

The Interplay between Traumatic Brain Injury, Cocaine Abuse, and Genetic Mutations in the Development of Parkinsons Disease: A Review

Mohamad Qutboddin^{1,2}, Syed Sagheer Ahmed^{1,*}, Nahid Abbas³, Mohammad Ali¹, Bharathi Doddla Raghunathanaidu¹

¹Department of Pharmacology, Faculty of Pharmacy, Sri Adichunchanagiri College of Pharmacy, Adichunchanagiri University, B. G. Nagara, Karnataka, INDIA.

²Department of Pharmacology, Akshaya Institute of Pharmacy, Tumkur, affiliated to Rajiv Gandhi University of Health Science, Bengaluru, Karnataka, INDIA.

³Department of Chemistry, Akshaya Institute of Pharmacy, Tumkur, affiliated to Rajiv Gandhi University of Health Science, Bengaluru, Karnataka, INDIA.

ABSTRACT

Parkinson's Disease (PD) is a neurodegenerative disorder characterized by the depletion of dopamine neurotransmitters and receptors. Although the etiology of PD is multifactorial, emerging evidence suggests that Traumatic Brain Injury (TBI), cocaine abuse, and genetic mutations may contribute to the development and progression of PD. This review summarizes the current understanding of the interplay between TBI, cocaine abuse, and genetic mutations in DA homeostasis in PD. Here, we discuss the molecular mechanisms underlying DA depletion and receptor dysfunction in PD, and how TBI, cocaine abuse, and genetic mutations may exacerbate these processes. Ultimately, a complex relationship between TBI, cocaine abuse, genetic mutations in dopamine, and PD has been identified.

Keywords: Cocaine abuse, Dopamine, Genetic mutation, Parkinson's disease, Traumatic brain injury.

Correspondence:

Dr. Syed Sagheer Ahmed

Department of Pharmacology, Faculty of Pharmacy, Sri Adichunchanagiri College of Pharmacy, Adichunchanagiri University, B. G Nagara-571448, Karnataka, INDIA.
Email: sysaha6835@gmail.com

Received: 16-12-2025;

Revised: 08-01-2026;

Accepted: 23-03-2026.

INTRODUCTION

Parkinson's Disease (PD) is a chronic and progressively debilitating condition that impacts the nervous system. The Reduced in dopamine levels in the basal ganglia results in symptoms such as Involuntary shaking, increased muscle tone, and impaired balance control as the disease advances.

The global rise in PD cases mirrors many aspects of the pandemic, except for the absence of an infectious cause. Only about 3-5% of Parkinson's cases are linked to mutations in identified Parkinson's-related genes, which are classified as genetically inherited Parkinson's disease. Parkinson's has a profound societal impact. Parkinson's is a prevalent disorder, affecting approximately 6.1 million people worldwide in 2016.¹ The incidence of Parkinson's increases sharply with age. Especially in the age group between 50-59; 17.4 cases were reported in the

100,000 population. Whereas 93.1 cases were reported for the age group of 70 to 79 years. The lifetime probability of developing the disease is estimated to be 1.5%.² Parkinson's disease is more prevalent in North America and Europe than in Asia and Africa. The underlying causes of these geographic variations remain unclear, but exposure to environmental factors, is the leading cause of this disease.³ Unlike in the general population, mortality rates in individuals with Parkinson's do not rise within the first five years of disease onset. However, after ten years, the risk of death increases by 3.5 times, according to the reported data. Drug abuse, pesticides, and head trauma are also associated with PD. Parkinson's happens because of genes, the environment, and how people live.⁴ An excess of dopamine can accumulate in the cytoplasm, triggering dopamine auto-oxidation and the production of toxic by-products, which ultimately contribute to neurodegeneration. To prevent this, neurons employ protective mechanisms such as converting cytosolic dopamine into neuromelanin or breaking it down through Monoamine Oxidase (MAO) and Catechol-O-Methyltransferase (COMT) to produce homovanillic acid.⁵ This review describes the interplay between traumatic brain injury, cocaine abuse, and genetic mutations in the development of Parkinson's disease.



DOI: 10.5530/ijper.20262583

Copyright Information :

Copyright Author (s) 2026 Distributed under Creative Commons CC-BY 4.0

Publishing Partner : Manuscript Technomedia. [www.mstechnomedia.com]

Traumatic brain injury

According to Graybiel (1990), Dopaminergic neurons in the brain arise from the Ventral Tegmental Area (VTA) and the Substantia Nigra (SN), projecting their axons to the striatum and cortex, limbic system, and hypothalamus.⁶ DA affects hormone secretion, motor control, motivation, emotion, and cognitive processing, among other physiological processes.⁷ Dopaminergic cell death and biochemical disruption of the dopaminergic system have been observed in animal models of TBI.⁸ The Figure 1 explain about the TBI leading to PD.

Injuries with acute, subacute, and chronic pathologies comprise a diverse and complicated disorder, known as traumatic brain injury.^{9,10} There are two separate sequential phases of brain trauma injury: the first phase and the second phase. Direct physical damage to neurons, glial cells, and vascular structures is linked to instantaneous biomechanical damage known as primary injury. The integrity of the brain parenchyma and Blood-Brain Barrier (BBB) is disrupted by the impact of damage. Chains of events that disrupt brain function cause further structural damage and accelerate cell death. These are manifestations of the secondary phase, which involves cortical impact injury, Drop-weight injury, Fluid Percussion Injury (FPI) and Blast-induced head injury. Twenty-three different injury processes have been developed using various animal injury models.¹¹ Changes in DA activity in TBI may lead to long-lasting cognitive impairment. Numerous animal studies have examined the pathophysiology of DA changes following TBI. Axonal damage leads to Wallerian degeneration, structural impairment, tissue edema, and disruption of the BBB. Disintegration can be caused by the original insult. Ischaemia, excitotoxicity, neuroinflammatory reactions, and the aftereffects of alterations in epigenetic and/or genetic expression are examples of secondary damage.¹²⁻¹⁴ Severe head trauma damage results in the loss of neurons and white matter, as well as the destruction of anatomical structures in the brain. Neurological deficits associated with permanent neuronal circuit damage are also associated with severe injury. Although direct anatomical damage is a major cause of many neurological deficits resulting from brain injury, inhibition of both electrical and chemical signalling also plays a crucial role. Both behavioral and epigenetic studies have examined these changes.^{15,16} All neurotransmitter systems, including Dopaminergic Systems (DAergic), are affected by extensive disruption of neural projections and physical damage to neuronal pathways, such as the nigrostriatal and mesolimbic circuits.¹⁷ In the striatum and FC, synaptic structure and dendritic complexity are altered by the loss of DAergic innervating fibers from the substantia nigra and ventral tegmental region.^{18,19} Reduced substantia nigra volume was linked to anomalies in the caudate DaT, and there was evidence of nigrostriatal tract injury, specifically impairing caudate projections. When combine these findings indicate that TBI frequently affects the dopaminergic system.²⁰ The cytoskeletal protein that stabilizes microtubules is

synthesis by Microtubule-Associated Protein Tau (MAPT), which is present on the chromosome-17.²¹ Tau proteins are involved in axonal transport regulation, membrane binding, and microtubule stabilisation.^{22,23} Tau proteins change shape and can no longer dissolve properly in pathological situations due to alterations in isoforms or phosphorylation patterns, neuronal damage, and axonal transport.^{24,25} In India, the incidence of TBI in both sexes was 7464800 (6383800-8667 200), in males was 4673 100 (3987400-5492900), and in females 2 791 700 (2 368100-3 273 300) were reported 2-1 times variation was observed in all states. Figure 2 illustrating the number of Traumatic Brain Injury (TBI) patients by age groups.

In India in 2019, in both males and females 770800 (635 100-919400) people had Parkinson's disease. Approximately 421800 (348300-502 100) males suffered from PD. Whereas the female count was 349000 (287 200-416500). An estimated 45 300 (95% UI 38 600-52 800) deaths were due to PD. However, a 2-3 times variation between the states was observed. Current data analysis reports that PD was uncommon in younger individuals, with prevalence rising significantly among older age groups, especially in those over 50 years of age in both men and women. The Figure 3 represents this data graphically. Note: Values in parentheses represent 95% uncertainty intervals. Prevalence or incidence is reported according to the metric most commonly applied in clinical practice for each disease.

Cocaine abuse

Cocaine attaches to the dopamine transporter, blocking dopamine reuptake. The extent of this binding is closely associated with self-administration of various stimulants.²⁶ Prolonged cocaine use may lead to the depletion of neurotransmitter reserves, which could explain the 'crash' and craving experience when the drug is discontinued. Supporting this hypothesis, a study of 20 chronic cocaine users found significantly higher prolactin levels (35.4±26.9 ng/mL) compared to the control group (7.0±5.0 ng/mL). Prolactin levels in cocaine users ranged from 5 to 96 ng/mL, with a median of 23 ng/mL. Notably, these increased levels did not fully return to normal within two weeks. This evidence has given rise to the 'dopamine depletion' hypothesis of cocaine addiction.²⁷ In a study involving 21 cocaine-dependent individuals seeking inpatient treatment, Ten patients showed high HDRS scores upon admission, which decreased by 50% or more during their hospital stay, despite the absence of targeted treatment.²⁸ This study revealed dopamine depletion occurrence was observed in cocaine addicts. In individuals who abused cocaine, a notable decline in dopamine D2 receptors was observed, which was linked to reduced metabolic activity in the cingulate gyrus and orbitofrontal cortex. This mechanism contributes to Cocaine disrupts the brain's dopamine system, leading to uncontrollable drug use. Additionally, cocaine users exhibit a significant decrease in dopamine secretion which, when combined with the reduction in D2 receptors, further exacerbates the condition.²⁹

Chronic cocaine use can lead to dopamine depletion due to the prolonged inhibition of dopamine reuptake. Normally, after reabsorption, dopamine is stored in secretory vesicles for reuse. However, when the reuptake is blocked, this recycling process is disrupted, causing the released dopamine to break down in the synapse. Evidence for this is the elevated levels of the synaptic dopamine metabolite, 3-methoxytyramine, observed after cocaine use.³⁰ Additionally, increased tyrosine hydroxylase activity indicates increased dopamine synthesis. However, if this synthesis is insufficient to offset the loss of dopamine in the synapse, dopamine depletion becomes inevitable.³¹ In contrast to the observed decrease dopamine levels in nucleus.³² Phasic dopamine levels declined in both regions as the cocaine intake rate increased. Notably, the reduction in dopamine levels within the ventral medial striatum was significantly correlated with the rate of escalation.³³

Genetic mutation

In Parkinson's, several genes are associated with mutations that contribute to disease development. They are classified into two types based on their mutations.

Autosomal Dominant Mutations

- a) *SNCA* (α -Synuclein),
- b) *LRRK2* (Leucine-Rich Repeat Kinase 2),

Autosomal Recessive Mutations

- a) *PARK2* (Parkin),
- b) *PINK1* (PTEN-Induced Putative Kinase 1),
- c) *DJ-1* (PARK7).

SNCA (A-Synuclein)

Deficient dopamine retention resulting from α -synuclein mutations may play a role in pathogenesis of PD.³⁴ Polymeropoulos *et al.*,³⁵ discovered a missense mutation in the α -synuclein gene on chromosome 4q21-23, which led to the replacement of alanine with threonine at position 53. α -Synuclein mutations follow an autosomal dominant inheritance pattern and lead to an earlier onset of PD compared to idiopathic cases 52 and 56. Mutations associated with PD accelerate α -synuclein oligomerization, and the development of non-fibrillar oligomers may be crucial for disease progression.³⁶ The presence of α -synuclein as the main element in Parkinson's disease.³⁷ Parkin ubiquitinates a higher-molecular-weight variant of α -synuclein, a role that is lost because of disease-related mutations in parkin.³⁸ A later study revealed that parkin targets synphilin-1, a protein that interacts with α -synuclein, rather than α -synuclein itself.³⁹ The ubiquitination of synphilin-1 is impaired by familial parkin mutations. The toxic buildup of abnormal α -synuclein forms can occur, as oxidative stress has been shown to trigger α -synuclein aggregation.^{40,41}

This implies that inadequate removal of oxidatively modified α -synuclein may contribute to the development of Parkinson's.

DRD2 Gene

This gene does not follow a clear autosomal dominant or recessive inheritance pattern, like *SNCA*, *LRRK2*, *PINK1*, or *PARK2*. This gene is mainly regarded as a risk factor for Parkinson's disease rather than a direct cause. *DRD2* comprises six introns. Alternative splicing gives rise to two primary variants: *D2S* and *D2L*.⁴² Over 200 polymorphisms have been discovered in *DRD2*, primarily located in introns and regions flanking downstream.⁴³ Disorders of mood, schizophrenia, movement-related conditions, and variations in drug response.^{44,45} A common single nucleotide polymorphism which involves a C-to-T substitution at position 957 in exon 7. The minor T allele is present in approximately 50% of the Caucasian population. This SNP has been linked to reduced mRNA stability and translation, diminished dopamine-induced upregulation of *DRD2* membrane expression *in vitro*, and reduced *DRD2* expression in the cortex and striatum of healthy individuals.⁴⁶⁻⁴⁸ The syndrome has been associated with the impairment of Dopamine (DA) receptors, which exhibit numerous mutations. To clarify the role of mesolimbic DA systems, three primary competing explanations "liking," "learning," and "wanting" were assessed in patients having symptoms of PD.⁴⁹ Certain genetic factors and environmental influences.⁵⁰ may lead to a deficiency in D2 receptors, increasing individuals' susceptibility to engaging in various addictive, impulsive, and compulsive behaviors.^{51,52} *DAT1* is located on chromosome 5p15.3, the *DAT1* gene (also known as *SLC6A3*) contains a *VNTR* polymorphism in its 3' non-coding region.^{53,54} Two significant alleles within this polymorphism may independently increase the risk of developing Reward Deficiency Syndrome (RDS) behaviours.

LRRK2 Gene

The *LRRK2* (Leucine-Rich Repeat Kinase 2) gene mutation is a major genetic factor in Parkinson's Disease (PD). It produces a kinase enzyme that plays a crucial role in neuronal signalling, autophagy, and mitochondrial processes. Variants of *LRRK2*, particularly the *G2019S* mutation, have been linked to both hereditary and sporadic forms of PD. The expression of *LRRK2* results in an age-related decline in dopamine (DA)-responsive motor activity and the degeneration of DA neurons.⁵⁵⁻⁵⁹ *LRRK2* is functional in the brain and comprises 51 exons. Dardarin contains five functional domains in its C-terminal region.^{60,61} A portion of this protein is known as the leucine-rich region because of its high concentration of the amino acid leucine, which serves as a building block of proteins. To assess the impact of *LRRK2* on tau phosphorylation, a SH-SY5Y cell clone was used that overexpressed *LRRK2*. The results showed a notable increase in tau phosphorylation at *Thr181* and *Ser396* in the cells with elevated *LRRK2* expression. This increase was mitigated when *LRRK2* was knocked down.⁶² Oxidative stress is a condition in

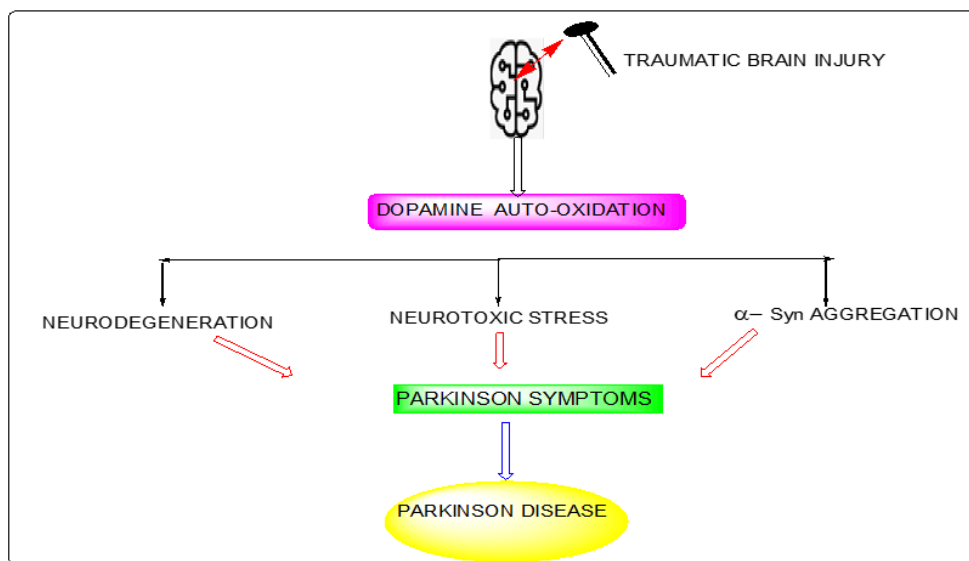


Figure 1: Neurodegeneration pathway caused due to TBI leading to PD.

which the body's antioxidant defenses fail to maintain normal levels of Reactive Oxygen Species (ROS) at normal levels.⁶³ Under such conditions, cells overexpressing either *LRRK2 WT* or the *p.G2019S* variant experience significantly increased cell death rates compared to those not under oxidative stress. This indicates that the expression of *LRRK2 WT* or *p.G2019S*, combined with oxidative stress, synergistically enhanced cell death, with the most pronounced effect observed in cells overexpressing the *p.G2019S* variant.⁶⁴ *LRRK2* may play a role in the progression of Parkinson's disease by directly impairing immune cell function. Where it is involved in immune responses to pathogens.^{65,66}

A significant increase in *LRRK2* protein levels has been detected in microglial cells within the Substantia Nigra Pars Compacta (SNpc) and striatum of mice following inflammation induced by Lipopolysaccharide (LPS).⁶⁷

PINK1 Gene

Alteration in *PINK1* Gene Cause Parkinson's disease (PD), resulting from either homozygous or compound heterozygous alterations. However, some studies have identified heterozygous *PINK1* mutations in patients with PD and in healthy controls. The presence of heterozygous mutations in healthy individuals suggests that other genetic or environmental factors may influence penetrance and expression of the disease. Additionally, the functional impact of these mutations can vary, with some leading to partial loss of kinase activity or mitochondrial dysfunction, which are central to *PINK1*'s role in mitochondrial quality control and protection against oxidative stress.⁶⁸⁻⁷⁰ The mutation results in the replacement of the amino acids arginine and glycine at a particular protein site, which contributes to the onset of parkinsonism in the population. This mutation was initially discovered in 2001 in a large Italian family pedigree located on chromosome 1 (*PARK 6* locus).^{71,72} It is hypothesized that a

mutation in *PINK1* may increase vulnerability to reactive oxygen species and other cellular stressors, ultimately contributing to the development of Parkinson's disease.⁷³

ATP13A2 Gene

ATP13A2, also known as *PARK9*, is a P-type ATPase that functions primarily as an ion pump. It is involved in the transport of polyamines such as spermidine and spermine, which are crucial for cellular homeostasis. *ATP13A2* is essential for maintaining lysosomal function and protecting against oxidative stress and neurodegeneration.⁷⁴ When *ATP13A2* function is compromised, lysosomal degradation is impaired, leading to accumulation of α -synuclein aggregates. This contributes to PD pathogenesis by promoting neuronal toxicity and degeneration. Additionally, *ATP13A2* has been shown to regulate autophagy, which is crucial for clearing damaged proteins and organelles.⁷⁵ *ATPase13A2*, which consists of 29 exons, encodes a protein composed of 1,180 amino acids. *ATP13A2* is typically located in the lysosomal membrane and features an ATPase domain along with ten transmembrane domains. This protein plays a role in preventing α -synuclein clumping, maintaining proper mitochondrial and lysosomal activities, and protecting against neurodegeneration. Mutations in *ATP13A2* have been identified in patients with juvenile-onset Parkinson's disease.⁷⁶

DJ-1 Gene

DJ-1 serves a vital function as a sensor for oxidative stress and a defender Against Reactive Oxygen Species (ROS). It engages in numerous cellular activities, including antioxidant defense, mitochondrial operations, and protein breakdown. Alteration in the *DJ-1* gene are associated with juvenile Parkinson's disease, which is inherited in an autosomal recessive pattern, highlighting its crucial role in promoting neuronal survival and protecting against oxidative damage.⁷⁷ *DJ-1*, also referred to as protein *DJ-1*,

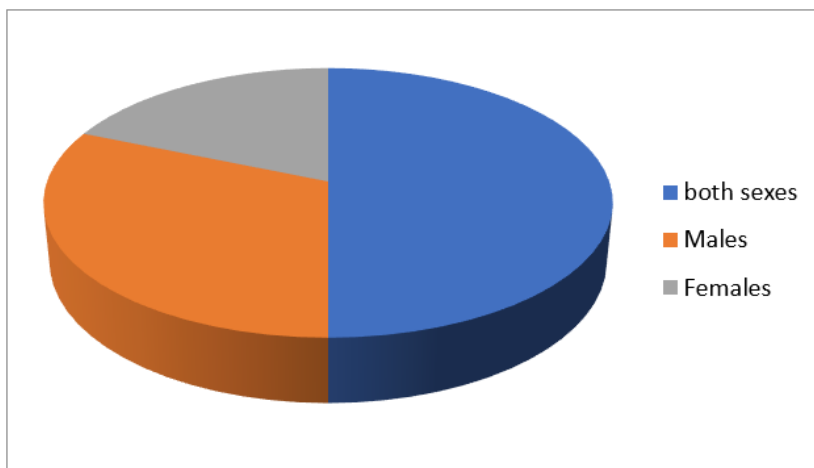


Figure 2: TBI cases reported in India.

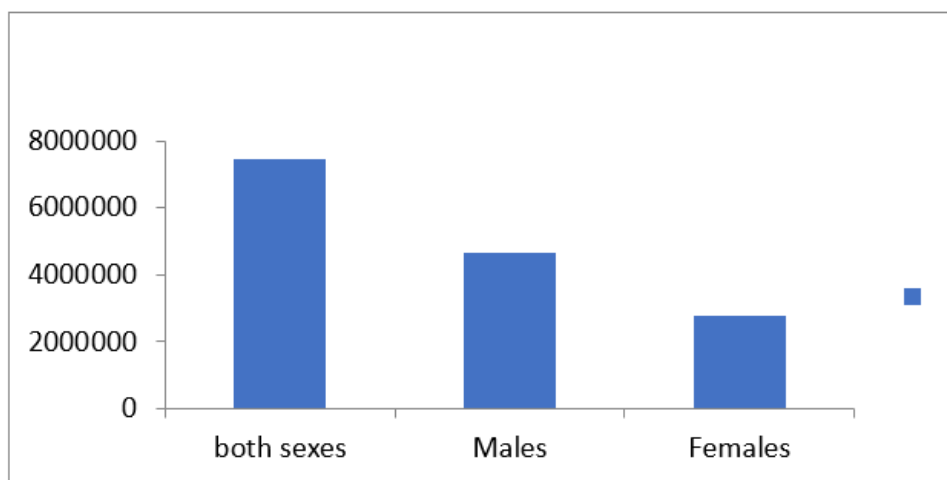


Figure 3: PD cases reported in India.

is composed of seven coding exons that generate a 189-amino acid protein referred to as Parkinson's protein 7. This protein is widely expressed and acts as a cellular sensor for oxidative stress.^{78,79} *DJ-1* functions as a chaperone in the protein-folding process, assisting newly formed proteins in attaining the correct three-dimensional shape, refolding damaged proteins, conveying specific proteins to proteasomes, and aiding RNA production and regulation. *DJ-1* gene mutations account for approximately 1-2% of autosomal recessive early onset Parkinson's disease cases.⁸⁰ The identification of *DJ-1* mutations in consanguineous families from secluded European areas is crucial for understanding the genetic foundation of Parkinson's. *DJ-1*, located on chromosome 1p36 (*PARK7 locus*).⁸¹ The *p.Arg610Gly* mutation is collocated in the GYF domain of the protein it encodes and is considered pathogenic because it disrupts ligand-binding function. This mutation leads to abnormalities in *GIGYF2*, resulting in insulin dysregulation and impaired signalling in the Insulin/IGF-1 Receptor (IGF-1R) pathway, which serves as an equilibrium maintenance of brain function. This disruption is regarded as a key mechanism underlying LOPD.^{82,83}

CONCLUSION

PD is characterized by the depletion of dopamine neurotransmitters and receptors. The evidence reviewed here suggests that traumatic brain injury, cocaine abuse, and genetic Alteration may contribute to the development and progression of Parkinson's disease by exacerbating dopamine depletion and receptor dysfunction. An increasing number of cases of injuries and genetic or abuse-related neurological disorders have been reported in India. Substantial state-level variations. These neurological disorders, especially Parkinson's disease, emphasize the importance of tailored, state-specific healthcare system responses. These gaps in neurology services-spanning awareness, early detection, treatment, and rehabilitation-should be addressed to improve the prevention and management of PD.

ACKNOWLEDGEMENT

I am grateful to be a part of Faculty of Pharmacy, Department of Pharmacology, Sri Adichuchanagiri college of Pharmacy, Adichuchanagiri University, BG. Nagara.

ABBREVIATIONS

PD: Parkinson's Disease; **LOPD:** Late-Onset Parkinson's Disease; **PARK7:** Parkinson's Disease Protein 7; **ROS:** Reactive Oxygen Species; **LPS:** Lipopolysaccharide; **LRRK2:** Leucine-Rich Repeat Kinase 2; **DA:** Dopamine; **SNP:** Single Nucleotide Polymorphism; **PINK1:** PTEN-Induced Putative Kinase 1; **MAO:** Monoamine Oxidase; **COMT:** Catechol-O-Methyltransferase; **FPI:** Fluid Percussion Impact; **BBB:** Blood-Brain Barrier; **HVA:** Homovanillic Acid; **MHPG:** 3-Methoxy-4-Hydroxyphenylglycol; **HDRS:** Hamilton Depression Rating Scale; **MAPT:** Microtubule-Associated Protein Tau; **SNpc:** Substantia Nigra Pars Compacta.

CONFLICT OF INTEREST

The authors declare that there is no conflict of interest.

SUMMARY

Parkinson's Disease (PD) is a progressive neurodegenerative disorder characterized by the loss of dopaminergic neurons in the substantia nigra and the presence of Lewy bodies. While aging is the strongest risk factor, growing evidence suggests that environmental, lifestyle, and genetic factors interact to influence disease onset and progression. This review examines the interplay between three critical factors-Traumatic Brain Injury (TBI), cocaine abuse, and genetic mutations-and their combined contribution to the development of PD.

TBI induces oxidative stress, neuroinflammation, mitochondrial dysfunction, and blood-brain barrier disruption, all of which can accelerate dopaminergic neuronal loss. Cocaine abuse, through repeated dopaminergic overstimulation, vascular injury, and neurotoxic metabolites, may further sensitize nigrostriatal pathways to degeneration. Meanwhile, pathogenic mutations in genes such as *LRRK2*, *SNCA*, *PINK1*, *DJ-1*, and *Parkin* increase vulnerability to cellular stress and protein aggregation.

When combined, these insults may act synergistically: TBI and cocaine exposure may unmask or amplify the effects of underlying genetic susceptibility, leading to earlier onset or faster progression of PD. Understanding this multifactorial interaction highlights the need for integrated research and preventive strategies, including targeted screening of at-risk individuals, neuroprotective interventions after brain injury, and harm reduction approaches for substance abuse.

REFERENCES

- GBD 2016 Neurology Collaborators. Global, regional, and national burden of neurological disorders, 1990-2016: a systematic analysis for the Global Burden of Disease Study 2016. *Lancet Neurol* 2019;18:459-80.
- Bower JH, Maraganore DM, McDonnell SDK, Rocca WA. Incidence and distribution of Parkinsonism in Olmsted County, Minnesota, 1976-1990. *Neurology* 1999;52:1214-20.
- De Lau LM, Breteler MM. Epidemiology of Parkinson's disease. *The Lancet Neurology*. 2006; 5(6): 525-35.

- Scorza FA, Guimarães-Marques M, Nejm M, *et al.* Sudden unexpected death in Parkinson's disease: insights from clinical practice. *Clinics (Sao Paulo, Brazil)* 2022;77:100001.
- Kouli A, Torsney KM, Kuan WL. Parkinson's disease: etiology, neuropathology, and pathogenesis. *Exon Publications*. 2018:3-26.
- Graybiel AM. The basal ganglia and the initiation of movement. *Revue neurologique*. 1990;146(10): 570-4.
- Jackson DM, Westlind-Danielsson A. Dopamine receptors: molecular biology, biochemistry and behavioural aspects. *Pharmacology and Therapeutics*. 1994;64(2):291-370.
- Van Bregt DR, Thomas TC, Hinzman JM, Cao T, Liu M, Bing G, *et al.* Substantial nigra vulnerability after a single moderate diffuse brain injury in rats. *Exp Neurol* 2012;234:8-19.
- Kochanek PM. Ischemic and traumatic brain injury: pathobiology and cellular mechanisms. *Critical care medicine*. 1993; 21(9):S333-4.
- Park C, Cho IH, Kim D, Jo EK, Choi SY, Oh SB, *et al.* Toll-like receptor 2 contributes to glial cell activation and heme oxygenase-1 expression in traumatic brain injury. *Neuroscience letters*. 2008;431(2):123-8.
- Xiong Y, Mahmood A, Chopp M. Animal models of traumatic brain injury. *Nat Rev Neurosci*. 2013;14(2):128-42.
- Huang EY, Tsai TH, Kuo TT, Tsai JJ, Tsui PF, Chou YC, *et al.* Remote effects on the striatal dopamine system after fluid percussion injury. *Behavioral brain research*. 2014;267:156-72.
- Xiong Y, Mahmood A, Chopp M. Animal models of traumatic brain injury. *Nature Reviews Neuroscience*. 2013;14(2):128-42.
- Lighthall JW. Controlled cortical impact: a new experimental brain injury model. *J Neurotrauma*. 1988;5(1):1-15.
- Wong VS, Langley B. Epigenetic changes following traumatic brain injury and their implications for outcome, recovery and therapy. *Neuroscience letters*. 2016;625:26-33.
- Walker KR, Tesco G. Molecular mechanisms of cognitive dysfunction following traumatic brain injury. *Frontiers in Aging Neuroscience*. 2013;5:29.
- Bales JW, Kline AE, Wagner AK, Dixon CE. Targeting dopamine in acute traumatic brain injury. *Open Drug Discovery Journal*. 2010;2:119.
- Blanchard V, Chritin M, Vyas S, Savasta M, Feuerstein C, Agid Y, *et al.* Long-term induction of tyrosine hydroxylase expression: a compensatory response to partial degeneration of the dopaminergic nigrostriatal system in the rat brain. *Journal of Neurochemistry*. 1995;64(4):1669-79.
- Onn SP, Berger TW, Stricker EM, Zigmond MJ. Effects of intraventricular 6-hydroxydopamine on the dopaminergic innervation of striatum: histochemical and neurochemical analysis. *Brain research*. 1986;376(1):8-19.
- Jenkins PO, De Simoni S, Bourke NJ, Fleminger J, Scott G, Towey DJ, *et al.* Dopaminergic abnormalities following traumatic brain injury. *Brain*. 2018;141(3):797-810.
- Shaw-Smith C, Pittman AM, Willatt L, Martin H, Rickman L, Gribble S, *et al.* Microdeletion encompassing MAPT at chromosome 17q21. 3 is associated with developmental delay and learning disability. *Nature genetics*. 2006;38(9):1032-7.
- Gauthier-Kemper A, Weissmann C, Golovyashkina N, Sebö-Lemke Z, Drewes G, Gerke V, *et al.* The frontotemporal dementia mutation R406W blocks tau's interaction with the membrane in an annexin A2-dependent manner. *Journal of Cell Biology*. 2011;192(4):647-61.
- Chen J, Kanai Y, Cowan NJ, Hirokawa N. Projection domains of MAP2 and tau determine spacings between microtubules in dendrites and axons. *Nature*. 1992;360(6405):674-7.
- Brandt R, Léger J, Lee G. Interaction of tau with the neural plasma membrane mediated by tau's amino-terminal projection domain. *Journal of Cell Biology*. 1995;131(5):1327-40.
- Alonso AD, Zaidi T, Novak M, Grundke-Iqbal I, Iqbal K. Hyperphosphorylation induces self-assembly of τ into tangles of paired helical filaments/straight filaments. *Proceedings of the National Academy of Sciences*. 2001;98(12):6923-8.
- Ritz MC, Lamb RJ, Goldberg SR, Kuhar MJ. Cocaine receptors on dopamine transporters are involved in the self-administration of cocaine. *Science*. 1987;237(4819):1219-23.
- Dackis, C. A. and Gold, M. S. New concepts in cocaine addiction: the dopamine depletion hypothesis. *Neuroscience Biobehavioral Reviews*, 1985;9:469-477.
- Gill K, Gillespie HK, Hollister LE, Davis CM, Peabody CA. Dopamine depletion hypothesis of cocaine dependence: A test. *Human Psychopharmacology: Clinical and Experimental*. 1991;6(1):25-9.
- Volkow ND, Fowler JS, Wang GJ. Imaging studies on the role of dopamine in cocaine reinforcement and addiction in humans. *Journal of psychopharmacology*. 1999;13(4):337-45.
- Di Giulio AM, Groppetti A, Cattabeni F, Galli CL, Maggi A, Algeri S, *et al.* Significance of dopamine metabolites in the evaluation of drugs acting on dopaminergic neurones. *European Journal of Pharmacology*. 1978;52(2):201-7.
- xTaylor D, Ho BT. Neurochemical effects of cocaine following acute and repeated injection. *Journal of Neuroscience Research*. 1977;3(2):95-101.
- Mateo Y, Lack CM, Morgan D, Roberts D, Jones SR. Reduced dopamine terminal function and insensitivity to cocaine following cocaine binge self-administration and deprivation. *Neuropsychopharmacology*. 2005;30(8):1455-63.

33. Willuhn I, Burgeno LM, Groblewski PA, Phillips PE. Excessive cocaine use results from decreased phasic dopamine signaling in the striatum. *Nature neuroscience*. 2014;17(5):704-9.
34. Lotharius J, Brundin P. Impaired dopamine storage resulting from α -synuclein mutations may contribute to the pathogenesis of Parkinson's. *Human Molecular Genetics*. 2002;11(20):2395-407.
35. Polymeropoulos MH, Lavedan C, Leroy E, Ide SE, Dehejia A, Dutra A, *et al.* Mutations in α -synuclein identified in families with Parkinson's disease. *science*. 1997;276(5321):2045-7.
36. Conway KA, Lee SJ, Rochet JC, Ding TT, Harper JD, Williamson RE, *et al.* Accelerated oligomerization by Parkinson's disease-linked α -synuclein mutants. *Annals of the New York Academy of Sciences*. 2000;920(1):42-5.
37. Spillantini MG, Schmidt ML, Lee VM, Trojanowski JQ, Jakes R, Goedert M. α -Synuclein in Lewy bodies. *Nature*. 1997;388(6645):839-40.
38. Shimura H, Schlossmacher MG, Hattori N, Froesch MP, Trockenbacher A, Schneider R, *et al.* Ubiquitination of a new form of α -synuclein by parkin from the human brain: Implications for Parkinson's disease. *Science*. 2001;293(5528):263-9.
39. Chung KK, Zhang Y, Lim KL, Tanaka Y, Huang H, Gao J, *et al.* Parkin ubiquitinates the α -synuclein-interacting protein, synphilin-1, which has implications for Lewy body formation in Parkinson's disease. *Nature medicine*. 2001;7(10):1144-50.
40. Hashimoto M, Hsu LJ, Xia Y, Takeda A, Sisk A, Sundsmo M, *et al.* Oxidative stress induces amyloid-like aggregate formation of NACP/ α -synuclein *in vitro*. *Neuroreport*. 1999;10(4):717-21.
41. Paxinou E, Chen Q, Weisse M, Giasson BI, Norris EH, Rueter SM, *et al.* Induction of α -synuclein aggregation by intracellular nitrate insult. *Journal of Neuroscience*. 2001;21(20):8053-61.
42. Gingrich JA, Caron MG. Recent advances in the molecular biology of dopamine receptors. *Annual review of neuroscience*. 1993;16(1):299-321.
43. Grandy DK, Litt M, Allen L, Bunzow JR, Marchionni M, Makam H, *et al.* The human dopamine D2 receptor gene is located on chromosome 11 at q22-q23 and identifies TaqI RFLP. *American Journal of Human Genetics*. 1989;45(5):778.
44. Noble EP. D2 dopamine receptor gene in psychiatric and neurological disorders and their phenotypes. *American Journal of Medical Genetics Part B: Neuropsychiatric Genetics*. 2003;116(1):103-25.
45. Zhang JP, Lencz T, Malhotra AK. D2 receptor genetic variation and clinical response to antipsychotic drug treatment: a meta-analysis. *American Journal of Psychiatry*. 2010;167(7):763-72.
46. Hirvonen MM, Lumme V, Hirvonen J, Pesonen U, Nägren K, Vahlberg T, *et al.* The C957T polymorphism of the human dopamine D2 receptor gene predicts extrastriatal dopamine receptor availability *in vivo*. *Progress in neuropsychopharmacology and biological psychiatry*. 2009;33(4):630-6.
47. Hirvonen MM, Laakso A, Nägren K, Rinne JO, Pohjalainen T, Hietala J. The C957T polymorphism of the dopamine D2 receptor gene affects striatal DRD2 availability *in vivo* by changing receptor affinity. *Synapse*. 2009;63(10):907-12.
48. Hirvonen M, Laakso A, Nägren K, Rinne JO, Pohjalainen T, Hietala J. C957T polymorphism of the dopamine D2 receptor (DRD2) gene affects striatal DRD2 availability *in vivo*. *Molecular psychiatry*. 2004;9(12):1060-1.
49. Berridge KC. The debate over dopamine's role in reward: The case of incentive salience. *Psychopharmacology*. 2007;191:391-431.
50. Gold MS, Graham NA, Cocores JA, Nixon SJ. Food addiction?. *Journal of Addiction Medicine*. 2009;3(1):42-5.
51. Rowe DC. Genetic and environmental components of antisocial behavior: A study of 265 twin pairs. *Criminology*. 1986;24(3):513-32.
52. Hällbus M, Magnusson T, Magnusson O. Influence of 5-HT1B/1D receptors on dopamine release in the guinea pig nucleus accumbens: a microdialysis study. *Neuroscience letters*. 1997;225(1):57-60.
53. Vandenbergh DJ. [34] Molecular cloning of neurotransmitter transporter genes beyond the coding region of cDNA. In *Methods Enzymol* 1998;296:498-514. Academic Press.
54. Michelhaugh SK, Fiskerstrand C, Lovejoy E, Bannon MJ, Quinn JP. The dopamine transporter gene (SLC6A3) variable number of tandem repeats domain enhances transcription in dopamine neurons. *Journal of neurochemistry*. 2001;79(5):1033-8.
55. Liu Z, Wang X, Yu Yi, Li X, Wang T, Jiang H, *et al.* Drosophila model of LRRK2-linked parkinsonism. *Proceedings of the National Academy of Sciences*. 2008;105(7):2693-8.
56. Ng CH, Mok SZ, Koh C, Ouyang X, Fivaz ML, Tan EK, *et al.* Parkin protects against LRRK2 G2019S mutant-induced dopaminergic neurodegeneration in *Drosophila*. *Journal of Neuroscience*. 2009;29(36):11257-62.
57. Venderova K, Kabbach G, Abdel-Messih E, Zhang Y, Parks RJ, Imai Y, *et al.* Leucine-Rich Repeat Kinase 2 interacts with Parkin, DJ-1 and PINK-1 in a *Drosophila melanogaster* model of Parkinson's disease. *Human molecular genetics*. 2009;18(22):4390-404.
58. Saha S, Guillily MD, Ferree A, Lanceta J, Chan D, Ghosh J, *et al.* LRRK2 modulates vulnerability to mitochondrial dysfunction in *Caenorhabditis elegans*. *Journal of Neuroscience*. 2009;29(29):9210-8.
59. Wolozin B, Saha S, Guillily M, *et al.* Investigating convergent actions of genes linked to familial Parkinson's disease. *Neurodegenerative disease*. 2008;5(3-4):182-5.
60. Mata IF, Wedemeyer WJ, Farrer MJ, Taylor JP, Gallo KA. LRRK2 in Parkinson's disease: protein domains and functional insights. *Trends in Neurosciences*. 2006;29(5):286-93.
61. Nuytemans K, Theuns J, Cruts M, Van Broeckhoven C. Genetic etiology of Parkinson disease associated with mutations in the SNCA, PARK2, PINK1, PARK7, and LRRK2 genes: a mutation update. *Human mutation*. 2010;31(7):763-80.
62. Kawakami F, Shimada N, Ohta E, Kagiya G, Kawashima R, Maekawa T, *et al.* Leucine-rich repeat kinase 2 regulates tau phosphorylation through direct activation of glycogen synthase kinase-3 β . *The FEBS journal*. 2014;281(1):3-13.
63. Shulman JM, De Jager PL, Feany MB. Parkinson's disease: genetics and pathogenesis. *Annual Review of Pathology: Mechanisms of Disease*. 2011;6(1):193-222.
64. Heo HY, Park JM, Kim CH, Han BS, Kim KS, Seol W. LRRK2 enhances oxidative stress-induced neurotoxicity via kinase activity. *Experimental cell research*. 2010;316(4):649-56.
65. Hakimi M, Selvanantham T, Swinton E, Padmore RF, Tong Y, Kabbach G, *et al.* Parkinson's disease-linked LRRK2 is expressed in circulating and tissue immune cells and is upregulated following recognition of microbial structures. *Journal of Neural Transmission*. 2011;118:795-808.
66. Gardet A, Benita Y, Li C, Sands BE, Ballester I, Stevens C, *et al.* LRRK2 is involved in the IFN- γ and host responses to pathogens. *Journal of Immunology*. 2010;185(9):5577-85.
67. Moehle MS, Webber PJ, Tse T, Sukar N, Standaert DG, DeSilva TM, *et al.* LRRK2 inhibition attenuates microglial inflammatory responses. *Journal of Neuroscience*. 2012;32(5):1602-11.
68. Bonifati V, Rohe CF, Breedveld GJ, Fabrizio E, De Mari M, Tassorelli C, *et al.* Early-onset parkinsonism associated with PINK1 mutations: frequency, genotypes, and phenotypes. *Neurology*. 2005;65(1):87-95.
69. Choi JM, Woo MS, Ma HI, Kang SY, Sung YH, Yong SW, *et al.* Analysis of PARK genes in a Korean cohort of early-onset Parkinson disease. *Neurogenetics*. 2008;9:263-9.
70. Weng, Y.-H.; Chou, Y.-H.W.; Wu, W.-S.; Lin, K.-J.; Chang, H.-C.; Yen, T.-C.; *et al.* PINK1 mutations in early onset Parkinsonism in Taiwanese patients 2007;254:1347-55.
71. Lohmann E, Periquet M, Bonifati V, Wood NW, De Michele G, Bonnet AM, *et al.* How much phenotypic variation can be attributed to parkin genotype?. *Annals of neurology*. 2003;54(2):176-85.
72. Lincoln SJ, Maraganore DM, Lesnick TG, Bounds R, de Andrade M, Bower JH, *et al.* Parkin variants in North American Parkinson's disease: cases and controls. *Movement disorders: Official Journal of the Movement Disorder Society*. 2003;18(11):1306-11.
73. Clark IE, Dodson MW, Jiang C, Cao JH, Huh JR, Seol JH, *et al.* *Drosophila* pink1 is required for mitochondrial function and interacts genetically with parkin. *Nature*. 2006;441(7097):1162-6.
74. Liu Y, Shoji-Kawata S, Sumpter Jr RM, Wei Y, Ginet V, Zhang L, *et al.* Autosis is a Na⁺, K⁺-ATPase-regulated form of cell death triggered by autophagy-inducing peptides, starvation, and hypoxia-ischemia. *Proceedings of the National Academy of Sciences*. 2013;110(51):20364-71.
75. Zhang F, Wu Z, Long F, Tan J, Gong N, Li X, *et al.* The roles of ATP13A2 gene mutations leading to abnormal aggregation of α -synuclein in Parkinson's disease. *Frontiers in cellular neuroscience*. 2022;16:927682.
76. Klein C, Westenberger A. Genetics of Parkinson's disease. *Cold Spring Harbor Perspective in Medicine*. 2012;2(1):a008888.
77. Taira T, Saito Y, Niki T, Iguchi-Ariga SM, Takahashi K, Ariga H. DJ-1 plays a role in antioxidative stress by preventing cell death. *EMBO reports*. 2004;5(2): 213-8.
78. Canet-Avilés RM, Wilson MA, Miller DW, Ahmad R, McLendon C, Bandyopadhyay S, *et al.* The Parkinson's disease protein DJ-1 is neuroprotective due to cysteine-sulfenic acid-driven mitochondrial localization. *Proceedings of the National Academy of Sciences*. 2004;101(24):9103-8.
79. Junn E, Taniguchi H, Jeong BS, Zhao X, Ichijo H, Mouradian MM. Interaction of DJ-1 with Daxx inhibits apoptosis signal-regulating kinase 1 activity and cell death. *Proceedings of the National Academy of Sciences*. 2005;102(27):9691-6.
80. Pankratz N, Pauciuolo MW, Elsaesser VE, Marek DK, Halter CA, Wojcieszek J, *et al.* Parkinson Study Group-PROGENI Investigators. Mutations in DJ-1 are rare in familial Parkinson disease. *Neuroscience letters*. 2006;408(3):209-13.
81. Bonifati V, Rizzu P, Van Baren MJ, Schaap O, Breedveld GJ, Krieger E, *et al.* Mutations in DJ-1 are associated with autosomal recessive early onset parkinsonism. *Science*. 2003;299(5604):256-9.
82. Ruiz-Martinez J, Krebs CE, Makarov V, Gorostidi A, Martí-Massó JF, Paisán-Ruiz C. GIGYF2 mutation in late-onset Parkinson's disease with cognitive impairment. *Journal of human genetics*. 2015;60(10):637-40.
83. Aleman A, Torres-Alemán I. Circulating insulin-like growth factor I and cognitive function: neuromodulation throughout the lifespan. *Progress in neurobiology*. 2009;89(3):256-65.

Cite this article: Qutboddin M, Ahmed SS, Abbas N, Ali M, Raghunathanaidu BD. The Interplay between Traumatic Brain Injury, Cocaine Abuse, and Genetic Mutations in the Development of Parkinson's Disease: A Review. *Indian J of Pharmaceutical Education and Research*. 2026;60(3):918-24.