Gamma Amino Butyric Acid Ameliorates Jejunal Oxidative Damage in Diabetic Rats

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ABSTRACT

Introduction: The small intestine exhibits morphological and functional changes in streptozotocin (STZ)-induced diabetes. This study was carried out to clarify the role of gamma-aminobutyric acid (GABA) in ameliorating the histopathological changes and the oxidative stress in small intestine of the diabetic rats. Material and Methods: For this purpose, 40 adult male Swiss albino rats (Sprague dawley strain) were divided into 4 groups (10 animals in each group); the control group, GABA treated group, the diabetic group and the fourth group was subjected to induction of diabetes and GABA treatment. All animals were sacrificed and specimens from jejunum were removed and processed for the histopathological examinations, cytomorphometric analysis and oxidative stress assessment. Results: In diabetic rats, Jejunal sections from the diabetic rats exhibited pleomorphism of villi, inflammatory cellular infiltration and edema in the villous core with lymphoid hyperplasia, an increase in goblet cells number was also observed with disappearance of Paneth cells or they were degranulated, increased crypt numbers and mucosal height and increased thickness of muscularis layer. Moreover, there was an increase in the oxidant biomarkers with decline in the antioxidant enzymes in Jejunal sections of the diabetic rats. Conclusion: Treatment of rats with GABA ameliorated the observed histopathological alterations, morphometric changes and improved the oxidant/antioxidant status of the jejunum in diabetic rats. Based on these results, it was concluded that GABA administration to rats ameliorated diabetes-induced jejunal histopathological and biochemical alterations.

Key words: Diabetes, Jejunum, Morphometry, Paneth Cells, Gamma-Aminobutyric Acid.

INTRODUCTION

Diabetes mellitus (DM) is a chronic, heterogeneous and multifactorial disease; it is characterized by hyperglycemia due to impaired insulin secretion and aberrant glucagon secretion resulting from changes in the pancreatic islet cell functions. Most of diabetic patients are non-insulin-dependent while small proportions have insulin-dependent diabetes, type 1 diabetes mellitus (T1DM) is a chronic disease results in a cellular-mediated autoimmune destruction of β-cells of the pancreas, insulin deficiency and hyperglycemia are the main outcomes of it, while in type 2 diabetes or adult-onset diabetes patients have insulin resistance and usually have relative insulin deficiency. Hyperglycemia is responsible for metabolic disturbances including oxidative stress which leads to injuries in the different cellular types of gastrointestinal tract and impair endogenous antioxidant defense system. Oxidative stress due to auto-oxidation of glucose and decreased antioxidant defense is believed to play a role in the onset of DM and its complications. The small intestine is unique in its variety of cell types including enterocytes (small
intestinal epithelial cells), goblet cells (for mucus secretion), entero-endocrine cells (for hormone secretion) and Paneth cells which are stem cells differentiate to form enterocytes. Paneth cells located at the base of the Lieberkühn glands and they synthesize substances with antimicrobial properties such as α-defensins, lysozyme and phospholipase A, in response to different stimuli. Paneth cells also control the composition of endogenous flora and play a role in the crypt formation and development of phagocytosis, digestion and detoxification processes.

Gamma-aminobutyric acid (GABA) is one of the main neurotransmitters in the central nervous system. It is produced from glutamate by decarboxylation via glutamic acid decarboxylase (GAD) activity and it creates biological signaling through activation of A and B types of GABA receptors. GABA and GABA-A receptors are found in non-neuronal cells of visceral organs. It is known that GABA is a transmitter of enteric interneurons and it regulates the gastrointestinal tract functions. GABA is also found in the epithelial cells of rat jejunum and it has immune inhibitory effects that may protect β-cells of pancreas from autoimmune destruction.

**MATERIAL AND METHODS**

**Streptozotocin treatment**

Streptozotocin (STZ) purchased from Sigma-Aldrich, St. Louis, Missouri, USA, within the type of one gram vials. Type 1 diabetes mellitus was induced by single intraperitoneal injection of rats with freshly prepared STZ at a dose of 60 mg/kg dissolved in 0.1M cold citrate buffer pH 4.5. Since STZ is capable of producing fatal hypoglycemia as a result of massive pancreatic release of insulin, the rats were given 5% glucose for the next 24 hr to prevent hypoglycemia. Blood glucose levels were determined using an Accu-check blood glucose meter (Roche Diagnostics, Basel, Switzerland). Three days following STZ injection the blood was withdrawn from the tail vein and glucose level was monitored. Rats with fasting blood glucose levels ≥ 250 mg/dl were considered as diabetic and included in the current study.

**Treatment of rats with Gamma amino butyric acid**

Gamma amino butyric acid (GABA) purchased from Sigma-Aldrich, St Louis, Missouri, USA in the form of 25 g vials was dissolved in distilled water and administered to rats daily by gastric gavages at doses of 200 mg/Kg body weight/day for successive 3 weeks.

**Animals**

The present study was conducted on 40 adult male Swiss albino rats (Sprague Dawley strain) (10 ± 2 weeks old; weighting 120 ± 10 g at the beginning of the experiment) purchased from the Egyptian company for Biological merchandise and Vaccines (Helwan, Cairo, EGYPT). They were kept for about 15 days, before the onset of the experiment under observation to be acclimatized to the laboratory conditions. Animals were housed collectively in plastic cages, maintained under standard conditions of light, ventilation, temperature and humidity and allowed free access of standard pellet diet and tap water. For biochemical analyses animals were sacrificed at 11:00 am ± 1 h. All animal procedures were carried out in accordance with the Ethics Committee of the National Research Centre followed to the “Guide for the Care and Use of Laboratory Animals” published by the US National Institutes of Health.

**Animal groups**

Experimental animals were randomly divided into 4 groups (n=10) treated in parallel and classified as following:

1. Control (C) group: Normal healthy rats administered 1 ml distilled water daily during 3 weeks via gastric gavages.
2. GABA (G) group: Normal healthy rats administered GABA (200 mg/Kg body weight/day) daily during 3 weeks via gastric gavages.
3. Diabetic (DM) group: STZ-treated rats administered 1 ml distilled water daily during 3 weeks via gastric gavages.
4. DM+GABA group: STZ-treated rats administered GABA (200 mg/Kg body weight/day) daily during 3 weeks via gastric gavages.
5. At the end of the third week rats were sacrificed after a fasting period of 12 hours next day to the last dose of GABA.

**Biochemical analysis**

Rats were anaesthetized with light ether, sacrificed and underwent laparotomy immediately. One cm segment of jejunum was dissected and washed with saline solution for the histological examinations. Other small segments of jejunum (10% w/v) were rapidly excised, homogenized in saline by using Teflon homogenizer (Glass-Col, Terre Haute, Ind., USA) and after centrifugation at 10 kg for fifteen minutes using refrigerated centrifuge (manufactured by K3 Centurion Scientific, Ltd, London, UK) the supernatant was used for the estimation of markers of oxidative stress. Chemicals and reagents...
were purchased from Sigma-Aldrich, St Louis, MO, USA. Measurement of absorbance was accomplished by utilizing a T60 UV/VIS spectrophotometer, PG instruments, London, UK.

Assessment of oxidative stress in jejunal section
The extent of lipid peroxidation was assayed as described by Yoshioka et al.,21 advanced oxidation protein products (AOPPs) was determined according to the method of Witko-Sarsat et al.,22 catalase activity was determined as described by Sinha23 and glutathione peroxidase (GSH-Px) activity was determined according to the method described by Necheles et al.24

Histopathologic examination
Tissue samples were fixed in 10% neutral buffered formalin and Carnoy’s fixative and embedded in paraffin. The sections were cut at 5 µm and stained with hematoxylin and eosin (H&E) and Masson’s trichrome stains for detection of the histological changes,25 periodic acid Schiff’s (PAS) method26 and Feulgen’s reaction.27 Sections were examined for mucosal injury, inflammation, congestion and graded in a blinded manner by a pathologist. Sections stained with PAS were evaluated for goblet cell prevalence and Masson’s trichrome stained sections were used to detect Paneth cells.

Morphometric and Cytometric analysis
For morphometric analysis, four whole circumference sections were examined. Also the number of villi and crypts per circumference, mucosal thickness, muscularis layer thickness and the outer intestinal boundary were estimated. Ten well orientated villi per segment were randomly selected and measured for villus height. Nuclear area, proliferation index and ploidy values of 100-150 cells within random fields were estimated in Feulgen’s stained sections. All morphometric and cytometric parameters were done in the Pathology Department, National Research Centre employing the Leica Qwin 500 Image Analyzer (LEICA Imaging Systems Ltd, Cambridge, England,) that consists of Leica DM-LB microscope provided with JVC color video camera attached to a computer system Leica Q 500IW. The results were recorded as the mean and standard deviation values.

Statistical analysis
The SPSS/PC computer program (version 20.0; SPSS Inc., Chicago, IL, USA) was used for statistical analysis of the results. Data were analyzed using one-way analysis of variance (ANOVA) followed by Post Hoc LSD test. The data which obtained were expressed as mean ± standard deviation (SD). Differences were thought of statistically significant once P value less than 0.05.

RESULTS

Biochemical measurements

Changes of oxidant and anti-oxidant biomarkers
The results obtained in the present study showed that the administration of gamma amino butyric acid (GABA) (200 mg/Kg/day) to normal rats (GABA group) for 3 weeks showed insignificant changes in catalase (CAT), glutathione peroxidase (GSH-Px) activities, malondialdehyde (MDA) and advanced oxidation protein products (AOPPs) levels and they were within the normal values in the small intestine tissues Tables 1 and 2. The administration of STZ to rats (DM group) showed a significant increase in AOPPs and MDA values and a significant decrease in CAT and GSH-Px values Tables 1 and 2. Administration of GABA to STZ-treated rats (DM+GABA group) improved the status of the above mentioned parameters Tables 2 and 3 which was evidenced by a significant increase in the antioxidants combined with a significant decrease in the oxidant biomarkers levels in small intestine tissues compared to those of the diabetic group.

Histopathologic observations

Control and GABA treated groups
Light microscopic examination of the control group H&E-stained sections showed that the villi are covered with tall columnar cells (enterocytes) with oval basal vesicular nuclei and connective tissue core. Crypts of Lieberkühn are invaginations of mucosa between the bases of the villi. Goblet cells were seen in the PAS stained sections, they were present in between the enterocytes at intervals. Paneth cells were found at the base of the crypts as pyramidal shaped cells with prominent eosinophilic granules in Masson’s trichrome stained sections Figure 1 A-C. The intestinal specimens obtained from the GABA treated group showed normal architecture of the villi and crypts Figure 1 D-F.

Diabetic group
Examined sections of this group showed villous and crypt hypertrophy, disturbed villous architecture was seen with some thick and other conical villi; other villi showed loss of epithelial coverings. Many of these villi appeared twisted or collapsed and others showed separation of the epithelium from the underlying connective tissue core. Inflammatory cellular infiltration and edema of the villous core were seen with lymphoid hyperplasia.
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<th>H&amp;E</th>
<th>PAS</th>
<th>Masson trichrom</th>
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Figure 1 – A-C Photomicrographs of jejunum sections from control rat showed that the villi are covered with tall columnar cells (enterocytes) with oval basal vesicular nuclei and connective tissue core. Crypts of Lieberkühn are invaginations of mucosa between the bases of the villi. Goblet cells were seen in the PAS stained sections, they were present in between the enterocytes at intervals. Paneth cells were found at the base of the crypts as pyramidal shaped cells with prominent eosinophilic granules in Masson’s trichrome stained sections. (D-F) Photomicrographs of jejunum sections from GABA treated rat showed normal architecture of the villi and crypts. (G-I) Photomicrographs of jejunum sections from diabetic rat showed villous and crypt hypertrophy, disturbed villous architecture was seen with some thick and other conical villi; other villi showed loss of epithelial coverings. Many of these villi appeared twisted or collapsed and others showed separation of the epithelium from the underlying connective tissue core. Inflammatory cellular infiltration and edema of the villous core were seen with lymphoid hyperplasia. Goblet cells increased in number and Paneth cells disappeared or appeared degranulated. (J-L) Photomicrographs of jejunum sections from GABA treated diabetic rat slight villi hypertrophic changes with moderate lymphocytic infiltration. Goblet cells appeared normal and Paneth cells were of normal count and shape. (H&E, x400)
Goblet cells increased in number and Paneth cells disappeared or appeared degranulated Figure 1 G-I.

**GABA treated diabetic group**

Sections of diabetic and GABA treated rats showed slight villi hypertrophic changes with moderate lymphocytic infiltration. Goblet cells appeared normal and Paneth cells were of normal count and shape Figure 1 J-L.

**Morphometric and Cytometric analysis**

Measured morphometric parameters of the examined sections showed a significant increase in the number of villi and crypts in the diabetic group. The mucosal height, the muscle layer thickness and the outer intestinal circumference increased significantly in the diabetic model compared to both the control and GABA groups demonstrating the effects of diabetes on the intestinal mucosa. The number of villi and crypts approximate somewhat near to the control value in the GABA treated diabetic group. Also, the mucosal height, the muscle layer thickness and the outer intestinal circumference were significantly lower than those of the diabetic group Table 3. Cytometric analysis of the diabetic group sections showed that the majority of epithelial cell nuclei have diploid DNA values with medium proliferation index and minor population of tetraploid cells comparable to the control and GABA treated groups.

**DISCUSSION**

Diabetes is one of the leading causes of death worldwide. It is a group of metabolic diseases characterized by hyperglycemia. Diabetes results from disorders in insulin production and/or insulin action and impaired operate within the metabolism of carbohydrates, lipids and proteins which lead to long term health complications. Type 1 diabetes mellitus is an autoimmune disease characterized by pancreatic islets infiltration by T lymphocytes, macrophages and other immune cells and loss of β-cells.

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**Table 1: Influence of GABA administration to different animal groups on the oxidant biomarkers values in the small intestine tissues of STZ-treated rats**

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Groups</th>
<th>AOPPs (µmol/L)</th>
<th>MDA (nmol/g)</th>
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<tr>
<td></td>
<td>Control</td>
<td>164.90±6.59</td>
<td>112.50±7.65</td>
</tr>
<tr>
<td></td>
<td>GABA</td>
<td>158±5.19</td>
<td>103.80±11.63</td>
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<td></td>
<td>P1</td>
<td>NS</td>
<td>&lt;0.05</td>
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<tr>
<td></td>
<td>DM</td>
<td>176.80±8.23</td>
<td>131.50±4.99</td>
</tr>
<tr>
<td></td>
<td>P1</td>
<td>7%</td>
<td>17%</td>
</tr>
<tr>
<td></td>
<td>DM+GABA</td>
<td>135.80±10.14</td>
<td>107.30±6.80</td>
</tr>
<tr>
<td></td>
<td>P1</td>
<td>-18</td>
<td>&lt;0.001</td>
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<tr>
<td></td>
<td>P2</td>
<td>-5%</td>
<td>&lt;0.001</td>
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</table>

Values are expressed as Means ± Standard Deviation (n=10). Values between brackets show percentage of change from Control. P1: significance vs control. P2: significance vs respective group (DM) not receiving GABA. NS: non-significant. P < 0.001 highly significant

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**Table 2: Influence of GABA administration to different animal groups on the antioxidants biomarkers values in the small intestine tissues of STZ-treated rats**

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Groups</th>
<th>GSH-Px (GSH /min/g fresh tissue)</th>
<th>Catalase (µmol of H₂O₂ consumed/min/mg protein)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Control</td>
<td>16.62±0.37</td>
<td>13.37±0.74</td>
</tr>
<tr>
<td></td>
<td>GABA</td>
<td>16.50±0.46</td>
<td>13.66±1.38</td>
</tr>
<tr>
<td></td>
<td>P1</td>
<td>NS</td>
<td>NS</td>
</tr>
<tr>
<td></td>
<td>DM</td>
<td>13.30±0.95</td>
<td>4.36±0.43</td>
</tr>
<tr>
<td></td>
<td>P1</td>
<td>-20%</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td></td>
<td>DM+GABA</td>
<td>16.88±0.60</td>
<td>5.68±0.85</td>
</tr>
<tr>
<td></td>
<td>P1</td>
<td>2%</td>
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</tr>
<tr>
<td></td>
<td>P2</td>
<td>&lt;0.001</td>
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Values are expressed as Means ± Standard Deviation (n=10). Values between brackets show percentage of change from Control. P1: significance vs control. P2: significance vs respective group (DM) not receiving GABA. NS: non-significant. P < 0.001 highly significant
In the current study, a single intraperitoneal injection of 60 mg/kg body weight of STZ to male albino rats induced hyperglycemia, and insulin deficiency (similar to type 1 DM) that was proved by our previous study. The decrease of insulin might result from hyperglycemia-induced beta-cell toxicity and degeneration of Langerhans islet beta cells. STZ, an antibiotic produced by *Streptomyces chromogenes*, is the most commonly used agent in experimental diabetes. It was demonstrated that STZ induced damage to pancreatic cell membranes and evoked oxidative stress in islet cells. In addition, STZ has been shown to induce DNA strand breaks and DNA alkylation in pancreatic islet cells. Experimental studies demonstrated that STZ induces Type 1 DM or Type 2 DM, depending on the dose administered.

Administration of STZ to rats induced oxidative stress indicated by a significant increase in the oxidant biomarkers values (AOPP and MDA) and a significant decrease in the antioxidants (catalase and GSH-Px) activities, such observations come in coincidence with those reported by Eltahawy et al. Oxidative stress plays an important role in cellular injury following hyperglycemia where high glucose level could stimulate free radical production processes whereas weak body defense system becomes unable to counteract the enhanced reactive oxygen species (ROS) generation resulting in imbalance between ROS and their protection which finally lead to oxidative stress processes. Several mechanisms have been reported to be responsible for oxidative stress in diabetes including excess oxygen free radicals production from auto-oxidation of glucose, glycated proteins and glycation of antioxidative enzymes which inhibit their capacity to detoxify oxygen free radicals in addition to high glucose levels stimulation of cytochrome *P*₄₅₀-like activity by excessive nicotinamide adenine dinucleotide phosphate-oxidase (NADPH) resulting from glucose metabolism and ketosis, a hallmark of T1DM, which may increase oxygen free radicals production.

Results of the current experiment showed numerous histopathological changes in jejunum of STZ-induced diabetic rats illustrated by villous and crypt hypertrophy, disturbed villous architecture of some villi and loss of epithelial covering of others. Inflammatory cellular infiltration and edema of the villous core with lymphoid hyperplasia were also depicted with increased number of goblet cells whereas Paneth cells disappeared or appeared degranulated. The majority of epithelial cell nuclei of the diabetic group had diploid DNA values with medium proliferation index. Such histopathologic findings come in collaboration with those observed by Abo Gazia and Hasan.

### Table 3: Morphometric parameters in the studied groups

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Control</th>
<th>GABA</th>
<th>DM</th>
<th>DM + GABA</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>No. of villi</strong></td>
<td>50</td>
<td>45</td>
<td>65*</td>
<td>50</td>
</tr>
<tr>
<td><strong>No. of crypts</strong></td>
<td>135</td>
<td>130</td>
<td>153*</td>
<td>120</td>
</tr>
<tr>
<td><strong>Mean mucosal thickness (μm)</strong></td>
<td>56.35±16.54</td>
<td>50.22±15.23</td>
<td>100.86±4.35*</td>
<td>67.44±7.73</td>
</tr>
<tr>
<td><strong>Mean mucosal thickness (μm)</strong></td>
<td>7.72±0.76</td>
<td>7.13±0.75</td>
<td>19.53±2.13*</td>
<td>11.32±1.54</td>
</tr>
<tr>
<td><strong>Outer intestinal circumference (μm)</strong></td>
<td>1379.46</td>
<td>1334.52</td>
<td>1477.99*</td>
<td>1399.46</td>
</tr>
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</table>

Values are expressed as Means ± Standard Deviation (n=10). (*) significantly different from the control at P<0.05

### Table 4: DNA cytometry parameters in the studied groups

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Control</th>
<th>GABA</th>
<th>DM</th>
<th>DM + GABA</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Diploid value %</strong></td>
<td>75.676 %</td>
<td>80.237 %</td>
<td>83.019 %</td>
<td>93.660 %</td>
</tr>
<tr>
<td><strong>Proliferation index</strong></td>
<td>18.018 %</td>
<td>15.961 %</td>
<td>15.094 %</td>
<td>6.422 %</td>
</tr>
<tr>
<td><strong>Tetraploid value %</strong></td>
<td>6.306 %</td>
<td>4.402 %</td>
<td>1.887 %</td>
<td>0.917 %</td>
</tr>
<tr>
<td><strong>Aneuploid value %</strong></td>
<td>0.0 %</td>
<td>0.0 %</td>
<td>0.0%</td>
<td>0.0%</td>
</tr>
<tr>
<td><strong>Nuclear area(μm²)</strong></td>
<td>27.74</td>
<td>25.23</td>
<td>25.97</td>
<td>24.70</td>
</tr>
</tbody>
</table>

Values are expressed as Means ± Standard Deviation (n=10).

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Eltahawy et al.: GABA ameliorates jejunal oxidative damage.

many environmental toxins such as concomitant exposure to aluminum and ionizing radiation increased detrimental structural changes in Paneth cells.  

In the past five decades, the function of GABA in the CNS has been well documented. However, the presence of a GABAergic system within the pancreas as a potential target for treatment diabetes mellitus appeared only recently. In α-cells, GABA induces membrane hyperpolarization and inhibits glucagon secretion. In β-cells, GABA induces membrane depolarization and improves insulin secretion. GABA also has valuable effects on β-cell survival and regeneration, which results in enlarged β-cell mass. Moreover, GABA inhibits insulitis and systemic inflammatory cytokine production. Furthermore, GABA treatment increased β-cell proliferation, while decreasing apoptosis, leading to enhanced β-cell mass. In addition, the activation of GABA-A receptors in islet β-cells increases insulin release. Nevertheless, according to the results of the current work, the significant improvement in all the studied parameters could be attributed to the role of GABA in improving insulin and glucose level as shown by our previous study that attributing to GABA treatment increased β-cell proliferation, while decreasing apoptosis, leading to enhanced β-cell mass. In addition, the activation of GABA-A receptors increases insulin release. Furthermore, GABA has been shown to attenuate oxidative stress through increasing endogenous superoxide dismutase (SOD) and CAT activities, decreasing lipid peroxidation due to its free radicals scavenging activities, its effectiveness in inhibiting the formation of reactive carbonyl intermediates and to react with MDA, increased in diabetic subjects as a side effect of increased protein glycation, to form different conjugated complexes thus protect the tissue against active carbonyl damage. Treatment of the diabetic rats with GABA reduced the severity of changes in the intestinal epithelial cells where the villi appeared slightly hypertrophic with moderate lymphocytic infiltration in the connective tissue core. Both goblet and Paneth cells were of normal count and shape. The majority of epithelial cells nuclei had diploid DNA values with low proliferation index, the mean nuclear area was lower in GABA treated diabetic group as compared to the other groups. In addition, treatment with GABA showed a significant increase in the antioxidants content associated with a significant decrease in the oxidant biomarkers values in the small intestine tissues indicating that GABA ameliorated the intestinal changes in STZ-induced diabetic rats. GABA exerts pancreatic β-cell regenerative effects as it protects β-cells against apoptosis and supported anti-inflammatory and immunoregulatory activities. Moreover, it exerted anti diabetic effects by acting on both the islet of β-cells and immune system, therapy preserving β-cell and preventing the development of T1DM. Moreover, it could suppress insulitis and inflammatory cytokines production. Furthermore, GABA has been shown to diminish inflammation by inhibition of activated T lymphocytes.

CONCLUSION

Oxidative stress plays an important role in progression and development of the different types of diabetes in addition to associated complications and its complications. The current results showed that gamma aminobutyric acid (GABA) ameliorated most of the degenerative changes in STZ-induced diabetic rats’ small intestine which may be attributed to its free radicals scavenging activities, its antioxidant properties and its role in regulating insulin level.

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CONFLICT OF INTEREST

The authors report no conflict of interest.

ABBREVIATIONS USED

STZ: Streptozotocin; GABA: Gamma aminobutyric acid; DM: Diabetes mellitus; GAD: Glutamic acid decarboxylase; AOPPs: Advanced oxidation protein products; GSH-Px: Glutathione peroxidase; MDA: Malondialdehyde; CAT: catalase; PAS: Periodic acid Schiff’s; ROS: Reactive oxygen species; CNS: Central Nervous System.

REFERENCES


SUMMARY

• This study was planned to investigate the role of GABA in ameliorating the histopathological changes and the oxidative stress in small intestine of the diabetic rats.
• Rats were injected with STZ to induce Diabetes and induce oxidative damage.
• Rats were treated with GABA to ameliorate damage that occurred in small intestine of the diabetic rats.
• GABA administration to rats ameliorated diabetes-induced jejunal histopathological and biochemical alterations.

PICTORIAL ABSTRACT

About Authors

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