperplasia and hypertrophy. Decrease in body weight gain of HFD-fed rats was escorted by a reduction of body fat stores since treatment with FCU and DCU also expressively decreased the weight and deposition of adipose tissues as compared with that control group. These data confirmed that FCU and DCU have an inhibitory effect on hypertrophy and hyperplasia of white adipose tissue. Results of histopathological evaluation shown in Figure 1, revealed that white adipose tissues size were increased in control group whereas decreased in groups treated with FCU and DCU and it was like the normal group. Therefore, FCU and DCU both are reducing the deposition of white adipose tissues and prevents obesity against high-fat diet. The outcomes of the current study supporting the lipid-lowering activity of DCU shown in guinea pig.

CONCLUSION
As per the results of the current research work, it has been determined that the treatment of FCU and DCU at doses 1.0 ml/kg and 2 ml/kg shown significant anti-obesity in HFD-induced obese rats. Further, research is required to be done to determine the different fraction and active principles and components of the FCU and DCU, followed by the establishment of a molecular mechanism of action for anti-obesity potential. Present research highlights the anti-obesity potential of fresh cow urine and Distillated cow urine, the strategies for the profit of mankind with the understanding that cow urine therapy desires instant attention, advancement, and extensive acceptance and appropriate support of the expert, scientists and clinicians to support alternative low-cost therapy having no or insignificant adverse effect.

ACKNOWLEDGEMENT
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CONFLICT OF INTEREST
There is no conflict of interest.

ABBREVIATIONS USED

REFERENCES
SUMMARY

Obesity has established significant attention as a major health impediments. In present scenario the management of obesity is a big challenge which reinforces the research to find out a new, suitable, safe and economical therapy. Traditionally, cow urine has been used for the management of many diseases and disorders. In this study cow urine and its distillate used for assement of Anti-obesity potential. The parameters have been estimated in High fat diet induced obese Wistar rats such as BMI, abdominal circumference, obesity index, atherogenic index, lipid profile along with histological evaluation of white adipose tissues. The results of present research, have successfully proved that cow urine and its distillate can cure obesity. Thus, it can be concluded that the cow urine and its distillate have anti-obesity potential, which supports its traditional claim. Further, studies will be carried out to determine the Bio active fraction of cow urine and its distillate along with probable mechanism of action.
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Ms. Sravani Karri: Lecturer (Pharmacology), NMIMS, SPTM, Shirpur, Maharashtra. Area of interest: Neuropharmacology, Clinical Trials and Basic Pharmacological Research.